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MISA

wpcp

**STOPPING
WATER POLLUTION
AT ITS SOURCE**



MISA

Municipal/Industrial Strategy for Abatement

**THIRTY SEVEN
MUNICIPAL WATER POLLUTION
CONTROL PLANTS**

PILOT MONITORING STUDY

VOLUME I

INTERIM REPORT

DECEMBER 1988



**Environment
Ontario**

Jim Bradley
Minister

THIRTY SEVEN
MUNICIPAL WATER POLLUTION CONTROL PLANTS

Pilot Monitoring Study

Volume 1
Interim Report

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Ontario Ministry of the Environment
Water Resources Branch

Report prepared by:
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EXECUTIVE SUMMARY

The Ontario Ministry of the Environment's Municipal Industrial Strategy for Abatement (MISA) Program is aimed at reducing contaminant loadings from direct industrial discharges and from municipal water pollution control plants (WPCPs). The MOE will address the municipal sector by the implementation of a Monitoring Regulation, requiring all Ontario WPCPs to monitor hazardous contaminants (HCs) in effluents and sludges. Subsequently, maximum concentration requirements of HCs in effluents and sludges will be established and a Compliance Regulation will be implemented.

The MOE, Environment Canada and the Municipal Engineers Association (MEA) sponsored this study to provide the information needed to support the development of a cost-effective and practical Monitoring Regulation.

This report is an interim report, summarizing study methodologies and presenting preliminary study findings. A more detailed analysis of the study data to determine factors affecting removal of HCs, and to allow prioritization of effluents and sludges and estimation of HC Loadings from study WPCPs will be presented in the final report.

Thirty-seven Ontario WPCPs were selected for the Pilot Monitoring Study, including 28 secondary treatment plants, 7 primary treatment plants and 2 lagoons. The field monitoring program involved sampling of influent, final effluent and raw and treated sludges for one to two 5 consecutive day periods at each of the study plants. In addition, plant performance parameters were monitored for the 2 weeks prior to sampling and during the sampling period.

Each sample was analyzed for all of the contaminants on a list established by MOE for this study. The monitoring list included 144 organic contaminants, 15 metals and conventional contaminants. Three laboratories contracted by the MOE, and the MOE Laboratory Services Branch (LSB), performed all of the analytical work.

A thorough field QA/QC program involved appropriate cleanliness and sample preservation procedures, duplicate sample collection of all field samples and field blank collection and analyses. Also, a comprehensive laboratory QA/QC program was carried out so that the applicability of each analytical result could be defined. This program involved analysis of method blanks, duplicate samples, field samples spiked with surrogate compounds and distilled water samples spiked with native compounds.

The individual plant data, including background information and analytical results from the sampling program were compiled and are presented in Appendix A (Volume II) of this report.

The analytical results from the sampling program were summarized for all of the plants for each type of sample. Metals were the most prevalent (most WPCPs) and most frequently detected contaminants in all sample types. Only 5 base neutral and acid extractable compounds were ever detected at more than 20 percent of plants for any sample type. Dioxin/furan compounds were detected at a maximum of 27 percent of plants in samples of raw sewage or final effluents (primary, secondary or lagoon) compared to 65 percent of plants for sludges. Approximately the same number (27 to 30) of pesticide/herbicide compounds were detected in raw sewage, secondary effluent and raw and treated sludge samples. The maximum frequency of detection and plant prevalence of pesticide/herbicide compounds was quite large and reasonably uniform for all sample types. The largest number of volatile organic compounds were detected in raw sewage and secondary effluent streams. The maximum frequency of detection of volatile organics compounds ranged from 15 to 55 percent, and the maximum plant prevalence ranged from 32 to 85 percent.

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1.0 INTRODUCTION AND PROJECT OBJECTIVES

1.1 Background

The Ontario Government's White Paper entitled, "Municipal-Industrial Strategy for Abatement (MISA)", released in June of 1986 by the Ontario Ministry of the Environment (MOE), outlined a new program to reduce the flow of toxic chemicals to the Province's receiving waters. MISA is aimed at reducing contaminant loadings from direct industrial discharges and from municipal water pollution control plants (WPCPs).

The program for municipal WPCPs will involve two stages. In the first stage, a Monitoring Regulation will be promulgated, requiring all WPCPs in Ontario to monitor HCs in effluents and sludges. In the second stage, maximum concentration requirements of HCs in effluents and sludges will be established and Compliance Regulations will be implemented.

In order that MOE can proceed with the establishment of the Monitoring Regulations, there is a need to determine the nature and incidence of HCs in WPCP effluents and sludges, to examine factors affecting the removal of HCs in wastewater treatment facilities and to evaluate the impact of upstream sanitary sewer users upon the HCs observed in WPCP effluents and sludges.

This study was jointly sponsored by the MOE, Environment Canada and the Municipal Engineers Association (MEA) with a goal of providing the information needed to support the development of a cost-effective and practical Monitoring Regulation.

The following is an interim report summarizing the study methodology used in the 37 WPCP study and presenting preliminary study findings. A more detailed analysis of the study data to determine factors affecting removal of HCs, and to allow prioritization of HCs in effluents and sludges and estimation of HC loadings from the study WPCPs will be presented in the final report.

The report has been organized into two volumes. Volume I has the following contents:

Section 2.0 - Field Program Methodology

This section presents a detailed description of the field program methodology.

Section 3.0 - Analytical Methodology

This section presents details of analytical methods as well as indicating the laboratories participating in the study.

Section 4.0 - WPCP Characteristics

This section outlines the characteristics of all WPCPs in Ontario and compares their characteristics with the 37 WPCPs studied.

Section 5.0 - Results and Analysis

This section presents in summary the preliminary results of the monitoring program.

Volume II of the preliminary report is comprised of individual appendices presenting design, operating and other background data for each WPCP as well as a complete summary of the preliminary results of monitoring.

1.2 Study Objectives

The goal of the Municipal WPCP Pilot Program was to obtain the information necessary to support the development of cost-effective and practical Monitoring Regulation for the municipal sector.

Specific project objectives included:

1. To carry out a well designed and rigorously controlled program of hazardous contaminant sampling of sludge and sewage streams at 37 Ontario WPCPs.
2. To summarize all study findings into a comprehensive project report and to provide a well organized database of HC and other results as well as process and flow measurements in a suitable electronic format.
3. To develop a prioritized list of contaminants observed in the study.
4. To assess the effectiveness of WPCPs in removing HCs and to identify (insofar as possible) the factors influencing removal efficiencies including any 'key' variables that may be employed as indicators of HC removal effectiveness.
5. To estimate the loadings discharged in the sludge and liquid effluents of the study WPCPs.

6. To assess the impact of industrial, residential and commercial sanitary sewer inputs upon the nature and loadings of HCs observed in the raw wastewaters, and sludges.
7. To identify and review major concerns affecting the implementation of the monitoring regulation and to make recommendations as needed to address any anticipated implementation problem areas.

1.3 Scope

The field monitoring program involved sampling of influent, final effluent and raw and treated sludges for one to two 5 consecutive day periods at each of the 37 study plants. Each sample was analyzed for all of the contaminants on a list established by MOE for this study. The monitoring list included 144 organic contaminants, 15 metals and conventional contaminants. A comprehensive QA/QC program was carried out in order to be able to define the applicability of the analytical results.

Three laboratories contracted by MOE (Zenon Environmental Ltd., Mann Testing Ltd. and Enviroclean Ltd.) and MOE Laboratory Services Branch (LSB) carried out all of the analytical work, under the direction of MOE.

The Project Liaison Committee directed the field program and defined the requirements for subsequent analytical data summarization, analysis and report writing.

2.1 Selection of WPCPs

The WPCPs for the study were selected by MOE on the basis of the following criteria:

No.	Criteria
1.	All WPCPs which discharged more than 45,000 m ³ /day effluent in 1986, regardless of treatment type. There were 16 secondary treatment plants and 7 primary treatment plants that fit this criterion. These WPCPs contributed 69.1 percent of the total flows discharged by the 406 plants in Ontario in 1986.
2.	Secondary WPCPs that were previously monitored by the Upper Great Lakes Connecting Channels (UGLCC) study in the summer of 1986. Eleven plants were monitored by UGLCC(Ref.1). Three of the 11 plants were already selected under Criterion 1. Consequently, only 5 additional secondary plants with an average daily flow in 1986 of less than 45,000 m ³ /day were selected under Criterion 2 alone. Three plants (Chatham secondary WPCP, Amhurstburg and Point Edward, primary WPCPs) were not remonitored in this study.
3.	Small secondary treatment plants with flows less than 20,000 m ³ /d which produced effluent quality typical of the effluent quality achieved in Ontario by well-operated secondary WPCPs were also selected. "Typical" effluent quality was defined as effluent BOD and TSS concentrations between 8 and 15 mg/L, and total phosphorus (TP) concentrations between 0.7 and 1.0 mg/L. Plant location was also considered in plant selection. It was more desirable in terms of economics and logistics, to sample at WPCPs that were in proximity to those selected in Criteria 1 and 2. Seven plants were selected under Criterion 3.
4.	Two lagoons were also selected for monitoring. Lindsay is served by the largest lagoon in Ontario. Niagara-on-the-Lake has a medium sized lagoon treatment facility.

In total, 37 WPCPs were selected by the above criteria. Table 2-1 presents a list of these plants indicating the criteria under which they were selected. Figure 2-1 presents a map showing the location of each WPCP.

Table 2-1
ONTARIO WPCPs SELECTED FOR THE MONITORING PROGRAM

Criteria	WPCP	Treatment Type
1. 1986 Average Daily Flow >45,000 m ³ /d	Brantford	Secondary
	Burlington (Skyway)	Secondary
	Cornwall	Primary
	Guelph	Secondary - Tertiary
	Hamilton	Secondary
	Kingston City	Primary
	Kitchener	Secondary
	London (Greenway)	Secondary
	Mississauga (Clarkson)	Secondary
	Mississauga (Lakeview)	Secondary
	Niagara Falls (Stamford)	Secondary
	Ottawa (Green Cr.)	Primary
	Pickering (Duffin Cr.)	Secondary
	Peterborough	Secondary
	Sarnia	Primary
	Sault Ste.Marie (East)	Primary
	Sudbury	Secondary
	Thunder Bay	Primary
	Toronto (Highland Cr.)	Secondary
	Toronto (Humber)	Secondary
	Toronto (Main)	Secondary
	Waterloo	Secondary
	Windsor (Westerly)	Primary
2. UGLCC and 1986 Average Daily Flow <45,000 m ³ /d	Belle River (Maidstone)	Secondary
	Moore (Corunna)	Secondary
	Sault Ste.Marie (West)	Secondary
	Wallaceburg	Secondary
	Windsor (Little River)	Secondary
3. Small Secondary WPCPs 1986 Average Daily Flow <20,000 m ³ /d	Grimsby (Baker Road)	Secondary
	Kingston Township	Secondary
	London (Pottersburg)	Secondary
	Oakville (South East)	Secondary
	Paris	Secondary
	Toronto (North)	Secondary
	Whitby (Pringle Creek 1)	Secondary
4. Lagoons	Lindsay	Lagoon
	Niagara-on-the-Lake	Lagoon

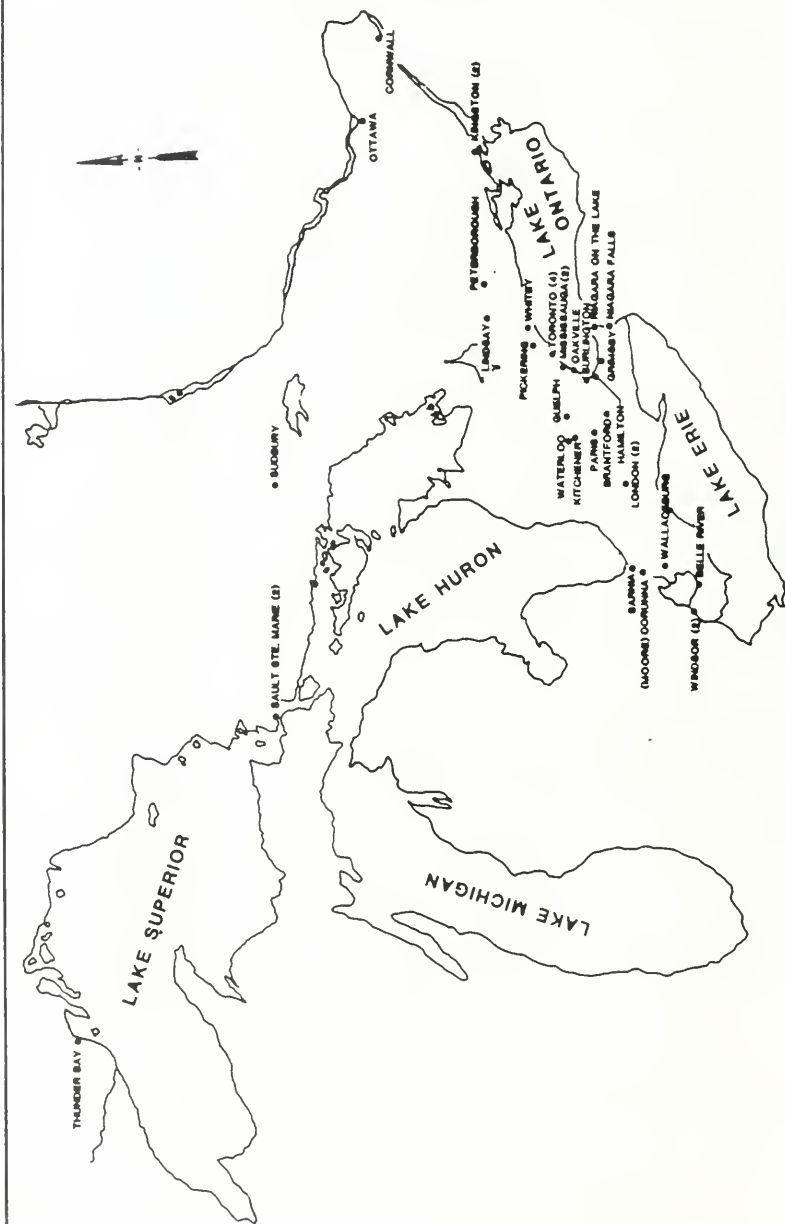


Figure 2-1
GEOGRAPHIC LOCATIONS OF PLANTS INCLUDED IN THE 37 WPCP STUDY

Selection of Target Compounds

The listing of target monitoring parameters for the entirety of the MISA program is presently embodied in the Environmental Monitoring Priority Pollutant List (EMPPL) (Ref. 2). At the time the 37 WPCP study was initiated the EMPPL listing was not as yet formulated and consequently an alternative but nonetheless comprehensive listing of contaminants was identified.

The organic contaminant list used in the present study includes all of the 126 organic contaminants monitored in the U.S. Environmental Protection Agency's (USEPA) Priority Pollutant List (PPL), established in 1972 (Ref. 3). The list also includes additional chlorinated pesticides, nitrogen phosphorus herbicides and phenoxy acid herbicides.

The organic contaminants that were monitored are presented in Table 2-2(a). The list includes 42 volatile organic compounds, 57 base neutral and acid extractable compounds, 35 pesticides and herbicides, and 10 dioxin furans (PCDD/PCDF) compounds. In total there were 144 organic contaminants monitored.

Fifteen metals were also monitored during the study, which are listed in Table 2-2(b). This list includes all of the metals presently regulated in Ontario in sludges utilized on agricultural land. A number of conventional contaminants, cyanide and total phenolics were monitored during the study. These contaminants were selected as general plant performance indicators. The list of conventional contaminants is presented in Table 2-2(c).

Table 2-3 presents those HCs included on the EMPPL but not monitored in this study. This list consists of 80 contaminants.

The organic contaminant list selected for this study is the same as the one used for monitoring by the City of Metropolitan Toronto at their 4 WPCPs in 1984, 1985 and 1986. This list was chosen because the MOE were confident that these contaminants can be satisfactorily monitored and that a long-term data base would be available for the 4 Toronto plants.

Monitoring Program

The sampling program at the 37 selected plants began in January 1987 and was completed in July 1987. In general, each monitoring period involved 2 weeks of pre-monitoring of plant performance and 5 days of sampling. Seventeen plants were monitored for two periods and 20 were monitored for one period only.

Table 2-2(a)
ORGANIC CONTAMINANTS MONITORED IN THE STUDY

Base Neutral and Acid Extractable Compounds	Dioxins/Furans	Pesticides and Herbicides	Volatile Organic Compounds
2,4,5-Trichlorophenol	Tetrachlorodibenzodioxins	Toxaphene*	1,1,1-Trichloroethane*
2,4,6-Trichlorophenol	Tetrachlorodibenzofurans	Total PCB	1,1,2,2-Tetrachloroethane
2,4-Dichlorophenol	Pentachlorodibenzodioxins	Strobane*	1,1,2-Trichloroethane
2,4-Dimethyl phenol	Pentachlorodibenzofurans	Silvex*	1,1-Dichloroethene
2,4-Dinitrotoluene	Hexachlorodibenzodioxins	PF-DDT*	1,2-Dichlorobenzene
2,4-Dinitro-o-cresol	Hexachlorodibenzafurans	PP-DDE*	1,2-Dichloroethane
2,6-Dinitrotoluene	Heptachlorodibenzodioxins	PP-DDD*	1,2-Dichloropropane
2 Hydroxy-toluene (O-Cresol)	Heptachlorodibenzofurans	Photomirex*	1,3-Dichlorobenzene
2-Chloronaphthalene	Octachlorodibenzodioxin	PCNB*	1,4-Dichlorobenzene
2-Chlorophenol	Octachlorodibenzofuran	Oxychlordane*	1-Octene*
2-Nitrophenol*		Mirex*	2-Chloroethylvinyl ether*
3 Hydroxy-toluene (m-Cresol)		Methoxychlor*	3-Chloro-1-propene*
4 Hydroxy-toluene (P-Cresol)		Hexachloroethane	3-Chloro-toluene*
4-Bromophenyl phenyl ether		Hexachlorocyclopentadiene	Acrolein
4-Chlorophenyl phenyl ether		Hexachlorobutadiene	Acrylonitrile
9H Fluorene		Heptachlor Epoxide*	Benzene
Acenaphthene		Heptachlor*	Bromodichlorobenzene*
Acenaphthylene		HCB	Bromodichloromethane
Alpha-naphthylamine*		Gamma-Chlordane*	Bromoethane
Ametryn* (PH)		Gamma-BHC8	Bromoform
Anthracene		Eldrin Aldehyde*	Carbon tetrachloride
Atrazine* (PH)		Eldrin*	Chlorobenzene
Benzo (A) anthracene		Endosulfan Sulphate*	Chloroethane*
Benzo (A) pyrene		Dieldrin*	Chloroform
Benzo (B) fluoranthene		Delta-BHC*	Chloromethane
Benzo (K) fluoranthene		Captan*	cis-1,3-Dichloropropene
Beta-naphthylamine*		Beta-Endosulfan*	cis-1,2-Dichloroethylene*
Biphenyl		Beta-BHC*	Dibromochloromethane
bis(2-Chloro ethoxy) methane		Alpha-Endosulfan*	Dichlorodifluoromethane*
bis(2-Chloro ethyl) ether		Alpha-Chlordane*	Diethyl ether*
bis(2-Chloroisopropyl) ether		Alpha-BHC*	Ethylbenzene
bis(2-ethyl hexyl) phthalate		Aldrin*	Hexane*
Butyl benzyl phthalate		2,4-Dichlorophenoxyacetic acid (2,4-D)*	Hexanol*
Chrysene		2,4-5Trichlorophenoxy- acetic acid (2,4,5-T)*	Methylene chloride
Diazinon* (PH)		1,2,4-Trichlorobenzene	Styrene
Dibenzo (AH) anthracene			Tetrachloroethene
Dicloran* (PH)			Toluene
Diethyl phthalate*			Trans-1,3-Dichloropropene
Dimethyl phthalate*			Trichloroethene
Diphenyl ether			Trichlorofluoromethane
Di-n-butyl phthalate			Vinyl bromide*
Di-n-octyl phthalate			Vinyl chloride
Fluoranthene			
Indeno (123-CD) pyrene			
Malathion* (PH)			
Naphthalene			
Nitrobenzene*			
N-Nitroso Diphenylamine			
N-Nitroso-di-n-propyl-amine			
Parathion ethyl* (PH)			
Parathion methyl* (PH)			
Pentachlorophenol			
Phenanthrene			
Phenol			
Pyrene			
P-chloro-M-cresol*			
Tri-n-tolyl phosphate			

Notes:

* Contaminants not included in EMPPL (Ref. 2)
(PH) Pesticide/Herbicide compound grouped with Base neutral
and acid extractable compounds for analyses

Table 2-2(b)
LIST OF METALS MONITORED IN THE STUDY

Priority Metals:	Arsenic	As
(Regulated by MOE in	Cadmium	Cd
sludge applied to	Chromium	Cr
agricultural land)	Cobalt	Co
	Copper	Cu
	Mercury	Mg
	Molybdenum	Mo
	Nickel	Ni
	Lead	Pb
	Selenium	Se
	Zinc	Zn
Other Metals:	Aluminum	Al
	Beryllium	Be
	Silver	Si
	Strontium	St

Table 2-2(c)
CONVENTIONAL CONTAMINANTS MONITORED IN THE STUDY

<u>Raw Wastewater & Effluent Streams</u>	<u>Sludges</u>
pH	pH
Biochemical Oxygen Demand (BOD ₅)	Chemical Oxygen Demand (COD)
Chemical Oxygen Demand (COD)	Nitrites (NO ₂)
Dissolved Organic Carbon (DOC)	Nitrates (NO ₃)
Total Suspended Solids (TSS)	Ammonia (NH ₃)
Total Volatile Suspended Solids (VSS)	
Filtered Nitrite (NO ₂)	Total Kjeldhal Nitrogen (TKN)
Filtered Nitrates (NO ₃)	Total Phosphorus (TP)
Filtered Ammonia (NH ₃)	Total Solids (TS)
Total Kjeldhal Nitrogen (TKN)	Total Volatile Solids (VS)
Total Phosphorus (TP)	Total Phenolics (4AAP)
Turbidity	Cyanide
Total Phenolics (4AAP)	
Cyanide (Total)	

Table 2-3
 EMPPL (Ref. 2) ORGANIC CONTAMINANTS NOT MONITORED
 IN THE PRESENT STUDY

1,1,3,3-Tetrachloroacetone	Indole
1,1,3-Trichloroacetone	Isopimaric
1,2,3,4-Tetrachlorobenzene	Levopimaric acid
1,2,3,5-Tetrachlorobenzene	Limonene
1,2,3-Trichlorobenzene	Mercapto benzothiazole
1,2,4,5-Tetrachlorobenzene	Methyl ethyl ketone
1,3-Butadiene	Methyl styrene
1,4-Dioxane	Neoabiatic acid
1-Chloronaphthalene	N-Methylformamide
1-Methylnaphthalene	N-Nitrosodimethylamine
1-Nitronaphthalene	Octachlorostyrene
2,3,4,5-Tetrachlorophenol	Oil and grease
2,3,4,6-Tetrachlorophenol	Oleic Acid
2,3,4-Trichlorophenol	Pentachlorobenzene
2,3,5,6-Tetrachlorophenol	Perylene
2,3,5-Trichlorophenol	Pimaric acid
2,3,7,8-Tetrachlorodibenzo-p-dioxin	Specific conductance
2,4,5-Trichlorotoluene	Sulphide
2,6-Di-t-butyl-4-methylphenol	Tetrachloroacetone
2-Hydroxybiphenyl	Tetrachlorogualacol
2-Methylnaphthalene	Tetraethyl lead
2-Nitronaphthalene	Tetra-alkyl lead
3,3 Dichlorobenzidene	(Total)
4,6-Dinitro-o-cresol	Thiourea
4-Aminoazobenzene	Total organic carbon
4-Chloro-3-methylphenol	(TOC)
4-Hydroxybiphenyl	Trichlorogualacol
Abiatic Acid	Triethyl lead
Acenaphthene, 5-nitro	Trimethylbenzenes
Acridine	(1,2,3 isomers)
Aniline	Trimethylnaphthalene
Benzaldehyde	Tri-alkyl lead (Total)
Benzeneacetone nitrile	Tri-n-Butylphosphate
Benzidine	
Benzyl alcohol	
Bis(2-chloroethyl)ether	
Bromomethane	
Butanal	
Camphene	
Chlorodehydroabiatic acid	
Chromium (hexavalent)	
Dehydroabiatic acid	
Dimethyl disulphide	
Diphenylamine	
Ethylene dibromide	
Ethylene thiourea	
Eugenol	
Formaldehyde	
Hydrazine	
Hydroxycyclohexane (Cyclohexanol)	

During the 5 day sampling period, 24-hour composite samples of raw sewage and final effluent (primary, secondary or lagoon) were collected daily. Also, 5-day composite samples of raw sludge and treated sludge were collected.

2.3.1 WPCP Sampling Schedule

The sampling schedule was organized into two separate survey programs. The "winter" program began on 19 January 1987 and continued until 30 March 1987, involving sampling at 25 WPCPs. The "summer" program occurred between 20 April 1987 and 26 July 1987 and included 29 WPCPs. Seventeen of the WPCPs were sampled in both programs, and the remaining 20 were sampled in only one program. Table 2-4 presents the plants sampled in each program.

It should be noted that the labels "winter" and "summer" were used throughout the study to describe the sampling schedules, however, did not necessarily imply a significant difference in weather temperature or operating conditions between the winter and summer programs.

The study requirements outlined that the monitoring of WPCPs take place during periods of "typical" operation. Operation was considered "typical" if the operating conditions were within a normal range expected for the particular plant. This encompassed a wide range of operational conditions, including minor process upsets and equipment malfunction.

Due to limited time and resources, it was not feasible to monitor plants during periods of exceptional circumstances. Therefore, plant monitoring during periods of process shut downs, process changeovers, large industrial spills and major upsets was avoided, where possible.

In general treatment plants were sampled for 5 consecutive days. Four plants were sampled for an extended 7 day period during the summer program. Specifically, they were: Kitchener WPCP, Ottawa (Green Creek) WPCP, Mississauga (Clarkson) WPCP and Toronto (Highland Creek) WPCP. Tables 2-5 and 2-6 respectively present the winter and summer sampling program schedules.

2.3.2 Pre-Monitoring Site Inspections

Prior to the monitoring at each of the 37 WPCPs in the present study, a senior CANVIRO engineer accompanied by a MOE staff member visited the site. The purposes of this initial visit were to define the sampling sites and to arrange for collection of design information, plant operating records and photographs of the site.

Table 2-4
WPCP SAMPLING PROGRAMS

<u>Plant</u>	<u>Code</u>	<u>Winter</u>	<u>Summer</u>
Belle River (Maidstone)	MA	X	
Brantford	BR	X	X
Burlington (Skyway)	BU	X	X
Cornwall	CO	X	X
Grimsby (Baker Rd.)	GR		X
Guelph	GU	X	X
Hamilton (Woodward)	HA	X	X
Kingston City	KC		X
Kingston Township	KT		X
Kitchener	KI	X	X
Lindsay	LI		X
London (Greenway)	LG	X	X
London (Pottersburg)	LP	X	X
Mississauga (Clarkson)	MC	X	X
Mississauga (Lakeview)	ML	X	X
Moore (Corunna)	CR	X	
Niagara Falls (Stamford)	NF	X	X
Niagara-on-the-Lake	NL		X
Oakville (SE)	OA		X
Paris	PA		X
Peterborough	PT	X	X
Pickering (Duffin Creek)	PD	X	X
Sarnia	SA	X	
Sudbury	SU		X
Sault Ste. Marie (East/Old)	SE	X	
Sault Ste. Marie (West/New)	SW	X	
Thunder Bay	TB		X
Toronto (Highland Creek)	TS	X	X
Toronto (Humber)	TH	X	X
Toronto (Main)	TM	X	X
Toronto (North)	TN		X
Wallaceburg	WA	X	
Waterloo	WT	X	X
Whitby (Pringle Creek #1)	WP		X
Windsor (Little River)	WL	X	
Windsor (Westerly)	WW	X	
Ottawa (Green Creek)	OT		X

Table 2-5
WINTER SAMPLING SCHEDULE

<u>Plant</u>	<u>Sampling Dates</u>
Kitchener	Jan 19 - Jan 23
Waterloo	Jan 19 - Jan 23
Burlington	Jan 19 - Jan 23
Toronto (Main)	Jan 26 - Jan 30
Mississauga (Clarkson)	Mar 2 - Mar 6
Mississauga (Lakeview)	Mar 2 - Mar 6
Toronto (Humber)	Feb 9 - Feb 13
London (Greenway)	Feb 16 - Feb 20
London (Pottersburg)	Feb 16 - Feb 20
Windsor (Little River)	Feb 16 - Feb 20
Windsor (Westerly)	Feb 16 - Feb 20
Hamilton (Woodward)	Feb 23 - Feb 27
Toronto (Highland Creek)	Mar 16 - Mar 20
Sault Ste. Marie (West)	Mar 16 - Mar 20
Sault Ste. Marie (East)	Mar 16 - Mar 20
Peterborough	Mar 9 - Mar 13
Sarnia	Feb 2 - Feb 6
Moore (Corunna)	Feb 2 - Feb 6
Wallaceburg	Feb 2 - Feb 6
Belle River (Maidstone)	Feb 2 - Feb 6
Brantford	Mar 9 - Mar 13
Pickering (Duffin Creek)	Mar 30 - Feb 3
Cornwall	Mar 30 - Feb 3
Guelph	Mar 27 - Mar 27
Niagara Falls (Stamford)	Mar 23 - Mar 27

Table 2-6
SUMMER SAMPLING SCHEDULE

<u>Plant</u>	<u>Sampling Dates</u>
Peterborough	Apr 20 - Apr 24
Lindsay	Apr 20 - Apr 24
Grimsby (Baker Rd.)	Apr 20 - Apr 24
Cornwall	May 4 - May 8
Guelph	May 4 - May 8
Paris	May 11 - May 15
Sudbury	May 11 - May 15
Pickering (Duffin Creek)	May 18 - May 22
Kingston City	May 18 - May 22
Kingston Township	May 18 - May 22
Whitby (Pringle Creek #1)	May 18 - May 22
Niagara Falls (Stamford)	May 25 - May 29
Niagara-on-the-Lake	May 25 - May 29
Thunder Bay	Jun 1 - Jun 5
Toronto (North)	Jun 8 - Jun 12
Oakville (SE)	Jun 8 - Jun 12
Toronto (Main)	Jun 15 - Jun 19
Mississauga (Clarkson)*	Jun 22 - Jun 26
Burlington (Skyway)	Jun 22 - Jun 26
Toronto (Highland Creek)*	Jun 29 - Jul 5
Hamilton (Woodward)	Jun 29 - Jul 3
London (Greenway)	Jul 6 - Jul 10
London (Pottersburg)	Jul 6 - Jul 10
Brantford	Jul 6 - Jul 10
Mississauga (Lakeview)	Jul 13 - Jul 17
Waterloo	Jul 20 - Jul 24
Ottawa (Green Creek)*	Jul 20 - Jul 26
Toronto (Humber)	Jul 20 - Jul 24
Kitchener*	Jul 20 - Jul 26

* Plants sampled for 7 days

The evaluation of the sampling site involved the selection of suitable locations for the sampling equipment and identification and resolution of sampling difficulties. The plant flow monitoring equipment (sewage and sludge meters) were reviewed to assess the equipment accuracy and the ability to flow proportion samples. In addition, the monitoring program was discussed with the plant staff for the purpose of locating field team equipment (refrigerators, monitoring equipment, etc.) and to ensure a full understanding of project requirements.

During the initial visit, arrangements were made to obtain historical performance data for each plant, plant operating record sheets, and plant design reports.

2.3.3 Background Monitoring

In order to ensure that the HC data obtained at the plants would be collected under conditions that were representative of typical plant operation, a two-week presampling process monitoring period was established. After the initial site visit, any limitations in the routine monitoring program were identified. Supplemental monitoring requirements were then determined for any additional process information for the two weeks prior to the sampling period.

In order to define plant performance during the premonitoring period, data was collected for influent and effluent conventional contaminants (BOD₅, TSS, TP, TKN, NH₃-N). At some plants, these parameters were not routinely monitored. It was therefore arranged that a sample be collected by WPCP staff at least once per week and submitted to MOE for analysis.

All of the available performance monitoring information was summarized on a spreadsheet. Figure 2-2 presents an example spreadsheet and shows monitored and derived data.

In certain instances, additional data not routinely collected was obtained by plant staff during the pre-monitoring and sampling period. This information included waste sludge rates, phosphorus removal chemical dosage, raw sludge volumes, digested sludge volumes, etc. The operation of the plant was evaluated for the study period based on discussions with plant operating staff, the background data and all pre-monitoring data.

2.3.4 Sampling Methodologies

2.3.4.1 Sampling Locations

Table 2-7 presents a summary of the sampling locations at each plant.

Figure 2-2 EXAMPLE OPERATIONAL EVALUATION SPREADSHEET

OPERATIONAL EVALUATION FOR:

EXAMPLE WPCP

TREATMENT FACILITY: Secondary
PERIOD ENDING: Feb. 20, 1987
SAMPLING SEASON: Winter (Cold Weather)
DESIGN AVG FLOW: 36,320 gpd

PARAMETER	PRE-SAMPLING PERIOD							PRE-SAMPLING PERIOD							SAMPLING PERIOD						
	DAY 1	DAY 2	DAY 3	DAY 4	DAY 5	DAY 6	DAY 7	DAY 8	DAY 9	DAY 10	DAY 11	DAY 12	DAY 13	DAY 14	DAY 15	DAY 16	DAY 17	DAY 18	DAY 19	DAY 20	DAY 21
RAW SEWAGE FLOW	43,500	40,800	36,300	35,400	35,800	39,000	41,700	34,000	34,900	34,000	34,000	41,700	31,300	27,600	27,600	28,100	28,100	29,500	29,000		
% of Design Flow	119.77%	112.13%	99.94%	97.47%	98.57%	107.38%	114.81%	93.41%	96.09%	93.61%	93.61%	114.81%	86.18%	75.49%	75.49%	77.37%	77.37%	81.22%	79.85%		
Influent BOD (mg/L)	171.0	210.0	164.0	148.0	113.0	124.0	140.0	145.0	140.0	107.0	172.0	226.0	230.0	145.0	173.0	159.0	125.0	217.0	107.0		
Primary BOD (mg/L)	8.0	5.0	5.0	11.0	4.0	3.0	3.0	7.0	6.0	4.0	3.0	5.0	4.0	21.0	27.0	27.0	27.0	3.0	28.0		
% PRIMARY REMOVAL	95.3	97.6	97.0	92.6	96.5	97.6	97.9	95.2	95.9	96.3	98.3	97.8	98.3	85.5	84.4	83.0	78.4	98.6	73.8		
% SECONDARY REMOVAL																					
Influent SS (mg/L)	244.0	290.0	192.0	187.0	151.0	150.0	182.0	171.0	174.0	128.0	172.0	228.0	216.0	214.0	135.0	140.0	174.0	217.0	106.0		
Primary SS (mg/L)	11.0	7.0	8.0	13.0	11.0	7.0	8.0	16.0	12.0	11.0	8.0	14.0	7.0	16.0	6.0	8.0	9.0	16.0	9.0		
% PRIMARY REMOVAL	95.5	97.6	95.8	93.2	92.7	95.3	95.6	90.6	93.1	91.4	95.3	93.9	96.8	92.5	95.6	94.3	92.7	92.6	91.5		
% SECONDARY REMOVAL																					
Influent NH ₄ (mg/L)	14.2	12.8					5.4	6.8	6.8						15.7	18.5					
Primary NH ₄ (mg/L)																					
Secondary NH ₄ (mg/L)	1.0	0.6					1.3	0.8	1.4						2.7	2.0					
% PRIMARY REMOVAL	93.2	95.3					75.9	88.4	79.4						82.8	89.2					
Influent TKN (mg/L)	25.3	23.2					14.0	11.7	12.7						27.4	32.6					
Primary TKN (mg/L)																					
Secondary TKN (mg/L)	2.4	2.4					1.3	2.6	1.7						9.0	14.3					
% PRIMARY REMOVAL																					
% SECONDARY REMOVAL	90.5	89.7					90.7	77.8	86.1						67.4	56.1					
Influent Total P (mg/L)	6.90	8.20	4.60	5.10	4.60	4.00	3.70	4.60	5.00	4.30	5.50	8.00	8.50	3.10	7.40	6.70	5.20	7.20	6.40		
Primary Total P (mg/L)																					
Secondary Total P (mg/L)	0.50	0.19	0.34	0.42	0.46	0.30	0.37	1.50	0.38	0.46	0.34	0.64	0.51	1.10	0.80	0.99	0.37	0.35	0.45		
% PRIMARY REMOVAL	92.8	96.9	92.6	91.8	90.0	92.5	91.4	67.4	92.4	89.3	93.8	92.0	94.0	78.4	89.2	84.0	92.7	95.3	93.0		
% SECONDARY REMOVAL																					

Table 2-7
SUMMARY OF SAMPLING LOCATIONS AT THE STUDY WPCP's

Plant	Number of Sampling Locations						Recycle to Raw Sewage Stream
	Raw Sewage	Primary Effluent	Secondary Effluent	Tertiary Effluent	Raw Sludge	Waste (Treated) Sludge	
<u>Tertiary</u>							
Guelph	1	-	1	1	1	AND/DW-1	-
<u>Secondary</u>							
Belle River (Maidstone)	1	-	1	-	1	AD-1	-
Brantford	1	-	1	-	2	AND-1	-
Burlington (Skyway)	1	-	1	-	1	AND-1	-
Grimsby (Baker Road)	1	-	1	-	1	AND-1	-
Hamilton	1	-	1	-	1	AND/DW-1	1
Kingston TWP	1	-	1	-	1	AND-1	1
Kitchener	1	-	1	-	1	AND-1	-
London (Greenway)	1	-	1	-	WAS-1/PRIM-1	DW-1	-
London (Pottersburg)	1	-	1	-	1	-	-
Mississauga (Clarkson)	1	-	1	-	2	COTH/AND-1	-
Mississauga (Lakeview)	2	-	3	-	1	DW/BL/TC-1	-
Moore (Corunna)	1	-	1	-	1 (RAS)	HT-1	-
Niagara Falls (Stamford)	1	-	1	-	1 (PRIM)1 (RBC)	AND-1	-
Oakville (SE)	1	-	1	-	1	AND-1	-
Paris	1	-	1	-	1	AD/TH/HT-1	-
Peterborough	1	-	1	-	1	AND-1	-
Pickering (Duffin Creek)	1	-	1	-	1	AND/DW-1	-
Sault Ste. Marie (West)	1	-	1	-	1	HT/DW-1	1
<u>Notes:</u>	AND - Anaerobically Digested	TC - Thermally Conditioned					
AD - Aerobically Digested	DW - Dewatered	TH - Thickened					
INC - Incinerated	COTH - Co-thickened in Primary Clarifiers	HT - Holding Tank - Supernatant Decanted					
HT - Heat Treated	THO - Thermally Oxidated	GR - Ground					
WAS - Waste Activated Sludge - no treatment		RAS - Return Activated Sludge					
		PRIM - Primary Sludge					
		RBC - Rotating Biological					
		HEAT - Heat Treatment					

Table 2-7
Continued

Plant	Number of Sampling Locations						Recycle to Raw Sewage Stream
	Raw Sewage	Primary Effluent	Secondary Effluent	Tertiary Effluent	Raw Sludge	Waste (Treated) Sludge	
Sudbury	1	-	1	-	-	HT-1	-
Toronto (Highland Creek)	1	-	1	-	1	COTH/AND/GR/HEAT/DW-1	-
Toronto (Humber)	1	-	1	-	PRIM-1/TH-1	TH/AND-1	-
Toronto (Main)	3	-	1	-	PRIM-1/TH-1	TH/AND/THO/DW-1	2
Toronto (North)	1	-	1	-	1	AND/DW-1	-
Waterloo	1	-	2	-	2	AND-2	-
Wallaceburg	1	-	1	-	1	AND/DW-1	-
Whitby (Pringle Cr #1)	2	-	1	-	1	AND-1	1
Windsor (Little River)	1	-	1	-	1	DW-1	1
<u>Primary</u>							
Cornwall	1	1	-	-	1	AND/DW-1	-
Kingston (City)	1	1	-	-	1	AND-1	-
Ottawa (Green Ck)	1	1	-	-	1	AND-1	2
Sarnia	1	1	-	-	1	AND-1	1
Sault Ste. Marie (East)	2	1	-	-	1	-	-
Thunder Bay	1	1	-	-	2	AND-1	-
Windsor (Westerly)	1	1	-	-	1	DW-1	1
<u>Lagoon</u>							
Lindsay	1	2	-	-	-	-	-
Niagara-on-the-Lake	1	1	-	-	-	-	-
<u>Notes:</u>							
AND	-	Anaerobically Digested			TC	- Thermally Conditioned	
AD	-	Aerobically Digested			TH	- Thickened	
DW	-	Dewatered			HT	- Holding Tank - Supernatant Decanted	
INC	-	Incinerated			GR	- Ground	
COTH	-	Co-thickened in Primary Clarifiers			RAS	- Return Activated Sludge	
HT	-	Heat Treated			PRIM	- Primary Sludge	
THO	-	Thermally Oxidated			RBC	- Rotating Biological Contactor Sludge	
WAS	-	Waste Activated Sludge - no treatment			HEAT	- Heat Treatment	

In general, raw sewage was sampled at a point at, or past, the grit removal area which provided good mixing characteristics. If there was aeration at the grit removal area, samples for volatile organic compounds only were collected upstream of this point.

At nine plants it was not possible to obtain a raw sewage sample at a point in the plant before an internal recycle (eg. digester supernatant, waste activated sludge etc.) entered the stream. In these cases, it was also necessary to collect a recycle stream sample so that the recycle contribution to the combined stream in terms of flows and contaminants, could be subtracted to obtain the actual raw wastewater characteristics.

Final effluent streams at all WPCPs except Mississauga (Lakeview) were sampled at a point beyond the point of chlorine addition. Due to logistics, at Lakeview, the effluent sample was taken prior to chlorination and manually chlorinated by the sampling team (subsection 2.3.4.3).

Sampling of raw sludges took place at one or more locations at each plant, depending on the configuration. Samples were only taken during operation of the sludge pumps. Typically, the treatment plants had multiple sumps from where raw sludge could be drawn. In the more complex cases, it was not practical to sample all of the locations each day. In these cases, a sample routine was determined which would allow all locations to be sampled on a regular basis over the 5 day study period. If waste activated sludge was sampled as a separate component of the raw sludge sample it was aliquoted on a flow weighted basis.

Treated sludges were either digested or digested and dewatered. For digested sludges, samples were taken from each digester in service and composited into one treated sludge sample. In plants with dewatering operations, the sludge cake was sampled.

2.3.4.2 Sample Collection Procedures

Raw sewage, final effluent (primary, secondary, lagoon and tertiary) and recycle streams were collected using the following methods:

<u>Sampling Method</u>	<u>Analyses</u>
24-hour flow proportioned composites	Conventionals, metals, cyanide, base-neutral and acid extractable compounds and pesticides and herbicides
5-day composite samples	Dioxin-furan compounds
Grab samples (3 per 24 hours)	Volatile organic compounds

The 24-hour flow proportioned composite samples were collected using one of three techniques, depending on the logistics of the sampling point. These included:

- o Automatic samplers withdrew one individual aliquot each hour. All of the aliquots were manually composited on a flow-proportioned basis at the end of each 24-hour period.
- o Automatic samplers were interfaced with the plant flow recorders so that flow weighted hourly aliquot volumes were added directly into one composite container.
- o Hourly aliquots were grab sampled and composited manually on a flow proportioned basis at the end of each 24-hour period.

For dioxin/furan analyses, flow proportioned aliquots were collected 3 times each day at least 2 hours apart and combined to form 5 day composite samples. Each aliquot was poured directly into the 5-day composite container.

Grab samples for volatile organic compound analyses were collected 3 times per day at least 2 hours apart. The equal volume samples were combined into daily composite samples at the analytical laboratory.

Raw and treated sludge samples collected for all analyses were 5-day flow proportioned composite samples. A minimum of three grab aliquots were collected each day, at least 2 hours apart. There were two methods of making up sludge samples as follows:

- o Individual aliquots were stored separately and combined flow proportionally at the end of the 5-day period to form one 5-day composite sample.
- o Individual flow proportioned aliquots were added directly to the 5-day composite container. This method was only used if the sludge flow was reasonably constant from day to day.

2.3.4.3 Sampling Handling Procotol

In order to ensure the integrity of sample results, a number of cleanliness, security and preservation procedures were carried out in the field.

All of the field equipment coming into contact with the sample was washed with methanol and rinsed with organic free distilled water prior to sampling. When sampling liquid streams, the equipment was also pre-rinsed with the stream

before the sample was taken. Equipment material was either glass, stainless steel, teflon or surgical graded silicon rubber. All equipment was site specific.

For security against breakages in transport or at the laboratory, all samples were collected in duplicate.

Tables 2-8 and 2-9 present the preservation methods used for liquid and sludge samples respectively. In addition to bottle specific preservation methods dependent on the nature of the analyses, all bottles were stored at 4°C in the field and during transport.

Table 2-8
INFLUENT, EFFLUENT AND RECYCLE SAMPLE PRESERVATION

Sample Group	Analysis	# of Samples	Bottle(2)	Preservation(1)
24 hr composite (Automatic or manual)	Base Neutral and Acid Extractable	2/day	1L (3P)	4°C
	Pesticides/Herbicides	2/day	1L (3P)	4°C
	ICAP	2/day	500 mL (20)	HNO ₃ + 4°C
	Mercury	2/day	250 mL (8c)	HNO ₃ + K ₂ CrO ₇ + 4°C
	Conventionals	2/day	1 L (3)	4°C
	Phenolics	2/day	250 mL (8p)	CuSO ₄ + H ₂ PO ₄ + 4°C
	Cyanide	2/day	500 mL (20)	NaOH + 4°C
5 day composite	Dioxins/Furans	2/wk	1L (3p)	4°C
Grabs	Volatile Organics	6/day	50 mL vials	4°C

Notes: (1) Sodium Thiosulphate was added to all effluent samples
(2) MOE bottle description code

Table 2-9
SLUDGE SAMPLE COLLECTION AND PREPARATION

Sample Type	Analysis	# of Samples	Bottle*	Preservation
Sludges (Raw & Treated)	Volatile Organics	2/wk	250 mL (5p)	Methanol 4°C
	Base Neutral & Acid Extractable	2/wk	250 mL (5p)	4°C
	Pesticides & Herbicides	2/wk	250 mL (5p)	4°C
	Dioxins/Furans	2/wk	250 mL (20)	4°C
	ICAP, Mercury, Cyanide	2/wk	500 mL (20)	4°C
	Conventionals	2/wk	250 mL (5)	4°C
	Phenolics	2/wk	250 mL (5)	4°C

* - MOE Bottle Description Code

In addition to the above procedures, sodium thiosulphate ($\text{Na}_2\text{S}_2\text{O}_3$) was added to all final effluent samples to neutralize the chlorine residual. The sodium thiosulphate was added to final effluent sample containers prior to sampling each day.

At one plant (Mississauga (Lakeview)), it was not possible to obtain a chlorinated effluent sample, and non-chlorinated effluent was sampled. In this particular case, the sample was dosed with a concentrated sodium hypochlorite solution to provide a chlorine concentration in the sample equal to the concentration in the chlorinated plant effluent, and mixed slowly for a time equal to the contact time at the plant. The sample was subsequently neutralized with sodium thiosulphate.

2.3.5 Documentation of Field Program

Field sampling personnel were responsible for maintaining two types of records:

- o Process information
- o Sample submissions

Any process information that related to sampled streams was recorded daily. Also, a wide range of sample submission information was recorded in field logs. Records maintained at each treatment plant depended on the type of processes being sampled. A list of typical field record information documented for each sample stream is listed in Table 2-10.

Table 2-10
FIELD RECORD INFORMATION

Information	Stream				
	Influent	Effluent	Recycle	Raw Sludge	Treated Sludge
Hourly Flows	X	X	X		
Daily Flows	X	X	X	X	X
Pump Times			X	X	X
Pump Volumes				X	X
Cl_2 Contact Times		X			
Sample Volume	X	X	X		
Sample Time	X	X	X	X	X
Sample Location	X	X	X	X	X
Sample Weight				X	X
Preservation	X	X	X	X	X
Sample Loss	X	X	X	X	X
Process Irregularity	X	X	X	X	X
Sampler Configuration	X	X	X		
QA/QC Samples	X	X	X	X	X
Samples Submitted	X	X	X	X	X

2.3.6 Field QA/QC Program

The field QA/QC program involved the collection of field blanks. A field blank was an organic free distilled water sample that underwent the same handling in the field and the laboratory as the samples. The purpose of the field blank collection and subsequent analysis was to establish if contamination was being introduced into the samples from the sampling equipment or preservation methods, transportation and/or laboratory handling. In order to determine the sources of contamination, if any, it was necessary to compare field blank results with the laboratory method blank results.

A "grab" sample field blank was prepared by rinsing organic free water in the grab sampling container prior to sampling. The rinse water was placed in the sample bottle and preserved using methods appropriate for the compounds to be analyzed (see Table 2-8).

An automatic sampler field blank was prepared from organic free water which was pumped through the sampler tubing prior to sampling. As above, the water was bottled and preserved according to prescribed methods (Table 2-8).

During the entire program, a total of 19 field blank samples were collected.

In addition to the field QA/QC program, a number of laboratory QA/QC measures were taken; some requiring duplicate sample collection in the field. The laboratory QA/QC procedures are described in Section 3.0.

3.1 Laboratory Analyses

Three laboratories were contracted by MOE to carry out the organics analyses on the samples from the 37 WPCPs. Zenon Environmental Incorporated in Burlington, Ontario did all of the analyses for the volatile organic compounds, dioxin/furan compounds and total phenols. The base neutral and acid extractable compounds were analyzed by Mann Testing Laboratories Ltd., in Mississauga, Ontario and the pesticides and herbicides analyses were carried out in the laboratories of Enviroclean Ltd., in London, Ontario.

In addition, the MOE Laboratory Services Branch (LSB) in Rexdale carried out the analyses for metals, conventional contaminants and cyanide. Table 3-1 presents a complete list of parameters analyzed by each analytical laboratory.

Table 3-2 presents the methods used by the laboratories for analysis of organic compounds and metals. A detailed description of these methods can be found in the individual laboratory reports, summarized by Zenon (Ref. 4). Table 3-3 presents the methods used to analyze the conventional contaminants.

3.2 Laboratory QA/QC Procedures For Trace Organic Compounds

A number of different techniques were regularly used in the laboratory for quality assurance of the analytical results. In addition to these methods, MOE Laboratory Services conducted an external quality assurance program. The quality assurance/quality control methods are presented in the following discussion.

Method Blanks

A method blank was analyzed routinely along with each batch of samples to identify possible contamination contributed by glassware, reagents, other samples, etc. A method blank consisted of an uncontaminated distilled water sample that underwent identical preparation methods (eg. extraction, purge and trap) and was analyzed with the field samples. A method blank was analyzed each time the instrumentation was set up for a new batch of samples.

The method blank analyses were used for two main purposes. Each day of analyses, method blank concentrations of each contaminant were averaged for all of the blanks analyzed that day. The average value was used to correct the concentrations of the particular contaminant in the samples on

Table 3-2
SUMMARY OF ANALYTICAL METHODS FOR TRACE ORGANICS AND METALS USED IN THE STUDY

Contaminant or Contaminant Group	Sample Preparation Method	Analytical Method	Instrumentation	Analytical Laboratory
Volatile Organic Compounds	Purge and trap	Gas Chromatography Mass Spectrometry (GC/MS) Capillary Column	NUTECH 8522 with Finnigan 4510 GCMS with Inco's Data System Enviroclean Series 810 and Hewlett Packard (HP) Mass Selective Detector (MSD)	Zenon Environmental Inc.
PCDD/PCDF	Liquid/liquid extraction and cleanup	GC/MS Capillary Column	Finnigan 4510 GC/MS with Inco's Data System	Zenon Environmental Inc.
Total Phenols	Dilution (if necessary) and distillation from acidified sample	Direct photometric method		Zenon Environmental Inc.
Acid Extractable Compounds	Liquid/liquid extraction and cleanup	GC/MS Capillary column	HP5970 B Mass Selective Detector HP5890 Gas Chromatograph HP9816 Computer System	Mann Testing Laboratories Ltd.
Base/Neutral Extractable Compounds	Liquid/liquid extraction	GC/MS Capillary column	HP5970 B Mass Selective detector HP5890 Gas Chromatograph HP9816 Computer System	Mann Testing Laboratories Ltd.
PCBs and Organochlorine Insecticides	Liquid/liquid extraction and cleanup	GC/MS Dual Capillary Column	Varian 6000 GC	Enviroclean Ltd.
Phenoxy Acid Herbicides	Liquid/liquid extraction and cleanup	GC/MS Dual Capillary Column	Hewlett Packard 5890	Enviroclean Ltd.
Metals (Ag, As, Cd, Cr, Co, Cu, Hg, Mo, Ni, Pb, Se, Zn, Al, Fe, Be, Ca, Mg)		Inductively Coupled Plasma Spectrometry (ICP) Direct Coupled Plasma Spectrometry (DCP)		MOE LSB

Table 3-3
SUMMARY OF ANALYTICAL METHODS FOR CONVENTIONAL CONTAMINANTS
USED IN THE STUDY

Contaminant	Method	Reference*
pH	pH Electrode	Code 001 A11 page 249
Chemical Oxygen Demand (COD)	Colourimetric measurement of trivalent chromium	Code 525 1C2 page 237
Biochemical Oxygen Demand (BOD)	Five day incubation	Code 001 A12 page 234
Dissolved Organic Carbon (DOC)	Filtration glass fibre filter $\leq 2 \mu\text{m}$, combustion at $<1000^\circ\text{C}$, colourimetric detection	Code 102 AC@ page 89
Ammonia plus Ammonium (NH_3)	Distillation, colourimetry	Code 103 DC2 page 191
Nitrate (NO_3)	Colourimetry	Code 102 DC2 page 210
Nitrite (NO_2)	Colourimetry	Code 102 DC2 page 222
Total Kjeldhal Nitrogen (TKN)	Digestion, distillation and colourimetry	Code 004 AC2 page 228
Total Solids (TS)	Drying at $103^\circ\text{C} \pm 3^\circ\text{C}$ and gravimetry	Code 202 A16 page 342
Total Suspended Solids (TSS)	Filtration glass fibre filter $\leq 2 \mu\text{m}$, drying at $103^\circ\text{C} \pm 3^\circ\text{C}$ and gravimetry	Code 506 AD4 Page 348
Volatile Suspended Solids (VSS)	As above, ignite filter for 4 hours at 550°C	Code 506 AD4 page 348
Total Phosphorus (TP)	Digestion, colourimetry	Code 504 AC2 page 279

* "1986 Performance Report", Water Quality Section,
Ministry of the Environment

that day. Secondly, the method blank results for all of the analyses were used to determine if the background "noise" level was too high to use the data for a particular contaminant with confidence.

Duplicates

Duplicate samples were defined as two aliquots taken from a single sample and carried through the same analytical process. The purpose of duplicate analyses was to provide a measure of analytical precision. This was carried out by comparing the differences of each set of duplicates, and determining if the differences were statistically significant.

Native Spikes in Distilled Water Samples

A known amount of standard mixture containing selected native compounds on the monitoring list (Table 2-2) was spiked into a reagent water sample, which subsequently underwent the same preparation and analysis as the field samples.

With each batch of samples analyzed, a blank was spiked with each of the compounds to be analyzed in the sample, and the percentage recovery was documented. The following 2 results were then used to evaluate the applicability of the data for each batch of samples:

- o The recovery of the native compound from the distilled water blank analyzed for each batch of samples.
- o The recovery of the spiked compound from all of the distilled water blanks for the entire study.

Surrogate Spikes in Field Samples

A known amount of mixture containing deuterated target compounds was spiked into the field sample, which was subsequently processed and analyzed. The amount of recovery of the deuterated spike was used to indicate the recovery of the target compound from the sample and the variability of compound recovery.

MOE Laboratory Spiking

Duplicate field samples were sent to the MOE LSB. Some liquid samples and all sludge samples were labelled at the MOE LSB, to indicate they were replicate samples, and were then taken to the contract laboratory for analyses. Other

samples were spiked at the MOE LSB with known concentrations of target compounds of volatiles, organics and pesticides. The spiked samples were relabelled for identification as a spiked replicate of the field sample and then submitted for analyses to the appropriate laboratory. The results of the spiked samples were compared to the replicate unspiked sample. The purpose of the MOE laboratory spiking was to ensure the integrity of the results if the concentrations of native compounds in spiked samples were greater than in non-spiked samples, and to establish parameter stability in transport, storage and analysis.

3.3 Data Management and Review

Analytical and QA/QC results from each of the contract laboratories and from MOE LSB were input into MOE Laboratory Information System (LIS). The results were reviewed and approved by the pertinent MOE laboratory supervisors. The approved results were then transferred from the mainframe LIS to a microcomputer database at MOE using dBase III Plus software (Ashton-Tate) for ease in data analysis and reporting.

The finalized database was sent to CANVIRO for formatting, analysis, interpretation and summarizing.

4.1 Background

Ideally, to effectively characterize wastewater treatment plant influents, effluents, sludges, removal abilities and drainage basin loadings in terms of HCs for Ontario WPCPs, all plants in Ontario would undergo the monitoring program. Since economic and time constraints would not allow for this, it was necessary to select a smaller group of plants that would be representative of all of the Ontario plants.

4.1.1 Ontario WPCPs

In 1987 in Ontario, there were 412 municipal treatment facilities, treating wastewater at a rate of 5.0 million cubic metres per day for a population of over 7 million people.

The 412 treatment facilities had a total hydraulic design capacity of over 6.0 million cubic metres per day. Figure 4-1 shows that in 1987, 82 percent of the facilities in Ontario had design capacities of less than 10,000 m³/day and 36 percent of the 412 facilities were less than 1,000 m³/day. Only 6.8 percent of the plants (27 plants) had capacities greater than 45,000 m³/day but they contributed greater than 70 percent of the total flow in 1987.

Figure 4-2 shows that in 1987, 52 percent of the facilities in Ontario provided secondary treatment, 7.5 percent provided primary treatment, 39 percent were lagoons and 1.7 percent were facilities with no discharge to surface waters (ie. septic tanks, exfiltration plants). Secondary facilities in Ontario generated the largest portion (76.8 percent) of flow in 1987; 70 percent of which was contributed by conventional activated sludge plants. Lagoons and septic tanks typically serve smaller communities. Consequently, total flow contribution from these types of facilities was less than 2 percent.

The 412 facilities in Ontario are located throughout the Province. Larger facilities are primarily located in the Lake Ontario drainage basin, accounting for 58.4 percent of the total flow (based on 1987 flow) from Ontario plants. Lake Erie and Lake Huron received 17.2 and 7.3 percent respectively. The Ottawa River and St. Lawrence River drainage basins received a total of 13.6 percent and flow into Lake Superior, James Bay and Lake Winnipeg was 3.5 percent.

4.1.2 37 WPCPs in the Study

The total flows in 1987 at the 37 WPCPs was 3.7 million cubic metres per day, or 73.6 of the total Ontario flow for that year. Of the study plants, secondary treatment facil-

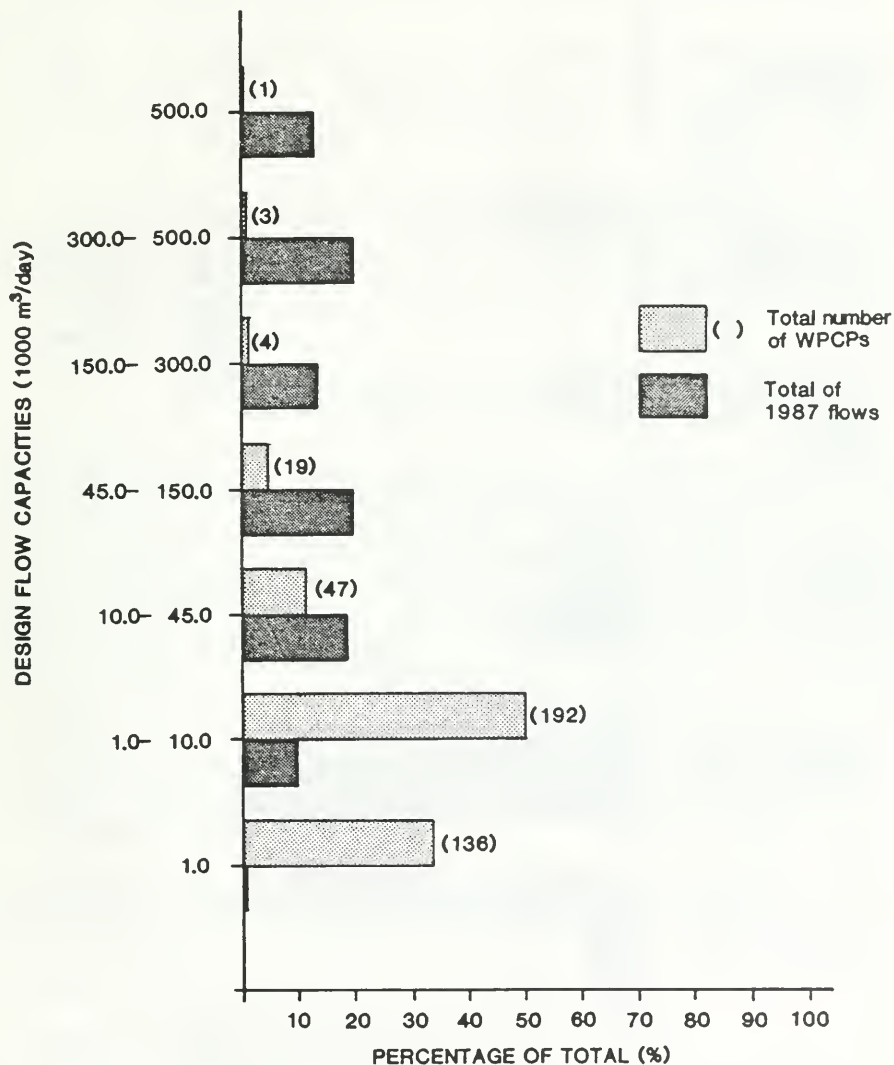


Figure 4-1
HISTOGRAM SHOWING DESIGN FLOW CAPACITIES
FOR ONTARIO WPCPs (1987)

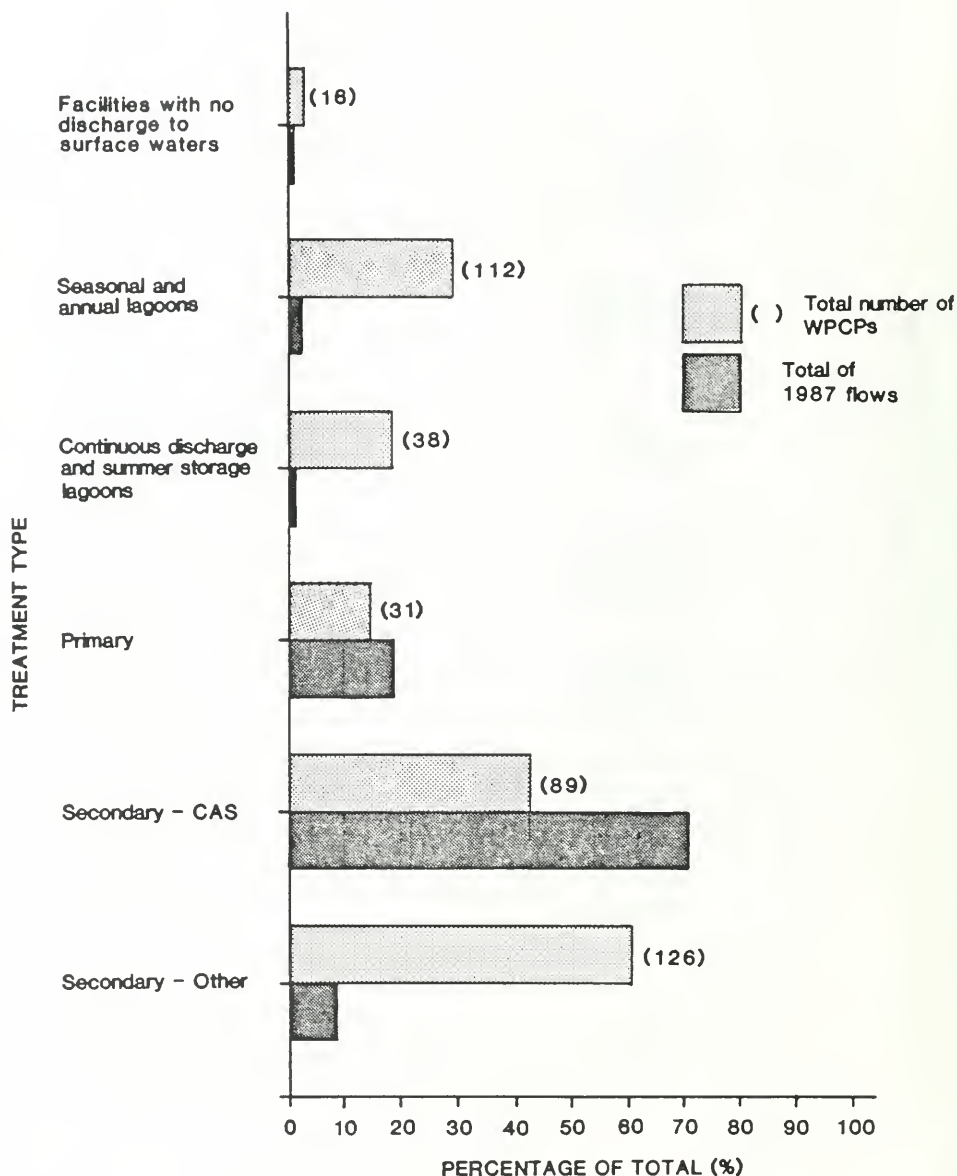


Figure 4-2
HISTOGRAM SHOWING TREATMENT
TYPES FOR ONTARIO WPCPs (1987)

ities contributed to 77.9 percent of flows, primary facilities contributed to 21.6 percent and lagoons to 0.5 percent of these flows.

There were 28 secondary treatment plants involved in the study, comprising 76 percent of the 37 plants. Of these, 23 were conventional activated sludge plants (1 with tertiary treatment), 3 were extended aeration, one was high rate and one used rotating biological contactors.

The largest portion of flows from the selected WPCPs were to the Lake Ontario drainage basin, comprising 64.5 percent. Flows to the Lake Erie drainage basin comprised 16.5 percent, to Lake Huron comprised 2.4 percent and to Lake Superior, 2.2 percent. The Ottawa River and St. Lawrence drainage basins received 14.4 percent of the total flow from the 37 WPCPs.

4.1.3 Comparison Between Ontario WPCPs and 37 WPCPs Selected For Study

The histograms in Figure 4-3 present a comparison of the 37 plants selected for the study to Ontario WPCPs. As noted previously, the study WPCPs represented more than 70 percent of flows from all plants in Ontario in 1987 (Figure 4-3a). The quantities of flows from each type of treatment process (ie. secondary, primary and lagoons) for the study WPCPs are of similar proportions to those for all Ontario WPCPs (Figure 4-3b). In addition, the division of the total Ontario flows to each drainage basin is also represented fairly accurately by the study WPCPs (Figure 4-3d).

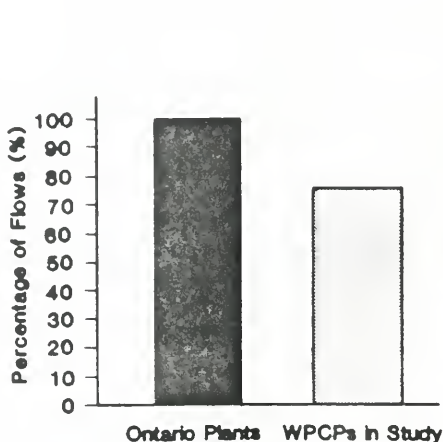
Figure 4-3c shows that a larger percentage of secondary treatment facilities were represented in the study than those existing in Ontario. Since one objective of the present study was to estimate the concentrations and/or removals of HCs in secondary WPCPs, a larger proportion of secondary plants was selected.

In summary, it can be observed that the study group of treatment facilities are a representative portion of all Ontario plants.

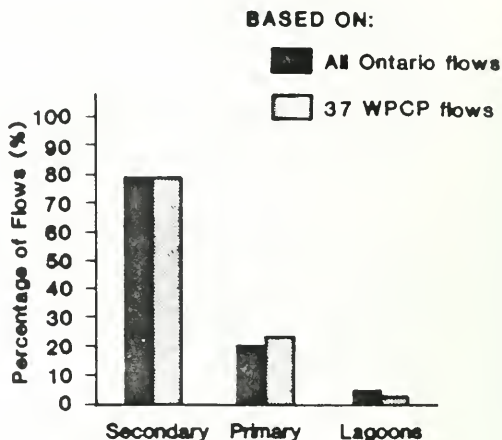
4.2 Characteristics of 37 WPCPs in Study

4.2.1 Summary of Communities

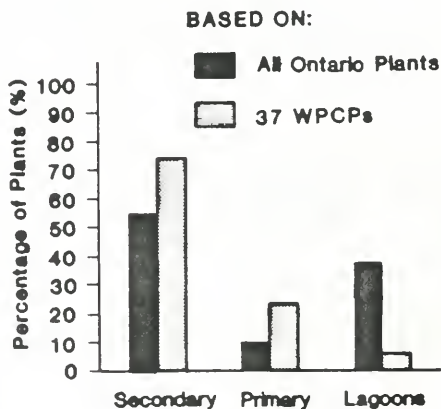
Table 4-1 presents a summary of the characteristics of the communities served by each of the study WPCPs.



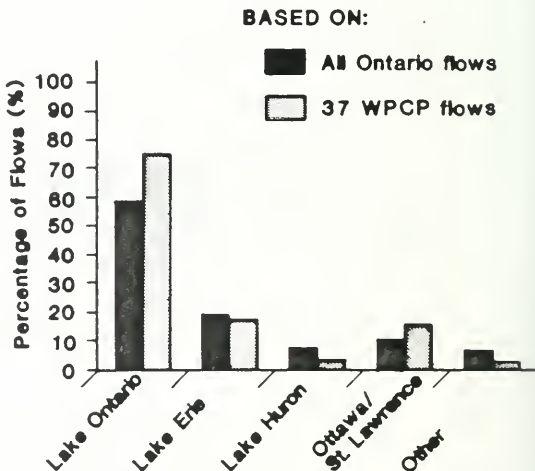
(a) TOTAL 1987 FLOWS



(b) 1987 FLOWS FOR EACH TREATMENT TYPE



(c) PERCENTAGE OF PLANTS WITH EACH TREATMENT TYPE



(d) 1987 FLOW DISTRIBUTION TO DRAINAGE BASINS

Figure 4-3
HISTOGRAM COMPARING ONTARIO WPCPs TO
WPCPs SELECTED FOR THE STUDY

Table 4-1
SUMMARY OF COMMUNITIES OF 37 WPCPs

Plant	Population Served	Receiving Watercourse	Drainage Basin	% Industrial Flow
Guelph	82,000	Grand River	Lake Erie	25%
Belle River (Maidstone)	7,581	Lake St. Clair	Lake Erie	8%
Brantford	73,000	Grand River	Lake Erie	40%
Burlington (Skyway)	120,000	Hamilton Harbour	Lake Ontario	17%
Grimsby (Baker Rd.)	19,850	Lake Ontario	Lake Ontario	0%
Hamilton (Woodward)	300,000	Redhill Creek	Lake Ontario	10%
Kingston TWP	18,700	Lake Ontario	Lake Ontario	4%
Kitchener	138,271	Grand River	Lake Erie	39%
London (Greenway)	164,614	Thames River	Lake Erie	8%
London (Pottersburg)	25,979	Thames River	Lake Erie	9%
Mississauga (Clarkson)	120,000	Lake Ontario	Lake Ontario	25%
Mississauga (Lakeview)	370,000	Lake Ontario	Lake Ontario	12%
Moore (Corunna)	3,405	St. Clair River	Lake Erie	<1%
Oakville (S.E.)	21,900	Lake Ontario	Lake Ontario	<1%
Niagara Falls (Stamford)	67,835	Chippawa Power Canal	Lake Ontario	18%
Paris	4,359	Grand River	Lake Erie	32%
Peterborough	61,063	Otonabee River	Lake Ontario	22%
Pickering (Duffin Creek)	64,386	Lake Ontario	Lake Ontario	25%
Sault Ste. Marie (West)		St. Mary's River	Lake Huron	-
Sudbury	95,000	Junction Creek	Lake Huron	1%
Toronto (Highland Creek)	290,000	Lake Ontario	Lake Ontario	22%
Toronto (Humber)	660,000	Lake Ontario	Lake Ontario	19%
Toronto (Main)	1,200,000	Lake Ontario	Lake Ontario	8%
Toronto (North)	55,000	Don River	Lake Ontario	8%
Waterloo	66,627	Grand River	Lake Erie	13%
Wallaceburg	9,200	Sydenham River	Lake Erie	37%
Whitby (Pringle Ck #1)	10,925	Pringle Creek	Lake Ontario	22%
Windsor (Little River)	64,000	Little River	Lake Erie	24%
Cornwall	46,800	St. Lawrence River	St. Lawrence River	10%
Kingston (City)	67,000	St. Lawrence River	St. Lawrence River	2%
Ottawa (Green Creek)	450,000	Ottawa River	Ottawa River	3%
Sarnia	64,475	St. Clair River	Lake Erie	7%
Sault Ste. Marie (East)	75,000	St. Mary's River	Lake Huron	26%
Thunder Bay	101,529	Kaministiquia River	Lake Superior	9%
Windsor (Westerly)	123,000	Detroit River	Lake Erie	28%
Lindsay	14,636	Scugog River	Lake Ontario	19%
Niagara-on-the-Lake	5,210	Lake Ontario	Lake Ontario	25%

The population of the communities (based on 1987 MOE data) served by the WPCPs range from under 5,000 for Moore (Corunna) and Paris to cities of greater than 100,000 (Burlington, Hamilton, London, Mississauga, Toronto, Thunder Bay, and Windsor).

The receiving water courses for the plants depend on the geographic location of the communities being served. Typically, the receiving water course is a creek or river that is a tributary of one of the Great Lakes. Only 10 plants discharge directly into the major drainage basin of Lake Ontario (3 Toronto plants, 2 Mississauga plants, Pickering, Grimsby, Kingston, Niagara-on-the-Lake and Oakville), and one plant into the St. Lawrence River (Cornwall).

The industrial flow contributions to the study WPCPs range from less than 1 percent for Grimsby (Baker Road) WPCP, Oakville S.E. WPCP and Moore (Corunna) WPCP to about 40 percent for a number of WPCPs. The industrial flow data was taken from a separate MOE study. Municipalities were requested to provide MOE with annual water use data for the industries that discharged to the 37 WPCPs. Industrial flow into each WPCP was then estimated as 85 percent of the total annual water use for 250 days per year. Since annual water use data were not available for many industries, the industrial flow data percentages in Table 4-1 should be considered very approximate (Ref. 4).

4.2.2 Summary of WPCP Design Characteristics

A summary of the WPCP design characteristics, including flows, process type and sludge treatment and disposal methods is presented in Table 4-2.

Ten of the study plants have design flow capacities of greater than 100,000 m³/d, 12 have capacities in the range of 45,000-100,000 m³/d, 9 are in the range of 10,000-45,000 m³/d, and 6 have design capacities of less than 10,000 m³/d.

The percentage utilization of the plant design capacity (based on 1987 average daily flows) ranged from 36% for the new Sault Ste. Marie (West) plant to plants operating at or beyond their hydraulic design capacities (Waterloo, Wallaceburg, Cornwall, Toronto (Humber) and Niagara-on-the-Lake).

All of the secondary plants with the exception of Hamilton (Woodward) and Sudbury practice continuous chemical addition for phosphorus removal. Woodward WPCP used the industrially contributed iron in the raw wastewater for phosphorus removal. Sudbury was not practicing phosphorus removal at the time of the study. However, phosphorus removal equipment is presently being installed. All of the primary plants, with the exception of the Sault Ste. Marie (East) plant, also had continuous addition of chemicals for phosphorus removal. Only Lindsay lagoon uses chemicals for phosphorus removal. The Niagara-on-the-Lake lagoon has no phosphorus removal facilities.

Table 4-2
SUMMARY OF WPCP DESIGN AND FLOW DATA

Plant	Design Flow Capacity (10 ³ m ³ /day)	1987 Average Flow (10 ³ m ³ /day)	1987 Flow as % of Design	Process	Phosphorus Removal	Sludge Treatment	Sludge Disposal
<u>Tertiary Plants</u>							
Guelph	54.55	43.42	79.6	Conventional activated sludge plus RBC's plus filtration	Continuous	Co-thickening/anaerobic digestion/filter dewater- ing	Drying beds or landfill
<u>Secondary Plants</u>							
Belle River (Maldstone)	6.82	5.60	82.1	Extended aeration	Continuous	Aerobic Digestion	Hauled
Brantford	81.83	52.10	63.7	Conventional activated sludge	Continuous	Co-thickening/anaerobic digestion	Agricultural land or lagoon
Burlington (Skyway)	93.19	67.03	71.9	Conventional activated sludge	Continuous	Co-thickening/anaerobic digestion	Incineration and/or agricultural land
Grimsby (Baker Road)	18.18	13.05	71.8	Conventional activated sludge	Continuous	Anaerobic digestion	Agricultural land
Hamilton (Woodward)	409.14	306.47	74.9	Conventional activated sludge	Without Chemicals	Co-thickening/anaerobic digestion/filter dewater- ing	Incineration
Kingston TWP	25.00	18.03	72.1	Conventional activated sludge	Continuous	Anaerobic digestion	Drying bed on lagoon
Kitchener	122.70	70.58	57.5	Conventional activated sludge	Continuous	Anaerobic digestion/ Dissolved air flotation/ belt press dewatering	Agricultural land
London (Greenway)	122.70	110.8	90.3	Conventional activated sludge	Continuous	Co-thickening/anaerobic digestion	Incineration
London (Pottersburg)	22.05	16.33	74.1	Conventional activated sludge	Continuous	Co-thickening/storage	Incineration
Mississauga (Clarkson)	109.10	74.7	68.5	Conventional activated sludge	Continuous	Co-thickening/anaerobic digestion	Incineration or Agricultural land
Mississauga (Lakeview)	284.13	256.9	90.4	Conventional activated sludge	Continuous	Centrifuge thickening/ thermal conditioning/ vacuum filtration/ anaerobic digestion	Incineration and ash lagoon
Moore (Corunna)	4.46	2.18	47.9	Extended aeration	Continuous	Holding tank, decanted	Lagoon
Niagara Falls (Stamford)	58.20	-	-	Rotating biological contactors	Continuous	Anaerobic digestion	Lagoon

Table 4-2
Continued

Plant	Design Flow Capacity (10 ³ m ³ /day)	1987 Average Flow (10 ³ m ³ /day)	1987 Flow as % of Design	Process	Phosphorus Removal	Sludge Treatment	Sludge Disposal
Oakville (S.E.)	22.73	13.52	59.5	Conventional activated sludge	Continuous	Co-thickening/anaerobic digestion	Agricultural land
Paris	7.05	2.52	35.7	Extended aeration	Continuous	Aerobic digestion/ thickening/storage	Agricultural land
Peterborough	68.19	50.79	74.5	Conventional activated sludge	Continuous	Co-thickening anaerobic digestion	Agricultural land
Pickering (Duffin Creek)	189.25	176.0	93.0	Conventional activated sludge	Continuous	Co-thickening anaerobic digestion/filter dewatering	Incineration
Sault Ste. Marie (West)	18.18	6.65	36.6	Conventional activated sludge	Continuous	Co-thickening/filter dewatering	Landfill
Sudbury	68.19	48.97	71.8	High rate	No removal	Anaerobic digestion	Hauled
Toronto (Highland Creek)	218.21	170.0	77.9	Conventional activated sludge	Continuous	Dissolved air flotation/ anaerobic digestion/ grinding/heat treatment/ centrifuge dewatering	Incineration
Toronto (Humber)	409.19	402.7	98.4	Conventional activated sludge	Continuous	Dissolved air flotation/ anaerobic digestion/ elutriation/vacuum filtration	Landfill
Toronto (Main)	818.3	767.2	93.8	Conventional activated sludge	Continuous	Dissolved air flotation/ anaerobic digestion/thermal oxidation/filter dewatering	Incineration
Toronto (North)	45.46	36.65	80.6	Conventional activated sludge	Continuous	Anaerobic digestion/ centrifuge dewatering	Landfill
Waterloo	45.46	46.38	102.0	Conventional activated sludge	Continuous	Co-thickening/anaerobic digestion	Agricultural land
Wallaceburg	6.82	6.76	99.1	Conventional activated sludge	Continuous	Anaerobic digestion/filter dewatering	
Whitby (Pringle Cr. #1)	5.68	3.61	63.6	Conventional activated sludge	Continuous	Co-thickening/anaerobic digestion	Hauled to Whitby (Corbett Cr.) WRCP
Windsor (Little River)	36.32	32.76	90.2	Conventional activated sludge	Continuous	Co-thickening/centrifuge dewatering	Landfill

Table 4-2
Continued

Plant	Design Flow Capacity (10 ³ m ³ /day)	1987 Average Flow (10 ³ m ³ /day)	1987 Flow as % of Design	Process	Phosphorus Removal	Sludge Treatment	Sludge Disposal
<u>Primary Plants</u>							
Cornwall	37.50	43.68	116.5	Primary	Continuous	Anaerobic digestion/ centrifuge dewatering	Landfill
Kingston (City)	61.37	63.48	103.4	Primary	Continuous	Anaerobic digestion (centrifuge dewatering)/ storage	Agricultural land
Ottawa (Green Creek)	545.0	400.25	73.4	Primary	Continuous	Anaerobic digestion	Lagoon
Sarnia	70.47	54.00	76.6	Primary	Continuous	Anaerobic digestion	Lagoon
Sault Ste. Marie (East)	54.55	32.02	58.7	Primary	No removal	Vacuum filtration	
Thunder Bay	109.11	81.11	74.3	Primary	Continuous	Anaerobic digestion	Landfill
Windsor (Westerly)	163.65	123.64	75.6	Primary with polymer addition	Continuous	Centrifuge dewatering/ composting	Agricultural land
<u>Lagoons</u>							
Lindsay	17.18	14.18	82.5	Aerated cells plus lagoon	No removal	No sludge production	-
Niagara-on-the-Lake	3.80	6.40	168.4	Conventional lagoon	No removal	No sludge production	-

A wide range of sludge treatment methods were used at the 37 WPCPs to reduce the sludge volume before ultimate disposal. Processes for pre-thickening (not including co-thickening in the primary clarifiers) are used at 5 facilities. Anaerobic digestion of sludges is used at the majority (26) of the facilities. Two of the plants (Belle River, Clarkson) use aerobic digestion and 7 plants do not have sludge digestion processes before disposal, but do utilize dewatering processes. Additional treatment of digested sludges included dewatering (13 plants), elutriation (Humber WPCP) and heat treatment (Highland Creek WPCP, Main WPCP and Lakeview WPCP).

Three main methods of sludge disposal are utilized, including incineration application to agricultural land and land-filling. In some cases, sludge is transferred to another WPCP for treatment and/or disposal.

4.2.3 Historical WPCP Performance Summary

Table 4-3 presents the annual average effluent concentrations of BOD_5 , suspended solids (TSS) and phosphorus for the 37 plants of this study, for 1986 and 1987. Also indicated is whether the plant complied with the MOE minimum effluent requirements for municipal treatment facilities presented in Table 4-4. It should be noted that plant specific effluent requirements, as required by some WPCPs, were not considered in the evaluation of compliance.

Table 4-3 shows that the secondary plants selected have generally complied in the past with the BOD_5 and TSS requirements. Out of the 7 primary plants, Cornwall WPCP did not comply with TSS requirements in both years, and Ottawa (Green Ck) WPCP did not comply with BOD_5 removal requirements in both years. Both Lindsay and Niagara-on-the-Lake lagoons were in compliance with respect to BOD_5 in both years. However, Lindsay did not comply with TSS limits in 1987.

In 1986 and 1987, only 24 of the study plants were in compliance with the phosphorus requirement of ≤ 1.0 mg/L, assessed on a monthly average basis. Seven plants were out of compliance in one year, and 3 plants (Peterborough, Cornwall and Green Creek) did not comply in either year. Three plants (Sudbury, Sault Ste. Marie East, and Niagara-on-the-Lake) did not have phosphorus removal in 1986 and 1987 and therefore were not subjected to a phosphorus requirement.

Table 4-3
SUMMARY OF HISTORICAL PERFORMANCE OF 37 WPCPs (1981-1986)

Plant	Annual Average Effluent BOD ₅ (mg/L)			Annual Average Effluent TSS (mg/L)			Annual Average Effluent TP (mg/L)			Comments
	1986	1987	Compliance	1986	1987	Compliance	1986	1987	Compliance*	
Tertiary										
Guelph	10.7	14.1		8.9	12.8		1.0	0.6	N	
Secondary										
Belle River (Maldstone)	16.7	4.9		14.8	11.1		0.8	0.7		
Brantford	12.2	13.4		10.4	11.2		0.8	1.0	N	
Burlington (Skyway)	10.8	9.3		9.5	7.3		0.7	0.7		
Grimsby (Baker Road)	13.8	20.6		9.4	11.0		0.5	0.5		
Hamilton	15.7	27.5	N	19.1	12.3		0.7	1.0	N	Phosphorus removal without chemicals
Kingston TWP	8.3	7.5		9.5	8.1		1.0	0.8	N	
Kitchener	12.1	15.5		5.2	5.2		0.7	0.8		
London (Greenway)	4.0	4.8		10.1	12.3		0.7	0.7		
London (Pottersburg)	2.6	3.3		4.3	5.3		0.6	0.6		
Mississauga (Clarkson)	12.7	14.2		9.2	10.0		0.8	0.9		
Mississauga (Lakeview)	16.4	17.8		14.0	14.3		0.6	0.7		
Moore (Corunna)	7.0	7.0		9.8	7.5		0.6	0.4		
Niagara Falls (Stamford)	20.9	11.9		16.5	15.8		0.6	0.7		Primary plant before 1987
Oakville (S.E.)	5.0	2.9		8.4	6.5		0.6	0.5		
Paris	8.7	5.0		6.6	5.4		0.6	0.5		
Peterborough	14.3	9.9		5.5	6.2		3.6	1.0	N	N
Pickering (Duffin Creek)	22.3	19.8		20.6	13.4		1.0	0.6	N	
Sault Ste. Marie (West)	12.2	11.5		9.9	8.6		1.0	0.9	N	
Sudbury	12.9	12.2		12.6	8.3		2.2	2.3	NR	NR
										No phosphorus removal at present. 1988 installation.
Toronto (Highland Creek)	18.7	9.8		23.8	19.0		0.8	0.8		
Toronto (Humber)	11.3	9.0		21.1	20.1		1.0	0.9	N	

Notes: * Compliance is assessed on a monthly average. For compliance, all months TP average ≤ 1.0 mg/L.
NR = No effluent requirements for plants without phosphorus removal.

Table 4-3
Continued

Plant	Annual Average Effluent BOD ₅ (mg/L)			Annual Average Effluent TSS (mg/L)			Annual Average Effluent TP (mg/L)			Comments
	1986	Compliance		1986	Compliance		1986	Compliance*		
		1987	1988		1987	1988		1987	1988	
Toronto (Main)	17.1	11.4		29.6	23.0	N	0.9	0.6		
Toronto (North)	17.9	18.1		6.6	9.3		0.7	0.8		
Waterloo	6.8	7.5		9.3	14.3		0.9	0.8		
Wallaceburg	16.9	11.1		14.2	9.6		0.5	0.7		
Whitby (Pringle Cr. #1)	10.2	6.3		10.4	12.3		0.6	0.6		
Windsor (Little River)	5.4	5.1		9.1	8.8		0.7	0.5		
Primary										
Cornwall	41.1	38.6	N	25.5	28.6		2.5	1.0	N	N
Kingston (City)	23.7	18.1		16.5	16.5		0.6	0.6		
Ottawa (Green Cr.)	34.5	38.6		72.9	32.5	N	1.9	2.2	N	N
Sarnia	33.1	39.1		20.2	24.1		0.7	0.9		
Sault Ste. Marie (East)	69.9	66.1		41.5	38.8		3.4	2.4	NR	NR
Thunder Bay	53.2	57.7		51.4	70.7		0.9	1.0		
Windsor (Westerly)	25.9	24.2		20.6	23.3		0.7	0.7		
Lagoons										
Lindsay	10.5	10.5		81.1	11.0	N	2.6	0.7		
Niagara-on-the-Lake	27.7	23.5		26.9	28.0		3.1	3.3	NR	NR
										No phosphorous removal

Notes: * Compliance is assessed on a monthly average. For compliance, all months TP average ≤ 1.0 mg/L.
NR = No effluent requirements for plants without phosphorus removal.

Table 4-4
MOE 1987 Effluent Discharge Requirements for
Ontario Wastewater Treatment Facilities

Treatment Type	Requirements	Basis
Secondary with phosphorus removal	BOD ₅ ≤25 mg/L	Annual Average
	TSS ₅ ≤25 mg/L	Annual Average
	TP ≤1.0 mg/L	Monthly Average
Secondary without phosphorus removal	BOD ₅ ≤25 mg/L	Annual Average
	TSS ₅ ≤25 mg/L	Annual Average
Primary with phosphorus removal	BOD ₅ Removal ≥50%	Annual Average
	TSS ₅ Removal ≥70%	Annual Average
	TP ≤1.0 mg/L	Monthly Average
Primary without phosphorus removal	BOD ₅ Removal >30%	Annual Average
	TSS ₅ Removal >50%	Annual Average
Lagoon with phosphorus removal	BOD ₅ ≤30 mg/L	Annual Average
	TSS ₅ ≤40 mg/L	Annual Average
	TP ≤1.0 mg/L	Monthly Average
Lagoon without phosphorus removal	BOD ₅ ≤30 mg/L	Annual Average
	TSS ₅ ≤40 mg/L	Annual Average
		Monthly Average

5.1 QA/QC Analytical Results

Detailed descriptions of the QA/QC program results from each contract laboratory are presented in individual laboratory reports, which in turn have been summarized in a report by Zenon Environmental Inc. (Ref. 4).

5.1.1 Detection Limits (DLs)

For the purposes of the present study, each target pollutant in each sample type was assigned a detection limit (DL). It was not intended that the DLs represent the lowest detection capability achievable, but rather, that they reflect a routinely available capability that would serve the needs of the study. In this regard, the statistical significance of a true method detection limit (MDL) (Ref. 5) cannot be used for the DLs. The resulting DLs for the present study for samples of raw wastewater, effluent water and sludges are presented in Tables 5-1(a) to 5-1(c).

For base neutral and acid extractable compounds (Table 5-1(a)) compound DLs were in the range of 10 to 75 µg/L for raw sewage, 2 to 15 µg/L for final effluents and 0.2 to 2 mg/L for sludges.

For volatile organic compounds, (Table 5-1(b)), compound DLs were in the range of 40 - 400 µg/L (with one exception of 5 mg/L for hexanol) for raw sewage, 2 to 100 µg/L (with the exception of 400 µg/L for hexanol) for final effluents, and from 40 to 400 µg/L (5 mg/L for hexanol) for sludges.

For pesticides and herbicides, compound DLs were in the range of 0.02 to 10 µg/L for raw sewage, 0.01 to 2 µg/L for final effluents and 0.2 to 100 µg/L for sludges.

For dioxin/furan analyses, due to the complexity of the samples, the DLs were highly variable depending on the cleanliness, homogeneity and interference associated with an individual sample. Therefore, a DL was established for each individual sample. The DLs for more than 95 percent of the samples ranged from 0.1 to 5 ng/L for raw sewage and final effluents, and from 0.05 to 4 µg/L for sludges. Table 5-1(d) presents the minimum DLs found for each of the compounds.

For metals analyses (Table 5-1(e)) DLs ranged from 0.01 to 0.05 mg/L for raw sewage and final effluents. The DL for mercury was the exception with a value of 0.01 µg/L. The range of metals DLs for sludges was 0.01 to 3 mg/L, with a DL for mercury of 0.01 µg/L. The method detection limit for cyanide was 1 µg/L for all three sample types.

Table 5-1(a)
DETECTION LIMITS BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS

Compound Name	Compound Code	DL Raw Sewage (ug/L)	DL Effluents (ug/L)	DL Sludges (liquid) (ug/L)
2,4,5-Trichlorophenol	X3245	25	5	500
2,4,6-Trichlorophenol	X3246	25	5	500
2,4-Dichlorophenol	PM24DP	25	5	500
2,4-Dimethyl phenol	PM24MP	25	5	500
2,4-Dinitrotoluene	PM26DT	15	3	300
2,6-Dinitrotoluene	PM24DT	15	3	300
2 Hydroxy-toluene (O-Cresol)	PMOCRE	15	3	300
2-Chloronaphthalene	PN2CNA	10	2	200
2-Chlorophenol	X30010	25	5	500
2-Nitrophenol	PM2NP	25	5	500
3 Hydroxy-toluene (M-Cresol)	PMMCRE	15	3	300
4 Hydroxy-toluene (P-Cresol)	PMPCRE	15	3	300
4-Bromophenyl phenyl ether	PM4BPE	15	3	300
4-Chlorophenyl phenyl ether	PM4CPE	10	2	200
9H Fluorene	PNFLUO	10	2	200
Acenaphthene	PNACNE	15	3	300
Acenaphthylene	PNACNY	15	3	300
Alpha-naphthylamine	PMANAA	50	10	1000
Ametryn	P2AMET	25	5	500
Anthracene	PNANTH	10	2	200
Atrazine	P2ATRA	25	2	500
Benzo(a)anthracene	PNBAA	10	2	200
Benzo(a)pyrene	PNBAP	10	2	200
Benzo(b)fluoranthene	PNBBFA	10	2	200
Benzo(k)fluoranthene	PNBKF	10	2	200
Beta-naphthylamine	PMBNAA	75	15	2000
Biphenyl	PNB1PH	15	3	300
bis(2-Chloro ethoxy)methane	PMB2EM	10	2	200
bis(2-Chloro ethyl)ether	PMB2NE	15	3	300
bis(2-Chloroispropyl)ether	PMB2IE	10	2	200
bis(2-ethyl hexyl)phthalate	PMBEHP	10	2	200
Butyl benzyl phthalate	PMBBP	10	2	200
Chrysene	PNCHRY	10	2	200
Diazinon	P4DIAZ	25	5	500
Dibenzo(ah)anthracene	PNDAHA	25	5	500
Dichloran	PODICH	50	10	1000
Diethyl phthalate	PMDEP	25	5	500
Dimethyl phthalate	PMDMP	10	5	200
Diphenyl ether	PMDPE	25	5	500
Di-n-butyl phthalate	PMDNBP	10	2	200
Di-n-octyl phthalate	PMDNOP	15	3	300
Fluoranthene	PNFLAN	10	2	200
Indeno(123-CD)pyrene	PNINP	25	5	500
Malathion	P4MALA	25	5	500
Naphthalene	PNNAPH	10	2	200
Nitrobenzene	PMNITB	10	2	200
N-Nitroso diphenylamine	PMNND	10	2	200
N-Nitroso-di-n-propyl-amine	PMNNP	2	10	200
Parathion ethyl	P4EPAR	25	5	500
Parathion methyl	P4MPAR	25	5	500
Pentachlorophenol	X3PCPH	25	5	500
Phenanthrene	PNPHEN	10	2	200
Phenol	PMPHEN	15	3	300
Pyrene	PNPYR	15	3	300
P-chloro-M-cresol	PMPCMC	25	5	500
Tri-n-tolyl phosphate	POTOC			

Table 5-1(b)
DETECTION LIMITS FOR VOLATILE ORGANIC COMPOUNDS

Compound Name	Compound Code	DL Raw Sewage (µg/L)	DL Effluents (µg/L)	DL Sludges (liquid) (µg/L)
1,1,1-Trichloroethane	X1111T	40	2	40
1,1,2,2-Tetrachloroethane	X11122	40	2	40
1,1,2-Trichloroethane	X1112T	40	5	40
1,1-Dichloroethene	X111CE	40	2	40
1,1-Dichloroethene	X1DCLE	40	2	40
1,2-Dichlorobenzene	X212CB	40	2	40
1,2-Dichloroethane	X112CE	40	2	40
1,2-Dichloropropane	X112CP	40	2	40
1,3-Dichlorobenzene	X213CB	40	2	40
1,4-Dichlorobenzene	X214CB	40	2	40
1-Octene	B10CTE	60	3	60
2-Chloroethylvinyl ether	PM2CEE	40	10	40
3-Chloro-1-propene	X2CPPE	40	2	40
3-Chloro-toluene	X23CTD	40	2	40
Acrolein	X2ACRO	400	100	400
Acrylonitrile	X1ACRY	400	100	400
Benzene	B2BENZ	40	2	40
Bromodichlorobenzene	B2BDCL	40	10	40
Bromodichloromethane	X1BDCM	60	3	60
Bromoethane	X1BETH	40	2	40
Bromoform	X1BROM	60	10	60
Carbon tetrachloride	X1CTET	40	2	40
Chlorobenzene	X2CBEN	40	2	
Chloroethane	X1CHLE	40	2	40
Chloroform	X1CHLO	40	2	40
Chloromethane	X1CHLM	40	20	40
cis-1,3-Dichloropropene	X113DP	60	3	60
cis-1,2-Dichloroethylene	X1CDCE	40	2	
Dibromochloromethane	X1CDBM	40	2	
Dichlorodifluoromethane	X1DCFM	40	20	40
Diethyl ether	E1DIEE	40	2	40
Ethylbenzene	B2BENZ	40	2	40
Hexane	B1HEXA	60	3	60
Hexanol	L1HEX	5000	400	5000
Methylene chloride	X1DCLM	60	3	60
Styrene	B2STYR	40	3	40
Tetrachloroethylene	X1TETR	40	2	
Toluene	B2TOLU	40	2	40
trans-1,3-dichloropropene	X113DR	40	2	40
Trichloroethylene	X1TRIC	40	2	
Trichlorofluoromethane	X1TCFM	40	2	40
Vinyl bromide	B1VBR	60	3	60
Vinyl chloride	X1VCL	100	50	100

Table 5-1(c)
DETECTION LIMITS FOR PESTICIDES AND HERBICIDES

Compound Name	Compound Code	DL Raw Sewage (µg/L)	DL Effluents (µg/L)	DL Sludges (liquid) (µg/L)
1,2,4-Trichlorobenzene	X2124	0.02	0.01	0.2
2,4,5-T	P3245T	0.1	0.05	1
2,4-D	P324D	0.04	0.02	0.4
Aldrin	PlALDR	0.04	0.02	0.2
Alpha-BHC	PlBHCA	0.02	0.01	0.2
Alpha-chlordane	PlCHLA	0.02	0.01	0.2
Alpha-endosulphan	PIEND1	0.02	0.01	0.2
Beta-BHC	PlBHCB	0.02	0.01	0.2
Beta-endosulphan	PlEND2	0.02	0.01	0.2
Captan	POCAPN	0.4	0.2	4
Delta-BHC	PlBCHD	0.02	0.01	0.2
Dieldrin	PlDIEL	0.02	0.01	0.2
Endosulphan sulphate	PlENDS	0.08	0.04	0.8
Eldrin	PlENDR	0.02	0.01	0.2
Eldrin aldehyde	PlENDA	0.4	0.2	4.0
Gamma-BHC	PlBHCG	0.02	0.01	0.2
Gamma-chlordane	PlCHLG	0.02	0.01	0.2
Heptachlor	PlHEPT	0.02	0.01	0.2
Heptachlor epoxide	PlHEPE	0.02	0.01	0.2
Hexachlorobenzene	X2HCB	0.02	0.01	0.2
Hexachlorobutadiene	X1HCB	0.2	0.1	2
Hexachlorocyclopentadiene	X1HCCP	0.2	0.1	2
Hexachloroethane	X1DCLE	10	2	40
Methoxychlor	PlDMDT	0.01	0.05	1.1
Mirex	PlMIRX	0.02	0.01	0.2
Oxychlordane	PlDCHL	0.02	0.01	0.2
PCNB	POPCNB	0.1	0.05	1
Photomirex	PlPMIR	0.02	0.01	0.2
PP-DDD	PlPPDD	0.02	0.01	0.2
PP-DDE	PlPPDE	0.02	0.01	0.2
PP-DDT	PlPPDT	0.02	0.04	0.8
Silvex	P3SILV	0.1	0.05	1
Strobane	PlSTRO	10	5	100
Total PCB	PlPCPT	0.08	0.04	0.8
Toxaphene	PlTOX	0.08	0.04	0.8

Table 5-1(d)
DETECTION LIMITS FOR DIOXIN/FURAN COMPOUNDS

Compound Name	Compound Code	DL Raw Sewage (ng/L)	DL Effluents (ng/L)	DL Sludges (liquid) (ug/L)
Tetrachlorodibenzodioxins	P94CDD	0.5	0.1	0.15
Tetrachlorodibenzofurans	P94CDF	0.2	0.1	1.50
Pentachlorodibenzodioxins	P95CDD	1.0	0.5	0.6
Pentachlorodibenzofurans	P95CDF	0.4	0.1	0.5
Hexachlorodibenzodioxins	P96CDD	1.0	0.3	0.4
Hexachlorodibenzofurans	P96CDF	0.7	0.1	1.0
Heptachlorodibenzodioxins	P97CDD	1.0	0.1	2.0
Heptachlorodibenzofurans	P97CDF	1.0	0.1	1.0
Octachlorodibenzodioxin	P98CDD	1.0	0.3	0.2
Octachlorodibenzofuran	P98CDF	1.0	0.2	0.5

Table 5-1(e)
DETECTION LIMITS FOR METALS AND CYANIDE

Compound Name	Compound Code	DL Raw Sewage (mg/L)	DL Effluents (mg/L)	DL Sludges (liquid) (mg/L)
Aluminum	ALUT	0.02	0.02	0.5
Beryllium	BEUT	0.01	0.01	0.5
Cadmium	CDUT	0.003	0.003	0.5
Calcium	CAUT	0.002	0.002	0.5
Chromium	CRUT	0.01	0.01	0.5
Cobalt	COUT	0.01	0.01	1
Copper	CUUT	0.01	0.01	0.5
Cyanide	CCNFUR	0.001	0.001	0.001
Lead	PBUT	0.03	0.03	0.5
Magnesium	MGUT	0.01	0.01	0.5
Mercury (ug/L)	HGUT	0.01	0.01	0.01
Molybdenum	MOUT	0.01	0.01	0.25
Nickel	NIUT	0.01	0.01	3
Selenium	SEUT	0.03	0.03	1
Silver	AGUT	0.01	0.01	0.5
Strontium	SRUT	0.01	0.01	0.5
Zinc	ZNUT	0.02	0.02	0.5

5.1.2 Method Blank Results

As previously noted in Section 3, one method blank with each batch of samples was routinely analyzed at each contract laboratory performing organics analyses. The results were used to establish the background contamination or "noise level" of each contaminant.

At the end of each day, the arithmetic mean of the compound concentration for each compound was calculated, and subtracted for the compound concentration measured in each sample for that day. This calculation was made to correct samples for the background "noise level".

When the analyses had been completed for all samples for a specific stream type, the arithmetic mean and standard deviation for the concentrations of each contaminant in the method blanks were calculated.

Since the "noise level" varied from day to day, MOE LSB staff felt that the "noise level" averaged over the duration of the study would be representative of the contamination problem with the compound in question, while the "noise level" established for each analytical run may not have been representative of that particular run. For this purpose, MOE established a criterion to determine if the "noise" level was too high to use the sample analytical data with confidence. If twice the standard deviation of the method blank results was greater than the analytical result for a compound averaged for a particular stream type, then the background concentration was considered too high, the entire analytical results for that compound were not considered valid and the compound was excluded from the data presentation for that stream types. Table 5-2(a) presents the compounds detected in 5 percent or more of the laboratory blank samples analyzed. Table 5-2(b) presents a list of the contaminants that did not meet the blank criterion, and are subsequently excluded from further result presentation.

5.1.3 Field Blank Results

There were a total of 19 field blank samples analyzed, including 8 for volatile organic compounds, 9 for base-neutral and acid extractable compounds and pesticide/herbicide compounds and 2 for dioxin/furan compounds. The field blanks were analyzed for all of the target compounds in each group.

Table 5-3 presents the contaminants identified in field blanks.

Table 5-2(a)
CONTAMINANTS DETECTED IN METHOD BLANK SAMPLES

Compound	Average Concentration (µg/L)	Standard Deviation (µg/L)	No. of Samples Analyzed	No of Times Compound was Detected
Di-n-butyl phthalate	1.5	2.8	75	49
Bis-2-Ethylhexyl phthalate	47.7	84.8	75	70
Di-n-octyl phthalate	0.2	0.9	75	6
Di-ethyl phthalate	0.3	1.7	75	7
Methylene chloride	9	6	73	66
Chloroform	3	3	73	29
Toluene	3	5	73	43
Benzene	4	3	73	42
Hexane	7	11	73	40
Bromodichloromethane	3	1	73	9
Methoxychlor	0.08	0.03	34	3

Table 5-2(b)
CONTAMINANTS WITH INVALID ANALYTICAL RESULTS DUE TO METHOD BLANK RESULTS

Contaminant		Sample Types						
Code	Name	Raw Sewage	Primary Effluent	Lagoon Effluent	Secondary Effluent	Tertiary Effluent	Raw Sludge	Treated Sludge
X1CHLO	Chloroform		X		X	X		
B1HEXA	Hexane		X	X	X		X	X
B2BENZ	Benzene		X	X	X		X	X
X1BDCM	Bromodichloromethane		X		X			
B2TOLU	Toluene		X	X	X		X	X
PMPEHP	Bis-2-Ethylhexyl phthalate	X	X	X	X		X	X
PMDNBP	Di-n-butyl phthalate		X	X	X		X	X
PMDNOP	Di-n-octyl phthalate		X	X			X	
PMDEP	Diethyl phthalate			X	X		X	
X1DCLM	Dichloromethane		X	X	X		X	X
PLPMDT	Methoxychlor		X		X			

Notes: X - indicates that the data for the contaminant in the indicated sample type did not meet study QA/QC criteria and was therefore deleted from subsequent evaluation.

Table 5-3
CONTAMINANTS DETECTED IN FIELD BLANKS

Compound Group	Parameter	Concentrations	Frequency
Base-neutral and acid extractables	Di-n-butyl phthalate	3.2 µg/L	1/9 samples
Pesticides/ Herbicides	Methoxychlor	0.1 µg/L, 0.1 µg/L	2/9 samples
	Endosulphan Sulphate	0.09 µg/L	1/9 samples
	2,4-Dichlorophenoxy-acetic acid	0.04 µg/L	1/9 samples
Volatiles	Hexane	13.0 µg/L	1/8 samples

Only 5 organic compounds were detected in the field blanks at concentrations greater than the DLs and in a maximum of 22 percent of tested samples. No dioxin compounds were detected in field blanks.

It was concluded from the field blank results that the level of contamination introduced from the field equipment field sample handling methods and sample transport was not significant.

5.1.4 Results of Duplicate Analyses

Duplicate analyses were carried out in each analytical laboratories in order to determine the variability of the sample results. These results are presented in detail in the individual laboratory reports (Ref. 4).

In a large percentage of the duplicate analyses carried out, one or both of the aliquots results were below the DL and comparisons could not be made. Since there were so few usable duplicate results, the analytical variability as evaluated using these results was inconclusive.

5.1.5 Surrogate Spike Recoveries

Deuterated compounds were added as surrogate spikes to each sample analyzed for volatile and base/neutral acid extractable compounds. Bromofluorobenzene was also added in the spike mixture for volatiles analysis. For dioxin and furan analyses C^{12} labelled tetra and octa dioxin congenors were spiked into the samples. No surrogate spikes were added to samples for pesticide/herbicide analysis since mass spectrometry was not employed for the analysis of these compounds.

Table 5-4(a) - Table 5-4(c) summarize the recoveries of the surrogates by sample type. The summary results show that recoveries of the same surrogate compound in each sample type are very similar (no statistically significant difference at 95% confidence level). Consequently, it was concluded that there was no significant effects of the stream type during this study and the compound recoveries obtained in the blank water spike samples could be used as an indication of average recoveries for all sample types.

Table 5-4(a)
BASE/NEUTRAL COMPOUND SURROGATE RECOVERY SUMMARY BY STREAM TYPE

	Surrogate Compound				
	<u>d₅-phenol</u>	<u>d₄-nitrophenol</u>	<u>d₈-naphthalene</u>	<u>d₁₀-anthracene</u>	<u>d₁₂-benzo-a-pyrene</u>
<u>Raw Sewage</u>					
Average	38%	71%	72.7%	110.3	89.2
Std. dev.	5.4%	15.6%	14.8%	19.7%	14.3%
No. of data averaged*	280	280	280	280	280
<u>Primary Effluent</u>					
Average	38.6%	71.0%	70%	107%	90.5%
Std. dev.	6.3	13.5%	16.8%	15.5%	16.7%
No. of data averaged*	37	37	37	37	37
<u>Secondary Effluent</u>					
Average	37%	71%	67.9%	102.5%	90.5%
Std. dev.	5.2%	15.6%	12.4%	19.2%	15.9%
No. of data averaged*	280	280	280	280	280
<u>Sludges</u>					
Average	52%	70%	72.6%	112.7%	80.5%
Std. dev.	10.9%	18.2%	17.8%	29.2%	17.8%
No. of data averaged*	117	117	117	117	117
<u>Method Blank</u>					
Average	36%	68%	73%	89%	89%
Std. dev.	22%	29%	30%	30%	30%
No. of data averaged*	72	72	72	72	72

Notes: * - No surrogate data were rejected in any case

Table 5-4(b)
DIOXIN/FURAN COMPOUND SURROGATE RECOVERY BY SAMPLE TYPE

Sample Type	13Cl2-T4CDD				13Cl2-08CDD			
	Average % Rec.	Std. Dev. %	No. of data averaged	No. of Data rejected	Average % Rec.	Std. Dev. %	No. of data averaged	No. of data rejected
Primary Effluent	60.8	18.9	8	0	64.3	14.7	7	1
Secondary Effluent	51.8	22.4	49	3	66.9	28.3	52	0
Recycle	59.1	14.4	8	2	64.7	20.5	9	1
Raw Sewage	58.4	27.3	54	3	63.6	24.8	57	0
Sludge	78.3	36.8	74	12	72.3	27.9	85	1
Method Blank	56.4	28.4	24	0	65.6	24.3	23	1
Native Spike	36.7	18.5	12	0	48.6	22.2	12	0

Table 5-4(c)
VOLATILE ORGANIC COMPOUND SURROGATE RECOVERY BY SAMPLE TYPE

Sample Type	d4-Dichloroethane				Bromofluorobenzene				d8-Toluene				d5-Chlorobenzene			
	Avg. % Rec.	Std. Dev. %	No. Averaged	No. Rejected	Avg. % Rec.	Std. Dev. %	No. Averaged	No. Rejected	Avg. % Rec.	Std. Dev. %	No. Averaged	No. Rejected	Avg. % Rec.	Std. Dev. %	No. Averaged	No. Rejected
Primary Effluent	97	12	35	1	92	15	33	3	95	13	36	0	93	16	36	0
Secondary Effluent	96	16	225	8	97	16	205	28	94	12	222	11	96	16	225	8
Recycle 103	18	43	1	99	17	37	7	100	11	42	2	102	11	41	3	
Raw Sewage	100	14	212	5	99	16	194	23	100	10	213	4	99	14	208	9
Sludge	104	12	87	7	100	18	83	11	103	11	93	1	103	13	88	6
Native Spike	99	10	26	2	101	13	25	3	98	11	24	4	100	13	28	0
Method Blank	102	14	65	8	96	31	64	9	98	16	66	7	95	20	69	4

5.1.6 Recovery of Native Spikes from Distilled Water

Tables 5-5(a) to 5-5(d) present summaries of the native spike recoveries from distilled water samples for each compound group. Where no data is presented, spiking of the compound in question was not done.

A system was established by the MOE to use the analytical QA/QC results to identify the qualitative and quantitative applicability of the analytical results. Each contaminant was given a code, which was used to label the value of the result in qualitative and quantitative terms. The codes describe recovery criteria, based on spiking of a native compound into a distilled water sample. If spiking of this native compound was not done, the code used was based on historical recovery data from MOE LSB.

The QA/QC codes and associated recovery criteria, as established by the MOE, are:

<u>QA/QC Code</u>	<u>Recovery Criteria</u>
1	The average recovery of the native compound in distilled water samples was within 50 and 150 percent inclusive, and 70 percent or more of individual recovery data were within 50 and 150 percent.
2	The average recovery of the native compound in distilled water samples and individual percent recovery data do not fit the criteria for 1, 3, 4 and 5.
3	The average recovery of the native compound in distilled water samples is either less than 30 percent, or, more than 30 percent of individual recovery data are less than 30 percent.
4	The average recovery in distilled water samples is greater than 150 percent, or, more than 30 percent of individual percent recovery data are greater than 150 percent.
5	More than 30 percent of individual recovery data are less than 30 percent, and, more than 30 percent of individual recovery data are greater than 150 percent.
0	Analyzed by MOE LSD using internal QA/QC procedures

TABLE 5-5a)
SUMMARY OF RECOVERY OF NATIVE BASE/NEUTRAL AND ACID EXTRACTABLE COMPOUND SPIKES FROM
FROM DISTILLED WATER SAMPLES

		% Recovery Data Obtained In Distilled Water Samples Spiked With Native Compounds											QA
Compound Code	Compound Name	AVG. % R	STD. DEV	DATA	USED DATA	USED DATA	DATA <30% REC	DATA <30% REC	DATA >150% REC	DATA >150% REC	QA //Code		
X3245	2,4,5-TRICHLOROPHENOL	68.7	22.5	71	52	73.2	1	1.4	0	0.0	1	1	
X3246	2,4,6-TRICHLOROPHENOL	83.0	16.0	68	66	97.1	0	0.0	0	0.0	1	1	
PM24DP	2,4-DICHLOROPHENOL	75.6	21.6	68	64	94.1	2	2.9	0	0.0	1	1	
PM24MP	2,4-DIMETHYLPHENOL	57.0	25.0	68	43	63.2	10	14.7	0	0.0	1	2	
PM24NP	2,4-DINITROPHENOL			0								3	
PM24OT	2,4-DINITROTOLUENE	96.0	16.0	68	68	100.0	0	0.0	0	0.0	1	1	
PM26DT	2,6-DINITROTOLUENE	89.0	16.0	68	67	98.5	0	0.0	0	0.0	1	1	
X30010	2-CHLOROPHENOL	67.0	22.0	68	50	73.5	3	4.4	0	0.0	1	1	
PM450P	2-METHYL,6-DINITROPHENOL	98.0	66.0	68	33	48.5	15	22.1	16	23.5	1	2	
PM2NP	2-NITROPHENOL	76.0	21.0	68	61	89.7	0	0.0	0	0.0	1	1	
PM4BPE	4-BROMOPHENYLPHENYLETHER	86.0	17.0	68	66	97.1	0	0.0	0	0.0	1	1	
PM4CPE	4-CHLOROPHENYLPHENYLETHER	83.0	18.0	68	64	94.1	0	0.0	0	0.0	1	1	
PM4NP	4-NITROPHENOL	47.0	19.0	68	32	47.1	11	16.2	0	0.0	1	2	
PM4CNE	ACENAPHTHENE	81.0	17.0	68	66	97.1	0	0.0	0	0.0	1	1	
PM4CNY	ACENAPHTHYLENE	84.0	19.0	68	65	95.6	0	0.0	0	0.0	1	1	
PM4NAA	ALPHA-NAPHTHYLAMINE	74.0	41.0	68	47	69.1	7	10.3	3	4.4	1	1	
P24MET	AMETRYNE	89.0	16.0	68	66	97.1	0	0.0	0	0.0	1	1	
PM4NTH	ANTHRACENE	89.0	15.0	68	67	98.5	0	0.0	0	0.0	1	1	
P24TRA	ATRAZINE	104.0	42.0	68	55	80.9	3	4.4	9	13.2	1	1	
PM4BA	BENZO (a) ANTHRACENE	86.0	21.0	68	65	95.6	0	0.0	0	0.0	1	1	
PM4BP	BENZO (a) PYRENE	83.5	16.8	71	68	95.8	1	1.4	1	1.4	1	1	
PM4BFA	BENZO (b) FLUORANTHENE	86.0	22.0	68	65	95.6	0	0.0	1	1.5	1	1	
PM4GHP	BENZO (g,h,i) PERYLENE	81.0	25.0	68	60	88.2	0	0.0	1	1.5	1	1	
PM4KFP	BENZO (k) FLUORANTHENE	84.0	19.0	68	64	94.1	0	0.0	1	1.5	1	1	
PM4NAA	BETA-NAPHTHYLAMINE	72.0	29.0	68	49	72.1	3	4.4	0	0.0	1	1	
PM4BPH	BIPHENYL	79.0	17.0	68	66	97.1	0	0.0	0	0.0	1	1	
PM4BEM	BIS (2-CHLOROETHOXY) METHANE	73.0	20.0	68	62	91.2	0	0.0	0	0.0	1	1	
PM4B2IE	BIS (2-CHLOROISOPROPYL) ETHER	71.0	24.0	68	53	77.9	1	1.5	0	0.0	1	1	
PM4BCHP	BIS-2-ETHYLHEXYLPHTHALATE	192.0	133.0	68	36	52.9	0	0.0	31	45.6	1	4	
PM4B2NE	BIS- (2-CHLOROMETHYL) ETHER			0								2	
PM4BBP	BUTYLBENZYLPHTHALATE	91.0	24.0	68	66	97.1	0	0.0	1	1.5	1	1	
PM4CNA	CHLORONAPHTHALENE	76.0	23.0	68	63	92.6	3	4.4	0	0.0	1	1	
PM4CHRY	CHRYSENE	87.0	22.0	68	64	94.1	0	0.0	0	0.0	1	1	
P401AZ	DIAZINON	90.0	22.0	68	63	92.6	0	0.0	0	0.0	1	1	
PM4DAH	DIBENZO (a,h) ANTHRACENE	81.0	19.0	68	64	94.1	0	0.0	0	0.0	1	1	
PM4DICN	DICHLORAN	91.0	17.0	68	67	98.5	0	0.0	1	1.5	1	1	
PM4DEP	DIETHYL PHTHALATE	96.0	30.0	68	65	95.6	0	0.0	2	2.9	1	1	
PM4DMP	DIMETHYL PHTHALATE	90.0	18.0	68	66	97.1	0	0.0	0	0.0	1	1	
PM4DPE	DIPHENYL ETHER	79.0	17.0	68	65	95.6	0	0.0	0	0.0	1	1	
PM4DNBP	DI-N-BUTYLPHTHALATE	91.0	20.0	68	66	97.1	0	0.0	0	0.0	1	1	
PM4DNOP	DI-N-OCTYLPHTHALATE	93.0	23.0	68	66	97.1	0	0.0	1	1.5	1	1	
PM4FLAN	FLUORANTHENE	87.0	16.0	68	67	98.5	0	0.0	0	0.0	1	1	
PM4FLDO	FLUORENE	85.0	15.0	68	67	98.5	0	0.0	0	0.0	1	1	
PM4INP	IDENO (1,2,3-cd) PYRENE	81.0	20.0	68	63	92.6	0	0.0	0	0.0	1	1	
P4MA4LA	MALATHION	92.0	15.0	68	68	100.0	0	0.0	0	0.0	1	1	
P4MEPAR	METHYL PARATHION	91.0	18.0	68	67	98.5	0	0.0	0	0.0	1	1	
PM4MCRE	M-CRESOL	77.0	30.0	71	57	80.3	3	4.2	3	4.2	1	1	
PM4NAPN	NAPHTHALENE	76.3	19.1	71	68	95.8	2	2.8	0	0.0	1	1	
PM4NITB	NITROBENZENE	74.9	22.2	71	59	83.1	3	4.2	0	0.0	1	1	
PM4NND	N-NITROSODIPHENYLAMINE	90.0	17.0	68	66	97.1	0	0.0	0	0.0	1	1	
PM4NNP	N-NITROSODI-N-PROPYLAMINE	79.3	29.7	71	58	81.7	6	8.5	2	2.8	1	1	
PM4MCRE	O-CRESOL	41.0	19.0	68	19	27.9	20	29.4	0	0.0	1	2	
P4CEPAR	PARATHION ETHYL	91.0	19.0	68	66	97.1	0	0.0	1	1.5	1	1	
X3PCPH	PENTACHLOROPHENOL	65.8	42.3	71	51	71.8	19	26.8	1	1.4	1	1	
PM4PNHN	PHENANTHRENE	88.0	17.0	68	65	95.6	0	0.0	0	0.0	1	1	
PM4PHEN	PHENOL	40.0	15.0	68	17	25.0	19	27.9	0	0.0	1	2	
PM4PYR	PYRENE	87.0	17.0	68	67	98.5	0	0.0	0	0.0	1	1	
PM4PCMC	P-CHLORO-M-CRESOL	86.0	17.0	68	66	97.1	0	0.0	0	0.0	1	1	
PM4PCRE	P-CRESOL	1.0	7.0	68	1	1.5	67	98.5	0	0.0	1	3	
POTOC	TRI-O-CRESYL PHOSPHATE	92.0	25.0	68	66	97.1	0	0.0	0	0.0	1	1	

TABLE 5-5b)
SUMMARY OF RECOVERY OF NATIVE DIOXIN/FURAN COMPOUND SPIKES FROM DISTILLED
WATER SAMPLES

		% Recovery Data Obtained in Distilled Water Samples Spiked With Native Compounds										
Compound Name		AVG. % R	STD. DEV.			DATA USED	USED DATA	DATA <30% REC	DATA >150% REC	DATA >150% REC	QA Code	
P97CDD	HEPTACHLORODIBENZODIOXIN	79.0	26.7	12	10	83.3	0	0.0	0	0	1	
P97CDF	HEPTACHLORODIBENZOFURAN	101.0	39.7	12	11	91.7	0	0.0	1	8.33	1	
P96CDD	HEXACHLORODIBENZODIOXIN	62.0	21.5	12	8	66.7	0	0.0	1	8.33	2	
P96CDF	HEXACHLORODIBENZOFURAN	80.0	29.9	12	11	91.7	0	0.0	0	0	1	
P98CDD	OCTACHLORODIBENZODIOXIN	83.0	36.8	12	10	83.3	0	0.0	0	0	1	
P98CDF	OCTACHLORODIBENZOFURAN	78.0	26.4	12	11	91.7	0	0.0	0	0	1	
P95CDD	PENTACHLORODIBENZODIOXIN	62.0	21.5	12	8	66.7	0	0.0	0	0	2	
P95CDF	PENTACHLORODIBENZOFURAN	71.0	24.7	12	11	91.7	0	0.0	0	0	1	
P94CDD	TETRACHLORODIBENZODIOXIN	66.0	21.9	12	10	83.3	1	8.3	0	0	1	
P94CDF	TETRACHLORODIBENZOFURAN	54.0	31.4	12	7	58.3	2	16.7	0	0	2	

TABLE 5-5c)
SUMMARY OF RECOVERY OF NATIVE PESTICIDE/HERBICIDE COMPOUND SPIKES FROM
DISTILLED WATER SAMPLES

		% Recovery Data Obtained In Distilled Water Samples Spiked With Native Compounds										QA Code	
Compound Code	Compound Name	AVG.	STD.	% R	DEV.	DATA OF % R	USED DATA	USED DATA	DATA	DATA	DATA	DATA	QA Code
		% R	<30% REC						<30% REC	>150% REC	>150% REC		
X2124	1,2,4-TRICHLOROBENZENE	27.8	18.4	63	31	49.2	13	20.6	0	0.00	0	0	3
P3245T	2,4,5-TRICHLOROPHENOXYACETIC ACID	13.0	45.8	46	1	2.2	45	97.8	0	0.00	0	0	3
P324D	2,4-DICHLOROPHENOXYACETIC ACID	2.4	4.9	46	1	2.2	45	97.8	0	0.00	0	0	3
P1ALDR	ALDRIN	80.6	87.4	63	54	85.7	8	12.7	0	0.00	0	0	1
P1BHCA	Alpha-BHC	77.8	32.1	63	50	79.4	4	6.3	1	1.59	0	0	1
P1CHLA	Alpha-CHLORDANE	85.8	39.2	61	50	82.0	4	6.6	3	4.92	0	0	1
P1BHCB	Beta-BHC	76.3	30.0	63	52	82.5	4	6.3	1	1.59	0	0	1
P0CAPN	CAPTAN	57.0	59.1	48	21	43.8	18	37.5	1	2.08	0	0	3
P1BHCD	Delta-BHC	72.9	40.6	62	43	69.4	11	17.7	1	1.61	0	0	1
P1DIEL	DIELDRIN	64.7	39.1	55	32	58.2	14	25.5	1	1.82	0	0	2
P1ENDA	ELDRIN ALDEHYDE	48.6	29.8	21	8	38.1	6	28.6	0	0.00	0	0	2
P1END1	ENDOSULFAN I	66.4	44.1	55	34	61.8	17	30.9	4	7.27	0	0	3
P1END2	ENDOSULFAN II	52.2	37.2	62	41	66.1	21	33.9	0	0.00	0	0	3
P1ENDS	ENDOSULFAN SULPHATE	55.8	40.7	65	37	56.9	20	30.8	0	0.00	0	0	3
P1ENDR	ENDRIN	55.3	40.8	62	31	50.0	23	37.1	1	1.61	0	0	3
P1BHCG	Gamma-BHC	65.8	33.0	62	41	66.1	10	16.1	1	1.61	0	0	2
P1CHLG	Gamma-CHLORDANE	84.6	41.7	62	48	77.4	10	16.1	3	4.84	0	0	1
P1HEPT	HEPTACHLOR	20.3	19.5	63	5	7.9	50	79.4	0	0.00	0	0	3
P1HEPE	HEPTACHLORPOXIDE	63.8	38.2	57	35	61.4	16	28.1	1	1.75	0	0	2
X2HCB	HEXACHLOROBENZENE	64.8	27.1	63	40	63.5	6	9.5	1	1.59	0	0	2
X1HCB0	HEXACHLOROBUTADIENE	29.7	14.5	62	3	4.8	29	46.8	0	0.00	0	0	3
X1HCCP	HEXACHLOROCHLOROPENTADIENE	42.3	44.4	43	10	23.3	23	53.5	3	6.98	0	0	3
X2HCE	HEXACHLOROETHANE	28.3	13.2	63	2	3.2	34	54.0	0	0.00	0	0	3
P1MDT	METHOXYCHLOR	57.8	35.7	60	46	76.7	17	28.3	1	1.67	0	0	1
P1MIRX	MIREX	63.0	51.0	63	51	81.0	4	6.3	0	0.00	0	0	1
P1PMIR	MIREX PHOTO	64.0	41.1	61	33	54.1	17	27.9	1	1.64	0	0	2
P1OCHL	OXYCHLORDANE	84.7	50.8	62	37	59.7	8	12.9	8	12.90	0	0	2
P1PCBT	PCB, TOTAL			0									2
P0PCNB	PCNB	65.5	27.1	58	39	67.2	5	8.6	1	1.72	0	0	2
P1PPDD	PP-DDD	90.4	47.2	62	46	74.2	9	14.5	7	11.29	0	0	1
P1PPDE	PP-DDE	78.7	37.9	59	43	72.9	7	11.9	2	3.39	0	0	1
P1PPDT	PP-DDT	63.8	65.1	62	34	54.8	21	33.9	7	11.29	0	0	3
P3SILV	SILVEX	4.9	9.7	46	0	0.0	44	95.7	0	0.00	0	0	3
P1STRO	STROBANE			0									3
P1T0X	TOXAPHENE			0									3

TABLE 5-5d)
SUMMARY OF RECOVERY OF NATIVE VOLATILE ORGANIC COMPOUND SPIKES FROM DISTILLED
WATER SAMPLES

Compound Code	Compound Name	% Recovery Data Obtained in Distilled Water Samples Spiked With Native Compounds										QA Code
		AVG. % R	STD. DEV. OF % R	# DATA AVE.	# DATA AVE.	% USED DATA	% USED DATA	% DATA <30% REC	% DATA <30% REC	% DATA >150% REC	% DATA >150% REC	
X1111T	1,1,1-TRICHLOROETHANE	87.0	24.0	28	25	89.28	1	3.6	0	0.0	1	1
X1112T	1,1,2,2-TETRACHLOROETHANE	80.0	19.0	27	26	96.29	0	0.0	0	0.0	1	1
X1112T	1,1,2-TRICHLOROETHANE	86.0	27.0	28	24	85.71	1	3.6	1	3.6	1	1
X111CE	1,1-DICHLOROETHANE	92.0	25.0	27	23	85.18	0	0.0	2	7.4	1	1
X1DCL	1,1-DICHLOROETHENE	84.0	23.0	26	24	92.30	0	0.0	0	0.0	1	1
X212CB	1,2-DICHLOROBENZENE	91.0	44.0	27	24	88.88	0	0.0	1	3.7	1	1
X112CE	1,2-DICHLOROETHANE	84.0	22.0	27	25	92.59	1	3.7	0	0.0	1	1
X112CP	1,2-DICHLOROPROPANE	91.0	22.0	27	26	96.29	0	0.0	0	0.0	1	1
X213CB	1,3-DICHLOROBENZENE	89.0	23.0	27	26	96.29	0	0.0	0	0.0	1	1
X214CB	1,4-DICHLOROBENZENE	91.0	22.0	27	26	96.29	0	0.0	0	0.0	1	1
B1OCT	1-OCTENE	95.0	32.0	25	23	92	0	0.0	2	8.0	1	1
PM2CEE	2-CHLOROETHYL VINYL ETHER	89.0	28.0	17	16	94.11	0	0.0	1	5.9	1	1
X2CPPE	3-CHLOROPROPENE	88.0	49.0	26	19	73.07	1	3.8	3	11.5	1	1
X23CTO	3-CHLOROTOLUENE	89.0	11.0	19	18	94.73	0	0.0	0	0.0	1	1
X1ACRO	ACROLEIN											3
X1ACRY	ACRYLONITRILE											3
X1ACTO	Alpha-CHLOROTOLUENE	82.0	24.0	18	16	88.88	2	11.1	0	0.0	1	1
B2BENZ	BENZENE	101.0	42.0	27	23	85.18	1	3.7	3	11.1	1	1
B2BDCL	BROMODICHLOROBENZENE	83.0	18.0	12	11	91.66	0	0.0	0	0.0	1	1
X1BDCM	BROMODICHLOROMETHANE	86.0	23.0	28	25	89.28	0	0.0	0	0.0	1	1
X1BETH	BROMOETHANE	79.0	18.0	25	24	96	0	0.0	0	0.0	1	1
BRFLB	BROMOFLUOROBENZENE											2
X1BROM	BROMOFORM	83.0	19.0	28	28	100	0	0.0	0	0.0	1	1
X1CTET	CARBON TETRACHLORIDE	79.0	25.0	27	23	85.18	0	0.0	1	3.7	1	1
X2CBEN	CHLOROBENZENE	88.0	16.0	28	27	96.42	0	0.0	0	0.0	1	1
X1CDBM	CHLORODIBROMOMETHANE	93.0	19.0	28	26	92.85	0	0.0	0	0.0	1	1
X1CHLE	CHLOROETHANE	89.0	40.0	14	11	78.57	0	0.0	1	7.1	1	1
X1CHLO	CHLOROFORM	99.0	35.0	28	25	89.28	0	0.0	3	10.7	1	1
X1CHLM	CHLOROMETHANE	86.0	60.0	17		0	2	11.8	3	17.6	1	2
X1CDCE	Cis-1,2-DICHLOROETHYLENE	88.0	19.0	26	25	96.15	0	0.0	0	0.0	1	1
X113DP	Cis-1,3-DICHLOROPROPENE	87.0	19.0	26	25	96.15	0	0.0	0	0.0	1	1
X1DCFM	DICHLORODIFLUOROMETHANE	79.0	37.0	21	13	61.90	1	4.8	1	4.8	1	2
X1DCIM	DICHLOROMETHANE	100.0	37.0	28	23	82.14	1	3.6	3	10.7	1	1
E1DIE	DIETHYL ETHER	97.0	27.0	24	24	100	0	0.0	0	0.0	1	1
B2EBNZ	ETHYLBENZENE	93.0	16.0	28	27	96.42	0	0.0	0	0.0	1	1
B1HEXA	HEXANE	103.0	43.0	28	21	75	0	0.0	4	14.3	1	1
L1HEX	HEXANOL	66.0	20.0	3	2	66.66	0	0.0	0	0.0	1	2
B2MXY	m-, and p-XYLENES	94.0	16.0	26	26	100	0	0.0	0	0.0	1	1
B2OXYL	O-XYLENE	95.0	20.0	20	20	100	0	0.0	0	0.0	1	1
B2STYR	STYRENE	91.0	12.0	19	19	100	0	0.0	0	0.0	1	1
X1TETR	TETRACHLOROETHYLENE	91.0	20.0	27	26	96.29	0	0.0	0	0.0	1	1
B2TOLU	TOLUENE	96.0	26.0	28	26	92.85	0	0.0	1	3.6	1	1
X113DR	Trans-1,3-DICHLOROPROPENE	100.0	32.0	24	23	95.83	0	0.0	2	8.3	1	1
X1TRIC	TRICHLOROETHYLENE	87.0	23.0	27	26	96.29	1	3.7	0	0.0	1	1
X1TCFM	TRICHLOROFUOROMETHANE	85.0	22.0	21	21	100	0	0.0	0	0.0	1	1
X1T12D	TR-1,2-DICHLOROETHYLENE	82.0	28.0	20	18	90	0	0.0	0	0.0	1	1
B1VBR	VINYL BROMIDE	77.0	24.0	21	20	95.23	0	0.0	0	0.0	1	1
X1VCL	VINYL CHLORIDE	64.0	45.0	11	5	45.45	1	9.1	1	9.1	1	2

The above QA/QC codes were used by the MOE to evaluate the applicability of the data, as follows:

<u>QA/QC Code</u>	<u>Data Application</u>
0, 1	Result can be used quantitatively.
2	The result can be used to confirm either the presence or absence of the contaminant, but may not be used quantitatively.
3	If the compound was detected in a sample stream, its presence can be confirmed and the reported concentration is a minimum. However, if it was not detected, its absence cannot be confirmed.
4	If the compound was not detected in a sample stream its absence can be confirmed. However, if it was detected, its presence cannot be confirmed.
5	Neither the absence nor presence of the compound detected or not detected in a sample stream can be confirmed, ie. no conclusions may be made.

The above criteria were applied to all of the contaminants analyzed. The majority of contaminants fell into criteria 1, 2 and 3. Only one contaminant (bis-2-ethylhexyl phthalate) fell into criteria 4 and the results for this compound were previously invalidated using the method blank criterion. There were no contaminants which fell into criteria 5.

5.1.7 MOE LSB Spiking

Duplicate samples were sent from the field to MOE LSB for native compound spiking before being sent to the contract laboratory for analysis, as described in Section 3.2.

The results from these tests were used for observation purposes and were not used for quality control or quality assurance purposes in this study. These results are presented in a separate report by the MOE LSB (Ref. 6).

5.2 Individual WPCP Reports

5.2.1 Background Data

Individual plant background data that was collected for the study included plant historical (1981 - 1985) performance summaries, raw water sources and estimated quantities, pre-monitoring operational data and design information.

Appendix A contains a sub-appendix for each plant in the study, containing (where available) the above background data.

5.2.2 Sampling Program Data

The results of the sampling program contaminant analyses were summarized in individual reports prepared for each plant. Each individual plant report consists of a number of tables, one for each stream sampled at the plant, including raw sewage, final effluent (primary, secondary or lagoon), raw sludge and treated sludge. If there was a recycle stream, an additional table presents this analytical data. Also, the raw sewage results presented are after the recycle contribution in terms of flows and contaminant concentrations has been subtracted.

The individual plant tables for each stream summarize the analytical data for each compound using the following parameters:

- o Compound name
- o Compound code
- o QA/QC code (Section 5.1.2)
- o Number of samples analyzed
- o Number of samples where compound was detected
- o The frequency of detection of the compound in all samples analyzed
- o The maximum concentrations analyzed

In addition, two statistical parameters describe the results; the geometric mean and spread factor. For the purpose of calculating the geometric mean and spread factor in cases where the analytical result was below the associated DL, the value below the DL was assumed to be one half of the DL.

For purposes of comparison, individual plant analytical summary tables include results for all of the plants (global) for a specific stream type.

The individual plant reports containing summary analytical data table for each stream are presented in Appendix A.

5.3 Summary of Sampling Program Results

5.3.1 Data Presentation

In order to satisfy the objectives of this study, it was necessary to summarize the analytical data on a combined WPCP or global level.

The global data base for a particular sample type was made up of all the data obtained from the analysis of all relevant samples at all WPCPs. For the purposes of these summaries, each sample (24 hour composite or 5 day composite) was considered independent of the number of samples taken at the plant or the number of sampling periods at the plant.

A global summary table was prepared for each of the following sample types:

- o Raw sewage (corrected for the effects of included recycle streams)
- o Primary effluent
- o Secondary effluent
- o Lagoon effluent
- o Tertiary effluent
- o Raw sludges
- o Treated sludges

Each table includes the following:

- o Compound code
- o Compound name
- o QA/QC Code (Section 5.1.2)
- o Number of samples analyzed for the compound
- o Number of samples in which the compound was detected
- o The frequency of detection of the compound in all samples
- o The maximum concentration analyzed
- o The minimum concentration above the DL

- o The number of plants at which the compound was analyzed
- o The plant prevalence ie. the percentage of the total number of plants where the compound was detected in at least one sample

In addition, global geometric means and spread factors were calculated, based on the assumptions described in subsection 5.2.2 for values less than the DL.

Also included in the summary tables for liquid streams (ie. raw sewage and effluents) is the compound DL. As previously discussed, this value occasionally varied during the laboratory analyses within the stream type depending on a number of factors. The DL values presented in the summary tables are the limits for over 90 percent of the samples in a stream type. For approximately 10 percent of the samples the analytical laboratory was able to achieve reliable results below those DLs. This will explain the reason that in some cases the minimum reported concentrations presented are lower than the "typical" DL.

For sludges, the DLs used in analyses were based on the liquid sludge sample analyses. Since the contaminant concentrations presented for sludges are on a dry weight basis, the DLs cannot be used for comparative purposes, and are therefore not presented.

5.3.2 Contaminants Not Detected in Any Sample Type

Table 5-6 lists those compounds that were never detected at concentrations above the DLs in any liquid or sludge sample at any of the 37 WPCPs sampled. The list has been partitioned into compounds that are confirmed as not detected and compounds which on the basis of QA/QC results cannot be confirmed as not detected.

In total, 34 compounds were never detected in any sample type, 4 of these were not confirmed. Of the total number, 17 were base neutral and acid extractable compounds, 13 were volatile organic compounds, 3 were pesticide and herbicide compounds and 1 was Tetrachlorodibenzodioxin. There are no metals on this list.

5.3.3 Summary of Contaminants in Raw Wastewater

Table 5-7(a) presents the compounds that were not detected above the DL in any raw wastewater sample. A total of 59 compounds were not detected, including 4 that were not confirmed. Also indicated in this Table are all compounds that were not detected in any stream type (Table 5-6).

TABLE 5-6 - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN ANY STREAM

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
	<u>BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS</u>		
P2ATRA	ATRAZINE	PMPCRE	P-CRESOL
P4DLAZ	DIAZINON		
P4EPAR	PARATHION ETHYL		
P4MALA	MALATHION		
P4MPAR	METHYL PARATHION		
PM24NP	2,4-DINITROPHENOL		
PMANAA	ALPHA-NAPHTHYLAMINE		
PMBNAA	BETA-NAPHTHYLAMINE		
PN2CNA	CHLORONAPHTHALENE		
PNACNE	ACENAPHTHENE		
PND4HA	DIBENZO(a,b)ANTHRACENE		
PNGHIP	BENZO(g,h,i)PERYLENE		
PNINP	IDENO(1,2,3-cd)PYRENE		
PODICH	DICHLORAN		
POTOC	TRI-O-CRESYL PHOSPHATE		
X3245	2,4,5-TRICHLOROPHENOL		
	<u>DIOXINS AND FURANS</u>		
P94CDD	TETRACHLORODIBENZODIOXIN		
	<u>PESTICIDES, HERBICIDES, PCBs</u>		
PISTRO	STROBANE	POCAPN	CAPTAN
		X1HCBD	HEXACHLOROBUTADIENE
	<u>VOLATILES</u>		
B1VBR	VINYL BROMIDE	X1ACRY	ACRYLONITRILE
PM2CEE	2-CHLOROETHYL VINYLETHER		
X11122	1,1,2,2-TETRACHLOROETHANE		
X1112T	1,1,2-TRICHLOROETHANE		
X1BETH	BROMOETHANE		
X1BROM	BROMOFORM		
X1CHLE	CHLOROETHANE		
X1CHLM	CHLOROMETHANE		
X1T12D	TR-1,2-DICHLOROETHYLENE		
X1TCFM	TRICHLOROFLUOROMETHANE		
X1VCL	VINYL CHLORIDE		
X2CPPE	3-CHLOROPROPENE		

TABLE 5-7 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN RAW SEWAGE

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
	<u>METALS AND CYANIDE</u>		
BELT	BERYLLIUM,UNFLT.TOTAL		
	<u>BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS</u>		
P2AMET	AMETRYNE	PMPCRE	P-CRESOL*
P2ATRA	ATRAZINE*		
P4DIAZ	DIAZINON*		
P4EPAR	PARATHION ETHYL*		
P4MALA	MALATHION*		
P4MPAR	METHYL PARATHION*		
PM24DT	2,4-DINITROTOLUENE		
PM24NP	2,4-DINITROPHENOL*		
PMANAA	ALPHA-NAPHTHYLAMINE*		
PM8NAA	BETA-NAPHTHYLAMINE*		
PMDPE	DIPHENYL ETHER		
PN2CNA	CHLORONAPHTHALENE*		
PNACNE	ACENAPHTHENE*		
PNACNY	ACENAPHTHYLENE		
PNANTH	ANTHRACENE		
PNBAP	BENZ(AX)PYRENE		
PNBPH	BIPHENYL		
PNCHRY	CHRYSENE		
PNDAHA	DIBENZO(A,H)ANTHRACENE*		
PNGHIP	BENZ(OK,G,H,I)PERYLENE*		
PNDP	IDENOX(1,2,3-CD)PYRENE*		
PODICH	DICHLORAN*		
POTOC	TRI-O-CRESYL PHOSPHATE*		
X3245	2,4,5-TRICHLOROPHENOL*		
	<u>DIOXINS AND FURANS</u>		
P94CDD	TETRACHLORODIBENZODIOXIN*		
P95CDD	PENTACHLORODIBENZODIOXIN		
P95CDF	PENTACHLORODIBENZOFURAN		
P96CDD	HEXACHLORODIBENZODIOXIN		
P96CDF	HEXACHLORODIBENZOFURAN		
P97CDD	HEPTACHLORODIBENZODIOXIN		
P97CDF	HEPTACHLORODIBENZOFURAN		
	<u>PESTICIDES, HERBICIDES, PCBS</u>		
P0PCNB	PCNB	P0CAPN	CAPTAN*
PIENDIA	ELDRIN ALDEHYDE	X1HCBD	HEXACHLOROBUTADIENE*
P1STRO	STROBANE*		
P1TOX	TOXAPHENE		
	<u>VOLATILES</u>		
B1OCTE	1-OCTENE	X1ACRY	ACRYLONITRILE*
B1VBR	VINYL BROMIDE*		
E1DIEE	DIETHYL ETHER		
PM2CEE	2-CHLOROETHYL VINYLETHER*		
X11122	1,1,2,2-TETRACHLOROETHANE*		
X1112T	1,1,2-TRICHLOROETHANE*		
X113DR	TRANS-1,3-DICHLOROPROPENE		
X1ACTO	ALPHA-CHLOROTOLUENE		
X1BETH	BROMOETHANE*		
X1BROM	BROMOFORM*		
X1CHLE	CHLOROETHANE*		
X1CHLM	CHLOROMETHANE*		
X1DCFM	DICHLORODIFLUOROMETHANE		
X1T12D	TRI-1,2-DICHLOROETHYLENE*		
X1TCFM	TRICHLOROFLUOROMETHANE*		
X1VCL	VINYL CHLORIDE*		
X213CB	1,3-DICHLOROBENZENE		
X214CB	1,4-DICHLOROBENZENE		
X2CBEN	CHLOROBENZENE		
X2CPPE	3-CHLOROPROPENE*		

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

A summary of the compounds that were detected in any raw sewage sample (all WPCPs) is presented in Table 5-7(b). A total of 85 organic contaminants, 14 metals and cyanide were detected at least once in raw sewage samples. However, only 7 metals (Al, Sr, Zn, Hg, Cu, Ni and Cr), 2 base neutral and acid extractable compounds (M-cresol and phenol) and two pesticides and herbicides (2,4-Dichlorophenoxyacetic acid and gamma-BHC) were found in the raw sewage streams at more than 50 percent of WPCPs. The most prevalent volatile organic compounds were detected in raw sewage at fewer than 40 percent of the 37 plants, and the most prevalent dioxin/furans at fewer than 11 percent of plants.

All detected volatile compounds, dioxin/furans compounds and all but the 2 pesticides and herbicides and 2 base neutral and acid extractable compounds (mentioned above) were detected in less than 20 percent of the samples. Metals were most frequently detected contaminant group.

5.3.4 Summary of Contaminants in Primary Effluents

Table 5-8(a) presents the contaminants that were not detected in any primary effluent samples from any of the 7 primary treatment plants. There were a total of 109 compounds not detected, including 7 that were not confirmed. Also indicated in the Table are the 34 compounds that were not detected in any sample at any plant.

A summary of the parameters detected in any primary effluent is presented in Table 5-8(b). A total of 33 organic contaminants, 13 metals and cyanide were detected at least once in the primary effluents.

Four of the 6 base neutral and acid extractable compounds, 4 of the 15 pesticides and herbicides, and 4 of 10 volatile organic compounds were detected in at least 3 of the 7 primary WPCPs. Metals were the most prevalent contaminant with 12 metals detected at at least 3 plants. Dioxin/furan compounds were only detected at 2 plants.

As noted, metals were the most frequently detected contaminants. Six metals (Zn, Sr, Hg, Al, Cr, Cu) were detected in greater than 60 percent of all primary effluent samples. Only one base neutral and acid extractable compound (M-cresol) two pesticides and herbicides (gamma-BHC and 2,4-Dichlorophenoxyacetic acid) and 1 volatile organic compound (Tetrachloroethylene) were present at least 40 percent of samples. The most frequently detected dioxin compound (Octachlorodibenzodioxin) was detected in only 25 percent of the samples.

TABLE 5-7b - GLOBAL SUMMARY OF CONTAMINANTS IN RAW SEWAGE

CONTAM- INANT	CONTAMINANT NAME	UNITS	QA/QC CODE	GLOBAL DET.	GLOBAL # SAMP.	GLOBAL % FREQ.	GLOBAL # PLANTS	GLOBAL PLANTS	GLOBAL % PLANT FREQ.	GLOBAL MEAN	GLOBAL FACTOR	GLOBAL CONC.	GLOBAL CONC. > DL	DET. FACTOR (DL)
CONVENTIONAL														
DOC	DISSOLVED ORGANIC CARBON	mg/L	0	271	271	100.0	37	37	100.0	22.39	1.81	184.00	1.50	1.00
NH4N	NITROGEN-TOT KIEL UNF.TOT	mg/L	0	273	273	100.0	37	37	100.0	23.44	1.47	76.50	1.20	0.10
PH	(LOGH+)(CONC)	mg/L	0	275	275	100.0	37	37	100.0	6.90	1.05	9.27	6.09	1.00
PHUT	PHOSPHORUS UNF.TOT.	mg/L	0	248	248	100.0	30	30	100.0	5.18	1.51	11.90	0.51	0.01
NH4N	AMMONIUM UNF.TOT.	mg/L	0	274	275	99.6	37	37	100.0	14.55	1.45	14.55	1.00	1.00
BOD5	BIOLOGICAL OXYGEN DEMAND	mg/L	0	266	267	99.6	37	37	100.0	140.23	1.93	802.00	11.00	1.00
CHL	CHLOROPHYLL UNF.TOT.	mg/L	0	266	267	99.6	37	37	100.0	126.88	1.93	667.00	6.10	1.00
COD	CHEMICAL OXYGEN DEMAND	mg/L	0	258	260	99.2	37	37	100.0	287.75	1.82	1620.00	12.00	5.00
RSPI OL	RESIDUE PAR LOSS ON IONL	mg/L	0	89	90	98.9	18	18	100.0	100.84	1.78	259.00	24.62	0.00
NH4N	NITRATE UNF.TOT.	mg/L	0	38	271	21.4	19	37	51.4	0.01	2.98	0.97	0.01	0.01
PHOL	PHENOLICS (GAAP)	mg/L	0	37	275	13.5	14	37	37.8	0.31	2.05	18.01	0.38	0.00
NH4N	NITRATES UNF.TOT.	mg/L	0	28	275	10.2	12	37	32.4	0.05	2.33	13.40	0.05	0.05
METALS AND CYANIDE														
SRUT	STRONTIUM UNF.TOT.	ug/L	0	318	319	99.7	37	37	100.0	370.70	2.14	2250.00	60.00	10.00
CUUT	COPPER UNF.TOT.	ug/L	0	48	49	98.0	34	35	97.1	110.60	2.28	660.00	30.00	10.00
ZNUT	ZINC UNF.TOT.	ug/L	0	315	322	97.8	37	37	100.0	21.00	2.94	530.00	19.00	20.00
HGUT	MERCURY UNF.TOT.	ug/L	0	274	275	99.6	36	37	97.3	100.00	2.65	22957.80	101.60	20.00
CUUT	COPPER UNF.TOT.	ug/L	0	237	322	73.6	33	37	89.2	51.10	3.44	820.00	10.00	10.00
NIUT	NICKEL UNF.TOT.	ug/L	0	103	322	32.0	20	37	54.1	38.80	2.70	1469.90	20.00	10.00
CCNUR	CYANIDE-FREEL UNF.TOT.	ug/L	0	82	271	30.3	12	37	32.4	1.90	6.88	19000.00	3.00	1.00
AGUT	SILVER UNF.TOT.	ug/L	0	82	321	25.6	28	37	75.7	10.40	2.55	90.00	8.20	10.00
CUUT	COPPER UNF.TOT.	ug/L	0	82	322	25.5	31	37	83.8	9.30	2.31	80.00	4.50	10.00
CDUT	CADMIUM UNF.TOT.	ug/L	0	76	322	23.6	26	37	70.3	6.50	2.69	259.20	30.00	10.00
PHUT	LEAD UNF.TOT.	ug/L	0	37	322	12.8	21	37	56.8	12.40	1.72	590.00	20.00	10.00
SRUT	STRONTIUM UNF.TOT.	ug/L	0	5	308	1.6	2	37	5.4	17.30	2.07	80.00	30.00	30.00
ASUT	ARSENIC UNF.TOT.	ug/L	0	5	308	1.0	1	37	2.7	16.80	1.83	60.00	60.00	30.00
BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS														
PMACRE	M-CRESOL	ug/L	1	167	275	60.7	32	37	86.5	25.59	3.45	783.95	0.18	15.00
PMPIEN	PIENOL	ug/L	2	118	275	42.9	29	37	78.4	14.52	2.46	276.20	9.97	15.00
PMNPH	BUTYL BENZYLPHENYLATH	ug/L	1	34	275	12.4	14	37	37.8	5.85	1.66	82.90	10.10	10.00
PMNPH	NAPHTHALENE	ug/L	1	16	275	5.8	8	37	21.6	5.37	1.47	46.50	10.30	10.00
PMNPH	PHENANTHRENE	ug/L	1	6	275	2.6	4	37	10.8	5.17	1.35	39.30	2.20	10.00
PMACRE	O-CRESOL	ug/L	2	6	275	2.2	4	37	8.1	7.72	1.39	216.50	15.30	10.00
PMACRE	P-CRESOL	ug/L	1	4	275	1.5	2	37	5.4	5.10	1.37	117.60	10.60	10.00
PMNPH	FLUORENTHENE	ug/L	1	4	275	1.5	2	37	5.4	5.09	1.29	36.60	2.40	10.00
PMNPH	4-NITROPHENOL	ug/L	2	3	275	1.1	3	37	8.1	12.65	1.29	53.30	32.05	25.00
PMNPH	BIS(2-CHLOROETHOXY)METHANE	ug/L	1	2	275	0.7	2	37	5.4	5.09	1.38	106.70	47.70	10.00
PMNPH	BIS(2-CHLOROMETHYL)ETHER	ug/L	2	2	275	0.7	2	37	5.4	7.61	1.34	101.50	39.40	15.00
PMNPH	N-NITROSO-DI-NITROPHENYLAMINE	ug/L	1	2	275	0.7	2	37	5.4	5.09	1.37	103.90	39.00	10.00
PMNPH	2,4-DICHLOROPHENOL	ug/L	2	1	275	0.4	1	37	2.7	12.54	1.27	33.90	25.00	10.00
PMNPH	2,6-DINITROPHENOL	ug/L	1	1	275	0.4	1	37	2.7	7.54	1.28	36.90	36.90	15.00
PMNPH	2-NITROPHENOL	ug/L	1	1	275	0.4	1	37	2.7	12.51	1.26	29.40	29.40	25.00
PMNPH	2-METHYL-6-DINITROPHENOL	ug/L	2	1	275	0.4	1	37	2.7	12.57	1.29	79.40	79.40	25.00
PMNPH	4-BROMOPHENYLPHENYL ETHER	ug/L	1	1	275	0.4	1	37	2.7	7.58	1.35	181.40	181.40	15.00
PMNPH	4-CHLOROPHENYLPHENYL ETHER	ug/L	1	1	275	0.4	1	37	2.7	5.05	1.33	95.50	95.50	10.00

TABLE 5-7b - GLOBAL SUMMARY OF CONTAMINANTS IN RAW SEWAGE

CONTAMINANT	CONTAMINANT NAME	UNITS	QA/QC CODE	GLOBAL # DET.	GLOBAL # SAMPS. TESTED	GLOBAL % DET.	GLOBAL # PLANTS	GLOBAL # PLANTS DET.	GLOBAL MEAN	GLOBAL SPREAD FACTOR	GLOBAL MAX. CONC.	GLOBAL MIN. CONC. > DL	DET. LIMIT (DL)
BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS													
PM20E	BIS(2-CHLOROPHENOXY)ETHYLB	ug/L	1	1	275	0.4	1	37	5.05	1.35	124.00	124.00	10.00
PM20B	NITROBENZENE	ug/L	1	1	275	0.4	1	37	5.03	1.29	28.96	28.96	10.00
PM20D	N-NITROSO DIETHANYLAMINE	ug/L	1	1	275	0.4	1	37	12.59	1.30	104.70	104.70	25.00
PM20C	P-CHLORO M-CRESOL	ug/L	1	1	275	0.4	1	37	5.01	1.29	37.60	37.60	10.00
PM20A	BENZOXANTHURACENE	ug/L	1	1	275	0.4	1	37	5.01	1.27	14.00	14.00	10.00
PM20F	BENZOPHANTHURACENE	ug/L	1	1	275	0.4	1	37	5.03	1.28	33.30	33.30	10.00
PM20H	FLUORENE	ug/L	1	1	275	0.4	1	37	5.02	1.26	15.00	15.00	15.00
PM20J	PYRENE	ug/L	1	1	275	0.4	1	37	7.51	1.26	32.20	32.20	25.00
X300O	2-CHLOROPHENOL	ug/L	0	1	275	0.4	1	37	12.53	1.26	30.50	30.50	25.00
X3246	2,4,6-TRICHLOROPHENOL	ug/L	1	1	275	0.4	1	37	12.53	1.26	30.50	30.50	25.00
DIOXINS AND FURANS													
P98CDD	OCTACHLORODIBENZO(DIOXIN)	ng/L	1	4	54	7.4	4	37	0.87	3.66	26.00	0.73	1.00
P94CDD	TETRACHLORODIBENZO(FURAN)	ng/L	2	1	54	1.9	1	37	0.41	3.08	4.80	4.80	0.20
P98CDD	OCTACHLORODIBENZO(FURAN)	ng/L	1	1	54	1.9	1	37	0.79	2.87	3.00	3.00	1.00
PESTICIDES, HERBICIDES, PCB'S													
P72AD	2,4-DICHLOROPHENOXYACETIC ACID	ug/L	3	214	276	77.5	37	37	100.0	3.72	2.60	0.90	0.04
P72BC	GAMMA-BHC(DICHLOROCYCLOHEXANE)	ug/L	2	143	276	51.8	35	37	94.6	0.02	2.29	0.01	0.02
P72BD	METHOXYCYCLO	ug/L	1	47	276	17.0	16	37	0.08	3.14	5.90	0.10	0.10
P72BT	PCB, TOTAL	ug/L	2	42	276	15.2	17	37	0.06	2.31	4.50	0.08	0.08
X2124	1,2,4-TRICHLOROBENZENE	ug/L	3	35	276	12.7	15	37	0.01	2.48	2.26	0.02	0.02
P72B	1,2,4-TRICHLOROBENZENE	ug/L	1	32	276	10.1	15	37	0.01	2.48	2.26	0.02	0.02
P72B	SILVEX	ug/L	3	28	276	8.0	14	37	0.06	1.08	1.00	0.08	0.10
P72AT	2,4,5-TRICHLOROPHENOXYACETIC ACID	ug/L	3	22	276	8.0	14	37	0.06	1.60	0.73	0.07	0.10
PM20A	ALPHA-BHC(DICHLOROCYCLOHEXANE)	ug/L	1	16	276	5.8	7	37	0.01	1.50	0.10	0.02	0.02
PM20B	PM20B	ug/L	1	15	276	5.4	9	37	0.01	1.74	5.80	0.01	0.02
PM20C	ENDOSULFAN SULPHATE	ug/L	3	13	276	4.7	6	37	0.04	1.56	0.99	0.12	0.08
PM20D	HEXACHLOROBENZENE	ug/L	2	11	276	4.0	7	37	0.01	1.39	0.10	0.02	0.02
PM20E	ENDOSULFAN II	ug/L	3	10	276	3.6	8	37	0.01	1.42	0.15	0.02	0.02
PM20F	ENDOSULFAN II	ug/L	3	10	276	3.6	8	37	0.01	1.42	0.15	0.02	0.02
PM20G	GAMMA-CYCLODIAN	ug/L	1	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20H	PM20H	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20I	PM20I	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20J	PM20J	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20K	PM20K	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20L	PM20L	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20M	PM20M	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20N	PM20N	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20O	PM20O	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20P	PM20P	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20Q	PM20Q	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20R	PM20R	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20S	PM20S	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20T	PM20T	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20U	PM20U	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20V	PM20V	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20W	PM20W	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20X	PM20X	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20Y	PM20Y	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20Z	PM20Z	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20A	PM20A	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20B	PM20B	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20C	PM20C	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20D	PM20D	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20E	PM20E	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20F	PM20F	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20G	PM20G	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20H	PM20H	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20I	PM20I	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20J	PM20J	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20K	PM20K	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20L	PM20L	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20M	PM20M	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20N	PM20N	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20O	PM20O	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20P	PM20P	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20Q	PM20Q	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20R	PM20R	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20S	PM20S	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20T	PM20T	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20U	PM20U	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20V	PM20V	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20W	PM20W	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20X	PM20X	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20Y	PM20Y	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20Z	PM20Z	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20A	PM20A	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20B	PM20B	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20C	PM20C	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20D	PM20D	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20E	PM20E	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20F	PM20F	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20G	PM20G	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20H	PM20H	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20I	PM20I	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20J	PM20J	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20K	PM20K	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20L	PM20L	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20M	PM20M	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20N	PM20N	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20O	PM20O	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20P	PM20P	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20Q	PM20Q	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20R	PM20R	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20S	PM20S	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20T	PM20T	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20U	PM20U	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20V	PM20V	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20W	PM20W	ug/L	3	9	276	3.3	6	37	0.01	1.49	2.10	0.02	0.02
PM20X	PM20X	ug/L	3	9	276	3.3	6	37	0.01	1.49			

TABLE 5-7b - GLOBAL SUMMARY OF CONTAMINANTS IN RAW SEWAGE

CONTAM- INANT	CONTAMINANT NAME	UNITS	QA/QC CODE	GLOBAL # SAMP. DET.	GLOBAL # SAMP. TESTED	GLOBAL % FREQ. DET.	GLOBAL # PLANTS DET.	GLOBAL # PLANTS	GLOBAL % PLANT PREV.	GLOBAL MEAN	GLOBAL SPREAD FACTOR	GLOBAL MAX. CONC.	GLOBAL MIN. CONC. S.D.	DET. LIMIT (ML)
VOLATILES														
B2MPX	M, AND P-XYLENES	ug/L	1	43	274	15.7	14	37	37.8	26.00	2.02	1700.00	3.80	40.00
B2BNZ	ETHYL BENZENE	ug/L	1	30	274	11.0	11	37	29.7	23.50	1.75	1200.00	3.50	40.00
X12HLO	CHLOROFORM	ug/L	1	28	274	10.2	12	37	32.4	23.90	1.75	340.00	41.00	40.00
ROXYL	O-XYLENE	ug/L	1	25	274	9.1	9	37	24.3	22.47	1.56	570.00	4.40	40.00
X12HCL	ORCUTANE	ug/L	1	21	274	7.7	7	37	18.9	22.18	1.74	440.00	43.42	40.00
X12HCL	THCHLOROTOLUENE	ug/L	1	11	274	4.0	4	37	10.2	21.59	1.82	980.00	72.00	40.00
X12HCL	TETRACHLOROTUENE	ug/L	1	12	274	4.4	7	37	10.8	21.59	1.82	300.00	43.42	40.00
B2STVR	STYRENE	ug/L	1	9	274	3.3	4	37	10.8	21.40	1.39	120.00	43.33	40.00
X12DCL	1,1-DICHLOROTUENE	ug/L	1	7	274	2.6	5	37	13.5	20.55	1.26	220.00	10.00	40.00
B2HOCCL	BROMODICHLOROBENZENE	ug/L	1	2	274	0.7	2	37	5.4	20.55	1.12	55.00	40.00	40.00
X12DCE	1,2-DICHLOROTUENE	ug/L	1	2	274	0.7	2	37	5.4	20.26	1.17	120.00	120.00	40.00
X12DCM	BROMODICHLOROMETHANE	ug/L	1	2	274	0.7	2	37	5.4	30.10	1.10	130.00	18.00	60.00
X12HCL	CHLOROMETHANE	ug/L	1	2	274	0.7	2	37	5.4	20.16	1.10	67.00	56.00	40.00
X12HCL	CARBON TETRACHLORIDE	ug/L	1	2	274	0.4	1	37	2.7	18.49	7.12	99.00	57.00	40.00
X12HCL	ETHANOL	ug/L	2	1	274	0.4	1	37	2.7	18.49	7.12	520.00	200.00	50.00
X111CE	1,1-DICHLOROTUENE	ug/L	1	1	274	0.4	1	37	2.7	20.19	1.10	250.00	250.00	40.00
X112CP	1,2-DICHLOROPROPANE	ug/L	1	1	274	0.4	1	37	2.7	20.05	1.05	42.00	42.00	40.00
X113DP	CIS-1,3-DICHLOROPROPANE	ug/L	1	1	274	0.4	1	37	2.7	29.24	1.11	40.00	40.00	60.00
X12ACRO	ACROLEIN	ug/L	3	1	274	0.4	1	37	2.7	202.81	1.26	9200.00	9200.00	400.00
X12DCE	CIS-1,2-DICHLOROTUENE	ug/L	1	1	274	0.4	1	37	2.7	20.14	1.12	130.00	130.00	40.00
X12DCE	TRANS-1,2-DICHLOROTUENE	ug/L	1	1	274	0.4	1	37	2.7	20.15	1.05	42.00	42.00	40.00
X22CTO	3-CHLOROTOLUENE	ug/L	1	1	274	0.4	1	37	2.7	20.51	1.11	44.00	44.00	40.00

TABLE 5-8 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN PRIMARY EFFLUENTS

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
ASUT	<u>METALS AND CYANIDE</u>		
BEUT	ARSENIC UNFLT TOTAL		
	BERYLLIUM UNFLT TOTAL		
	<u>BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS</u>		
P2AMET	AMETRYNE	PMPCRE	P-CRESOL*
P2ATRA	ATRAZINE*		
P4DIAZ	DIAZINON*		
P4EPAR	PARATHION ETHYL*		
P4MALA	MALATHION*		
P4MPAR	METHYL PARATHION*		
PM24DP	2,4-DICHLOROPHENOL		
PM24DT	2,4-DINITROTOLUENE		
PM24MP	2,4-DIMETHYLPHENOL		
PM24NP	2,4-DINITROPHENOL*		
PM26DT	2,6-DINITROTOLUENE		
PM2NP	2-NITROPHENOL		
PM46DP	2-METHYL 4,6-DINITROPHENOL		
PM4BPE	4-BROMOPHENYLPHENYLETHYR		
PM4CPE	4-CHLOROPHENYLPHENYLETHYR		
PM4NP	4-NITROPHENOL		
PMANAA	ALPHA-NAPHTHYLAMINE*		
PMB2EM	BIS(2-CHLOROETHOXY)METHANE		
PMB2IE	BIS(2-CHLORISOPROPYL)ETHER		
PMB2NE	BIS-(2-CHLOROMETHYL)ETHER		
PMBNAA	BETA-NAPHTHYLAMINE*		
PMDMP	DIMETHYL PHTHALATE		
PMDPPE	DIPHENYL ETHER		
PMNTB	NITROBENZENE		
PMNND	N-NITROSO-DI-PHENYLAMINE		
PMNNP	N-NITROSO-DI-N-PROPYLAMINE		
PMPCCMC	P-CHLORO-M-CRESOL		
PN2CNA	CHLORONAPHTHALENE*		
PNACNE	ACENAPHTHENE*		
PNACNY	ACENAPHTHYLENE		
PNANTH	ANTHRACENE		
PNBAA	BENZOA)ANTHRACENE		
PNBAP	BENZOA)PYRENE		
PNBBFA	BENZOKB)FLUORANTHENE		
PNBPH	BIPHENYL		
PNBKF	BENZOKK)FLUORANTHENE		
PNCHRY	CHRYSENE		
PNDAAH	DIBENZOA)ANTHRACENE*		
PNFLAN	FLUORANTHENE		
PNGHIP	BENZOKJ)FLUORENE*		
PNINP	IDENOK1,2,3-CD)PYRENE*		
PNPHEN	PHENANTHRENE		
PNPYR	PYRENE		
PODICH	DICHLORAN*		
POTOC	TRI-O-CRESYL PHOSPHATE*		
X300O	2-CHLOROPHENOL		
X324S	2,4,5-TRICHLOROPHENOL*		
X3246	2,4,6-TRICHLOROPHENOL		
X3PC7H	PENTACHLOROPHENOL		
	<u>DIOXINS AND FURANS</u>		
P94CDD	TETRACHLORODIBENZODIOXIN*		
P95CDD	PENTACHLORODIBENZODIOXIN		
P95CDF	PENTACHLORODIBENZOFURAN		
P96CDD	HEXACHLORODIBENZODIOXIN		
P96CDF	HEXACHLORODIBENZOFURAN		
P97CDD	HEPTACHLORODIBENZODIOXIN		
P97CDF	HEPTACHLORODIBENZOFURAN		
P98CDF	OCTACHLORODIBENZOFURAN		
	<u>PESTICIDES, HERBICIDES, PCBs</u>		
POPCNB	PCNB	POCAPN	CAPTAN*
P1ALDR	ALDRIN	P1END1	ENDOSULFAN I
P1BHCD	DELTA-BHC(HEXACHLOROCYCLOHEXANE)	P1ENDR	ENDRIN
P1CHLA	ALPHA-CHLORDANE	X1HCBD	HEXACHLOROBUTADIENE*
P1CHLG	GAMMA-CHLORDANE		
P1DIEL	DELDRIN		
P1ENDA	ELDRIN ALDEHYDE		
P1HEPE	HEPTACHLORPOXIDE		
P1OCHL	OXYCHLORDANE		
P1PMIR	MIREX PHOTO		

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

TABLE 5-8 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN PRIMARY EFFLUENTS

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
P1PPDD P1PPDE P1STRO P1TOX X2HCB	PESTICIDES, HERBICIDES, PCBS PP-DDD PP-DDE STROBANE* TOXAPHENE HEXACHLOROBENZENE VOLATILES ORGANIC		
B1VBR B2BDCL E1DIEE L1HEX PM2CEE X111Z2 X111ZT X111CE X112CE X112CP X113DP X113DR X1ACTO X1BETH X1BROM X1CDBM X1CDCE X1CHLE X1CHLM X1DCFM X1T12D X1TCFM X1VCL X212CB X213CB X214CB X23CTO X2CBEN X2CPPE	VINYL BROMIDE* BROMODICHLOROBENZENE DIETHYL ETHER HEXANOL 2-CHLOROETHYL VINYLETHER* 1,1,2,2-TETRACHLOROETHANE* 1,1,2-TRICHLOROETHANE* 1,1-DICHLOROETHANE 1,2-DICHLOROETHANE 1,2-DICHLOROPROPANE CIS-1,3-DICHLOROPROPENE TRANS-1,3-DICHLOROPROPENE ALPHA-CHLOROTOLUENE BROMOETHANE* BROMOFORM* CHLORODIBROMOMETHANE CIS-1,2-DICHLOROETHYLENE CHLOROETHANE* CHLOROMETHANE* DICHLORODIFLUOROMETHANE TRI-1,2-DICHLOROETHYLENE* TRICHLOROFLUOROMETHANE* VINYL CHLORIDE* 1,2-DICHLOROBENZENE 1,3-DICHLOROBENZENE 1,4-DICHLOROBENZENE 3-CHLOROTOLUENE CHLOROBENZENE 3-CHLOROPROPENE*	X1ACRO X1ACRY	ACROLEIN ACRYLONITRILE*

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

TABLE 5-B - GLOBAL SUMMARY OF CONTAMINANTS IN PRIMARY EFFLUENTS

CONTAM. INANT	CONTAMINANT NAME	UNITS	QA/QC CODE	GLOBAL # SAMP.	GLOBAL # SAMP. TESTED	GLOBAL % SAMP. DET.	GLOBAL # PLANTS DET.	GLOBAL # PLANTS	GLOBAL % PLANT PREV.	GLOBAL GEO. MEAN	GLOBAL SPREAD FACTOR	GLOBAL MAX. CONC.	GLOBAL MIN. CONC. > DL	DET. LIMIT (DL)
CONVENTIONAL														
BOD5	BOD, 5 DAY - TOTAL DEMAND	mg/L	0	40	40	100.0	7	7	100.0	48.40	2.05	139.00	8.20	1.00
COD	CHEMICAL OXYGEN DEMAND	mg/L	0	40	40	100.0	7	7	100.0	18.40	1.36	376.00	36.00	1.00
DOC	DISSOLVED ORGANIC CARBON	mg/L	0	40	40	100.0	7	7	100.0	12.90	1.66	21.60	2.20	0.20
TOC	TOTAL ORGANIC CARBON	mg/L	0	40	40	100.0	7	7	100.0	10.46	1.30	24.10	8.30	0.01
TPH	NITROGEN TOTAL NITROGEN	mg/L	0	40	40	100.0	7	7	100.0	15.36	1.03	7.44	6.16	1.00
PH	(GLOCH)(CONC)	mg/L	0	40	40	100.0	7	7	100.0	6.88	1.03	7.44	6.16	1.00
PHIT	PHOSPHORUS UNFILT TOTAL	mg/L	0	34	34	100.0	5	5	100.0	1.34	1.96	4.02	0.33	0.01
RSP	RESIDUE PARTICULATE	mg/L	0	39	39	100.0	7	7	100.0	29.57	1.73	111.00	7.30	1.00
RSLOI	RESIDUE PARLOSS ON IGN	mg/L	0	10	10	100.0	2	2	100.0	28.16	1.95	76.80	12.80	0.00
TURB	TURBIDITY	mg/L	0	5	5	100.0	1	1	100.0	1.88	1.35	16.00	1.00	0.10
PHOL	PHENOLICS (4A&P)	mg/L	0	7	40	100.0	7	7	100.0	0.08	2.01	2.60	0.11	0.10
PHOZP	PHOSPHATE	mg/L	0	4	40	100.0	1	7	14.3	0.00	4.03	0.53	0.15	0.01
NHONTR	NITRATE, TOTAL FILT REAC	mg/L	0	4	40	100.0	1	7	14.3	0.05	3.16	1.55	0.90	0.05
METALS AND CYANIDE														
ZNIT	ZINC UNFILT TOTAL	ug/L	0	48	48	100.0	7	7	100.0	69.80	3.04	1360.00	10.00	20.00
ZNIT	ZINC UNFILT TOTAL	ug/L	0	43	48	97.9	7	7	100.0	304.90	2.81	1010.00	60.00	10.00
ALUT	ALUMINUM UNFILT TOTAL	ug/L	0	38	39	97.4	7	7	100.0	0.05	2.42	0.36	0.01	0.01
ALUT	ALUMINUM UNFILT TOTAL	ug/L	0	46	48	95.8	7	7	100.0	550.00	3.45	4860.00	100.00	20.00
CUIT	COPPER UNFILT TOTAL	ug/L	0	7	8	87.5	7	7	100.0	18.20	1.85	60.00	10.00	10.00
CRUT	CHROMIUM UNFILT TOTAL	ug/L	0	29	48	60.4	6	7	85.7	10.80	1.94	40.00	10.00	10.00
CDUT	CADMIUM UNFILT TOTAL	ug/L	0	18	48	37.5	6	7	85.7	2.50	1.96	7.00	3.00	3.00
COIT	COBALT UNFILT TOTAL	ug/L	0	11	48	22.9	5	7	71.4	6.60	1.66	20.00	10.00	10.00
NIUT	NICKEL UNFILT TOTAL	ug/L	0	10	48	20.8	4	7	57.1	8.70	2.86	140.00	10.00	10.00
PHIT	LEAD UNFILT TOTAL	ug/L	0	9	48	18.8	5	7	71.4	20.80	1.94	140.00	30.00	30.00
AGUT	SILVER UNFILT TOTAL	ug/L	0	8	48	16.7	4	7	57.1	6.40	1.61	20.00	10.00	10.00
CCNTR	CYANIDE-FRE UNFILT REAC	ug/L	0	3	40	7.5	3	7	42.9	0.90	2.61	39.00	7.00	1.00
SEUT	SELENIUM UNFILT TOTAL	ug/L	0	1	45	2.2	1	7	14.3	16.50	1.70	30.00	30.00	30.00
BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS														
PMCSRE	M-CRESOL	ug/L	1	18	39	46.2	5	7	71.4	3.90	2.97	32.40	4.30	3.00
PMBBP	BUTYL-BENZYLPHENATE	ug/L	1	8	39	20.5	4	7	57.1	1.42	2.10	9.20	2.10	2.00
PMBPN	PHENOL	ug/L	2	5	39	12.8	3	7	42.9	1.78	1.60	8.70	3.40	2.00
PNAPLE	NAPHTHALENE	ug/L	1	5	39	15.8	3	7	42.9	1.31	1.43	14.00	2.00	2.00
PMFLU	FLUORENE	ug/L	2	2	39	5.1	1	7	14.3	1.60	1.33	7.20	3.80	3.00
PNFLU	FLUORENE	ug/L	1	1	39	2.6	1	7	14.3	1.05	1.38	7.60	7.60	2.00
DIOXINS AND FURANS														
PSKCD	OCTACHLORODIBENZODIEN	ug/L	1	2	8	25.0	2	7	28.6	0.25	3.33	1.40	0.08	0.30
PSKCDP	TETRACHLORODIBENZOFURAN	ug/L	2	1	8	12.5	1	7	14.3	0.09	2.20	0.10	0.10	0.10
PESTICIDES, HERBICIDES, FUNGICIDES														
PHICG	GAMMA-BHC(HCH, CYCLOHEXANH)	ug/L	2	29	40	72.5	6	7	85.7	0.02	2.28	0.09	0.01	0.01

TABLE 5.8 b--GLOBAL SUMMARY OF CONTAMINANTS IN PRIMARY EFFLUENTS

CONTAM- INANT	CONTAMINANT NAME	UNITS	QC CODE	GLOBAL # SAMP.	GLOBAL # SAMP. TESTED	GLOBAL % SAMP.	GLOBAL % FREQ.	GLOBAL # PLANTS DET.	GLOBAL # PLANTS	GLOBAL % PREV.	GLOBAL MEAN	GLOBAL SPREAD FACTOR	GLOBAL MAX. CONC.	GLOBAL MIN. CONC. > DL	DET. LIMIT (DL)
PESTICIDES, HERBICIDES, PCBs															
P24D	2,4-DICHLOROPHENOXYACETIC ACID	ug/L	3	27	40	67.5	0.06	6	7	85.7	0.06	4.96	1.70	0.02	0.02
P51LV	50 VEX	ug/L	3	9	40	22.5	0.03	4	7	57.1	0.03	2.19	1.40	0.05	0.05
P1PCBT	PCB TOTAL	ug/L	2	7	40	17.5	0.03	2	7	28.6	0.03	3.06	0.43	0.05	0.04
P24ST	2,4,5-TRICHLOROPHENOXYACETIC ACID	ug/L	3	7	40	17.5	0.04	3	7	42.9	0.04	2.61	2.80	0.06	0.05
P1BHC	BETA-BHC (DIECHLOROCYCLOHEXANE)	ug/L	1	6	40	15.0	0.01	2	7	28.6	0.01	1.36	0.02	0.01	0.01
P1BHC	BETA-BHC (DIECHLOROCYCLOHEXANE)	ug/L	1	5	40	12.5	0.01	1	7	14.3	0.01	1.64	0.05	0.02	0.01
P1BHC	ALPHA-BHC (DIECHLOROCYCLOHEXANE)	ug/L	1	2	40	5.0	0.01	1	7	14.3	0.01	1.12	0.01	0.01	0.01
P1ENDS	ENDOSULFAN II	ug/L	3	1	40	2.5	0.01	1	7	14.3	0.01	1.12	0.01	0.01	0.01
P1ENDS	ENDOSULFAN SULPHATE	ug/L	3	1	40	2.5	0.02	1	7	14.3	0.02	1.36	0.14	0.14	0.04
P1MEX	HEPTACHLOR	ug/L	3	1	40	2.5	0.01	1	7	14.3	0.01	1.25	0.02	0.02	0.01
P1MEX	MIREX	ug/L	1	1	40	2.5	0.01	1	7	14.3	0.01	1.39	0.04	0.04	0.01
P1DDT	DDT	ug/L	3*	1	40	2.5	0.02	1	7	14.3	0.02	1.31	0.11	0.11	0.04
X1DCP	1,2-DICHLOROCYCLOPENTADIENE	ug/L	3	1	40	2.5	0.05	1	7	14.3	0.05	1.03	0.06	0.06	0.10
X124	1,2,4-TRICHLOROBENZENE	ug/L	3	1	40	2.5	0.01	1	7	14.3	0.01	1.33	0.03	0.03	0.01
VOLATILES ORGANIC COMPOUNDS															
X1TTR	TETRACHLOROETHYLENE	ug/L	1	21	38	55.3	4.39	6	7	85.7	4.39	5.84	380.00	2.20	2.00
X1TTR	PERCHLOROETHYLENE	ug/L	1	13	38	34.2	1.94	6	7	85.7	1.94	3.05	39.00	2.70	2.00
B1MEXY	M-XYLENES	ug/L	1	9	38	26.3	2.84	5	7	71.4	2.84	3.42	83.00	2.20	2.00
X111T	1,1,1-TRICHLOROETHANE	ug/L	1	10	38	26.3	2.34	2	7	28.6	2.34	1.97	16.00	1.60	2.00
X1C1T	CARBON TETRACHLORIDE	ug/L	1	9	38	23.7	2.05	3	7	42.9	2.05	3.84	53.00	4.60	2.00
X1TTR	TRICHLOROETHYLENE	ug/L	1	7	38	18.4	1.71	3	7	42.9	1.71	3.60	420.00	6.40	2.00
B1BENZ	BENZENE	ug/L	1	5	38	13.2	1.29	2	7	28.6	1.29	2.03	17.00	2.20	2.00
X1DC1E	1,1-DICHLOROETHYLENE	ug/L	1	3	38	7.9	1.43	1	7	14.3	1.43	3.44	110.00	70.00	2.00
B1OC1E	1-OCTENE	ug/L	1	1	38	2.6	1.36	1	7	14.3	1.36	1.36	10.00	10.00	3.00
B1STTR	STYRENE	ug/L	1	1	38	2.6	1.45	1	7	14.3	1.39	1.45	15.00	15.00	3.00

5.3.5 Summary of Contaminants in Lagoon Effluents

Table 5-9(a) presents the contaminants that were not detected in any effluent sample from either of the two lagoons sampled. In total, 133 contaminants were not detected, including 12 that were not confirmed. Also indicated in the Table are these 34 contaminants not detected in any sample at any WPCP.

Table 5-9(b) shows that in the lagoon effluents, only 7 organic compounds were detected, all from the pesticide/herbicide group. Only one herbicide (2,4-Dichlorophenoxyacetic acid) was detected at both lagoons. Of the 10 metals detected, 9 were detected at both lagoons.

Only 3 organic compounds (2,4-Dichlorophenoxyacetic acid, Methoxychlor and 1,2,4-Trichlorobenzene) were detected in more than 1 or 10 percent of the samples. Four metals (Al, Hg, Sr and Zn) were detected in more than 90 percent of the samples, while the other metals (Cd, Co, Mo, Ni, Cr, Cu) were detected in at least 17 percent of the samples.

5.3.6 Summary of Contaminants in Secondary Effluents

Table 5-10(a) presents the contaminants that were not detected in any effluent sample from any of the 28 secondary WPCPs. A total of 74 contaminants were never detected including 5 that were not confirmed. Also indicated in the Table are the 34 compounds never detected in any type of sample at any WPCP.

Data regarding contaminants detected in secondary WPCP effluents is presented in Table 5-10(b). Sixty-eight organic compounds, 14 metals and cyanide were detected in at least one secondary effluent sample. However, none of the base-neutral and acid extractable compounds or dioxins and furans were detected at more than 15 percent of the WPCP (4 plants).

Only 8 of the 23 compounds in the pesticide/herbicide group and 3 of the 17 volatile organic compounds were detected at more than 15 percent of the WPCPs. Metals were the most prevalent contaminants, with 11 metals detected at more than 50 percent of the WPCPs.

As noted, the most frequently detected contaminant group was metals, with 7 metals detected in greater than 50 percent of the secondary effluent samples. The most frequently detected base neutral and acid extractable compounds were found in less than 4 percent of samples; dioxin/furan compounds were found in less than 9 percent of samples, and volatile organic compounds were found in less than 10 percent of samples. Two pesticide/herbicide compounds (2,4-Dichlorophenoxyacetic and gamma-BHC) were detected in at least 70 percent of all the final effluent samples.

TABLE 5-9 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN LAGOON EFFLUENTS

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
AGUT ASUT BELT CCNFUR PBUT SEUT	<p><u>METALS AND CYANIDE</u></p> <p>SILVER,UNFLT.TOTAL ARSENIC,UNFLT.TOTAL BERYLLIUM,UNFLT.TOTAL CYANIDE-FREE,UNFLT.REAC. LEAD,UNFLT.TOTAL SELENIUM,UNFLT.TOTAL</p> <p><u>BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS</u></p>		
P2AMET P2ATRA P4DIAZ P4EPAR P4MALA P4MPAR P4M4DP P4M4DT P4M4MP P4M4NP P4M6DT P4M2NP P4M6DP P4M4PE P4M4NP P4MANAA P4MB2EM P4MB2IE P4MB2NE P4MBBP P4MBNAA P4MDMP P4MDPE P4MOCRE P4MNTTB P4M4ND P4M4NP P4MOCRE P4MPCMC P4MPCHE P4M2CNA P4NACNE P4NACNY P4NANTH P4NBAA P4NBAP P4NBIFA P4NBIPH P4NBKF P4NBRY P4NDAHA P4NFLAN P4NFLUO P4NGHP P4NDP P4NNAPH P4NPHEN P4NPYR P4ODICH P4OTOC X3301O X3345 X3346 X33PCFH	<p>AMETRYNE ATRAZINE* DIAZINON* PARATHION ETHYL* MALATHION* METHYL PARATHION* 2,4-DICHLOROPHENOL 2,4-DINITROTOLUENE 2,4-DIMETHYLPHENOL 2,4-DINITROPHENOL* 2,6-DINITROTOLUENE 2-NITROPHENOL 2-METHYL 4,6-DINITROPHENOL 4-BROMOPHENYLPHENYLETHER 4-CHLOROPHENYLPHENYLETHER 4-NITROPHENOL ALPHA-NAPHTHYLAMINE* BIS(2-CHLOROETHOXY)METHANE BIS(2-CHLOROISOPROPYL)ETHER BIS(2-CHLOROMETHYL)ETHER BUTYLBENZYLPHthalate BETA-NAPHTHYLAMINE* DIMETHYL PHthalate DIPHENYL ETHER M-CRESOL NITROBENZENE N-NITROSO-DI-PHENYLAMINE N-NITROSO-DI-N-PROPYLAMINE O-CRESOL P-CHLORO-M-CRESOL PHENOL CHLORONAPHTHALENE* ACENAPHTHENE* ACENAPHTHYLENE ANTHRACENE BENZOX(A)ANTHRACENE BENZOX(A)PYRENE BENZOX(B)FLUORANTHENE BIPHENYL BENZOX(K)FLUORANTHENE CHRYSENE DIBENZOX(A,H)ANTHRACENE* FLUORANTHENE FLUORENE BENZOX(G,H)PERYLENE* IDENOX(1,2,3-CD)PYRENE* NAPHTHALENE PHENANTHRENE PYRENE DICHLORAN* TRI-O-CRESYL PHOSPHATE* 2-CHLOROPHENOL 2,4,5-TRICHLOROPHENOL* 2,4,6-TRICHLOROPHENOL PENTACHLOROPHENOL</p> <p><u>DIOXINS AND FURANS</u></p> <p>TETRACHLORODIBENZODIOXIN* TETRACHLORODIBENZOFURAN PENTACHLORODIBENZODIOXIN PENTACHLORODIBENZOFURAN HEXACHLORODIBENZODIOXIN HEXACHLORODIBENZOFURAN HEPTACHLORODIBENZODIOXIN HEPTACHLORODIBENZOFURAN OCTACHLORODIBENZODIOXIN OCTACHLORODIBENZOFURAN</p>	P4MPCRE	P-CRESOL*

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

TABLE 5-9 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN LAGOON EFFLUENTS

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
	<u>PESTICIDES, HERBICIDES, PCBs</u>		
POPCNB	PCNB	POCAPN	CAPTAN*
PIALDR	ALDRIN	PIENDI	ENDOSULFAN I
PIBHCA	ALPHA-BHCHEXCHLORCYCLHEXANE)	PIEND2	ENDOSULFAN II
PIBHCB	BETA-BHC (HEXCHLORCYCLHEXANE)	PIENDR	ENDRIN
PIBHCD	DELTA-BHCHEXCHLORCYCLHEXANE)	PIPPDT	PP-DDT
PICHLA	ALPHA-CHLORDANE	P324ST	2,4,5-TRICHLORPHENOXYACETIC ACID
PICHLG	GAMMA-CHLORDANE	P35ILV	SILVEX
PIDIEL	DIELDRIN	X1HCBD	HEXACHLOROBLTADIENE*
PIENDA	ELDRIN ALDEHYDE	X2HCE	HEXACHLOROETHANE
PIHEPE	HEPTACHLOREPOXID		
PIMIRX	MIREX		
PIOCGL	OXYCHLORDANE		
PIPCBT	PCB, TOTAL		
PIPMIR	MIREX PHOTO		
PIPPDD	PP-DDD		
PIPPDE	PP-DDE		
PISTRO	STROBANE*		
PI TOX	TOXAPHENE		
X2HCB	HEXACHLOROBENZENE		
	<u>VOLATILES</u>		
B1OCTE	1-OCTENE	X1ACRO	ACROLEIN
B1VBR	VINYL BROMIDE*	X1ACRY	ACRYLONITRILE*
B2BDCL	BROMODICHLOROBENZENE		
B2EBNZ	ETHYLBENZENE		
B2MPXY	M- AND P-XYLENES		
B2OXYL	O-XYLENE		
B2STYR	STYRENE		
E1DIEE	DIETHYL ETHER		
L1HEX	HEXANOL		
PM2CEE	2-CHLOROETHYLVINYLETHER*		
X1111T	1,1,1-TRICHLOROETHANE		
X11122	1,1,2,2-TETRACHLOROETHANE*		
X1112T	1,1,2-TRICHLOROETHANE*		
X111CE	1,1-DICHLOROETHANE		
X112CE	1,2-DICHLOROETHANE		
X112CP	1,2-DICHLOROPROPANE		
X113DP	CIS-1,3-DICHLOROPROPENE		
X113DR	TRANS-1,3-DICHLOROPROPENE		
X1ACTO	ALPHA-CHLOROTOLUENE		
X1BDCM	BROMODICHLOROMETHANE		
X1BETH	BROMOETHANE*		
X1BROM	BROMOFORM*		
X1CDBM	CHLORODIBROMOMETHANE		
X1CDCE	CIS-1,2-DICHLOROETHYLENE		
X1CHLE	CHLOROETHANE*		
X1CHLM	CHLOROMETHANE*		
X1CHLO	CHLOROFORM		
X1CTET	CARBON TETRACHLORIDE		
X1DCFM	DICHLORODIFLUOROMETHANE		
X1DGLE	1,1-DICHLOROETHENE		
X1T12D	TRI-1,2-DICHLOROETHYLENE*		
X1TCFM	TRICHLORODIFLUOROMETHANE*		
X1TETR	TETRACHLOROETHYLENE		
X1TRIC	TRICHLOROETHYLENE		
X1VCL	VINYL CHLORIDE*		
X212CB	1,2-DICHLOROBENZENE		
X213CB	1,3-DICHLOROBENZENE		
X214CB	1,4-DICHLOROBENZENE		
X23CTO	3-CHLOROTOLUENE		
X2CBEN	CHLOROBENZENE		
X2CPPE	3-CHLOROPROPENE*		

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

TABLE 5-9b - GLOBAL SUMMARY OF CONTAMINANTS IN LAGOON EFFLUENTS

CONTAM- INANT	CONTAMINANT NAME	UNITS	QA/QC CODE	GLOBAL # SAMP. DET.	GLOBAL % SAMP. TESTED	GLOBAL % FREQ. DET.	GLOBAL # PLANTS DET.	GLOBAL % PLANT PREV.	GLOBAL GEO. MEAN	GLOBAL SPREAD FACTOR	GLOBAL MAX. CONC.	GLOBAL MIN. CONC. > DL	DET. LIMIT (DL)
CONVENTIONAL													
BOD5	BOD 5 DAY - TOTAL DEMAND	mg/L	0	10	10	100.0	2	100.0	24.95	1.66	48.20	14.40	1.00
COD	CHEMICAL OXYGEN DEMAND	mg/L	0	10	10	100.0	2	100.0	70.64	1.67	110.00	4.00	5.00
DOC	DISSOLVED ORGANIC CARBON	mg/L	0	10	10	100.0	2	100.0	10.09	1.14	11.60	8.30	1.00
NO3-PR	NITRATE-FILT REACT.	mg/L	0	10	10	100.0	2	100.0	0.42	4.77	3.62	0.11	0.01
NO3-NR	NITRATE-TOTAL FILT REAC.	mg/L	0	10	10	100.0	2	100.0	1.65	3.25	6.15	0.40	0.05
NH4-NR	NITROGEN-TOT KIEL UNF-TOT	mg/L	0	10	10	100.0	2	100.0	10.85	1.64	43.00	8.30	0.01
PHOS	PHOSPHORUS UNFILT-TOTAL	mg/L	0	10	10	100.0	2	100.0	7.92	1.03	8.38	7.57	1.00
PRPT	PHOSPHORUS PARTICULATE	mg/L	0	6	6	100.0	1	100.0	0.226	1.18	0.31	0.20	0.01
RSPT	RESIDUE PARTICULATE	mg/L	0	10	10	100.0	2	100.0	29.40	1.18	35.00	19.40	1.00
RSPT-01	RESIDUE PARTICLOSS ON IONE	mg/L	0	2	2	100.0	1	100.0	40.98	1.04	42.20	39.40	0.01
TURB	TURBIDITY	mg/L	0	5	5	100.0	1	100.0	25.76	1.20	30.00	29.40	0.25
NH4-NR	AMMONIUM-TOTAL FILT REAC.	mg/L	0	6	10	60.0	2	100.0	0.60	12.80	6.35	0.65	0.20
PHENOL	PHENOLICS (AAP)	mg/L	0	2	10	20.0	1	50.0	0.07	1.91	0.34	0.13	0.10
METALS AND CYANIDE													
ALUT	ALUMINUM UNFILT-TOTAL	ug/L	0	12	12	100.0	2	100.0	171.20	1.31	240.00	100.00	20.00
IGUT	MERCURY UNFILT-TOTAL	ug/L	0	10	10	100.0	2	100.0	0.01	1.25	0.02	0.01	0.01
SRUT	STRONTIUM UNFILT-TOTAL	ug/L	0	12	12	100.0	2	100.0	230.70	1.36	330.00	160.00	10.00
ZNUT	ZINC UNFILT-TOTAL	ug/L	0	11	12	91.7	2	100.0	11.90	1.37	20.00	10.00	10.00
CUUT	COPPER UNFILT-TOTAL	ug/L	0	1	2	50.0	1	50.0	10.00	0.00	10.00	10.00	10.00
COUT	CADMIUM UNFILT-TOTAL	ug/L	0	3	12	25.0	2	100.0	1.80	1.49	4.00	3.00	3.00
COBT	COBALT UNFILT-TOTAL	ug/L	0	3	12	25.0	2	100.0	6.30	1.41	10.00	10.00	10.00
MOUT	MOLYBDENUM UNFILT-TOTAL	ug/L	0	3	12	25.0	2	100.0	3.70	1.37	10.00	10.00	10.00
NIUT	NICKEL UNFILT-TOTAL	ug/L	0	3	12	25.0	2	100.0	6.90	1.74	30.00	10.00	10.00
CRUT	CHROMIUM UNFILT-TOTAL	ug/L	0	2	12	16.7	1	50.0	5.90	1.37	10.00	10.00	10.00
PESTICIDES, HERBICIDES, INTRIS													
P2AD	2,4-DICHLOROPHENOXYACETIC ACID	ug/L	3	7	10	70.0	2	100.0	0.03	2.08	0.10	0.03	0.02
P2MDT	METHOXYCIBOZOL	ug/L	1	4	10	40.0	1	50.0	0.07	3.77	0.70	0.05	0.05
X2124	1,2,4-TRICHLOROBENZENE	ug/L	3	2	10	20.0	1	50.0	0.01	2.11	0.04	0.02	0.01
PH1CCO	GAMMA-HCH (HCHACTH OR CYCLOHEXANEH)	ug/L	2	1	10	10.0	1	50.0	0.01	1.25	0.01	0.01	0.01
PH1NDS	ENDOSULFAN SULPHATE	ug/L	3	1	10	10.0	1	50.0	0.02	1.89	0.15	0.15	0.04
PH1NDS	ENDOSULFAN SULPHATE	ug/L	3	1	10	10.0	1	50.0	0.01	1.25	0.01	0.01	0.01
X11CCP	HEXACID-OROCYCLOPENTADIENE	ug/L	3	1	10	10.0	1	50.0	0.06	1.35	0.13	0.13	0.10

TABLE 5-10 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN SECONDARY EFFLUENTS

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
BELT	<u>METALS AND CYANIDE</u>		
	BERYLLIUM, UNFILT. TOTAL		
	<u>BASE NEUTRAL & ACID EXTRACTABLE COMPOUNDS</u>		
P2AMET	AMETRYNE	PMPCRE	P-CRESOL*
P2ATRA	ATRAZINE*		
P4DAZ	DIAZINON*		
P4EPAR	PARATHION ETHYL*		
P4MALA	MALATHION*		
P4MPAR	METHYL PARATHION*		
PM2AMP	2,4-DIMETHYLPHENOL		
PM2ANP	2,4-DINITROPHENOL*		
PM4BPE	4-BROMOPHENYLPHENYLETHER		
PM4CPE	4-CHLOROPHENYLPHENYLETHER		
PMANAA	ALPHA-NAPHTHYLAMINE*		
PMBZE	BIS(2-CHLOROISOPROPYL) ETHER		
PMBNAA	BETA-NAPHTHYLAMINE*		
PMDPE	DIPHENYL ETHER		
PMNND	N-NITROSO-DI-PHENYLAMINE		
PN2CNA	CHLORONAPHTHALENE*		
PNACNE	ACENAPHTHENE*		
PNACNY	ACENAPHTHYLENE		
PNANTH	ANTHRACENE		
PNBAP	BENZOA-PYRENE		
PNB8FA	BENZOBIFLUORANTHENE		
PNB8PH	BIPHENYL		
PNCHRY	CHRYSENE		
PNDABA	DIBENZ(A,H)ANTHRAENE*		
PNFLAN	FLUORANTHENE		
PNFLLO	FLUORENE		
PNGHIP	BENZOGHJPERYLENE*		
PNNP	IDENOL 2,3-CD-PYRENE*		
PODICH	DICHLORAN*		
POTOC	TRI-O-CRESYL PHOSPHATE*		
X3245	2,4,5-TRICHLOROPHENOL*		
	<u>DIOXINS AND FURANS</u>		
P94CDD	TETRACHLORODIBENZODIOXIN*		
P95CDD	PENTACHLORODIBENZODIOXIN*		
P95CDF	PENTACHLORODIBENZOFURAN		
P96CDD	HEXACHLORODIBENZODIOXIN		
P96CDF	HEXACHLORODIBENZOFURAN		
P97CDF	HEPTACHLORODIBENZOFURAN		
	<u>PESTICIDES, HERBICIDES, PCBs</u>		
POPCNB	PCNB	POCAPN	CAPTAN*
P1ALDR	ALDRIN	X1HCBD	HEXACHLOROBTADIENE*
P1BHCD	DELTA-BHC(HEXACHLOROCYCLOHEXANE)		
P1ENDA	ELDRIN ALDEHYDE		
P1HEPE	HEPTACHLOREPOXIDE		
P1OCHL	OXYCHLORDANE		
P1PMIR	MIREX PHOTO		
P1STRO	STROBANE*		
X2HCB	HEXACHLOROBENZENE		
	<u>VOLATILES</u>		
B1OCTE	1-OCTENE	X1ACRO	ACROLEIN
B1VBR	VINYL BROMIDE*	X1ACRY	ACRYLONITRILE*
E1DIEE	DIETHYL ETHER		
L1HEX	HEXANOL		
PM2CEE	2-CHLOROETHYL VINYLETHER*		
X11122	1,1,2,2-TETRACHLOROETHANE*		
X1112T	1,1,2-TRICHLOROETHANE*		
X112CP	1,2-DICHLOROPROPANE		
X113DP	CIS-1,3-DICHLOROPROPENE		
X1BETH	BROMOETHANE*		
X1BROM	BROMOFORM*		
X1CHLE	CHLOROETHANE*		
X1CHLM	CHLOROMETHANE*		
X1CTET	CARBON TETRACHLORIDE		
X1T12D	TRI-1,2-DICHLOROETHYLENE*		
X1TCFM	TRICHLOROFLUOROMETHANE*		
X1VCL	VINYL CHLORIDE*		
X212CB	1,2-DICHLOROBENZENE		
X213CB	1,3-DICHLOROBENZENE		
X214CB	1,4-DICHLOROBENZENE		
X23CTO	3-CHLOROTOLUENE		
X2CPPE	3-CHLOROPROPENE*		

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

TABLE 5-10.b - GLOBAL SUMMARY OF CONTAMINANTS IN SECONDARY EFFLUENTS

CONTAM. INANT	CONTAMINANT NAME	UNITS	QA/QC CODE	# SAMP. DET.	GLOBAL # SAMP. TESTED	GLOBAL % FREQ. DET.	GLOBAL # PLANTS DET.	GLOBAL # PLANTS	GLOBAL % PLANT PREV.	GLOBAL GEO. MEAN	GLOBAL SPREAD FACTOR	GLOBAL MAX. CONC.	GLOBAL MIN. CONC. > DL	DET. LIMIT (DL)
CONVENTIONAL														
DOC	DISSOLVED ORGANIC CARBON	mg/L	0	220	100.0	28	28	28	100.0	8.09	1.60	297.00	2.70	1.00
NH4-N	NITROGEN-TOT. NH4-N/UNT. TOT	mg/L	0	222	100.0	28	28	28	100.0	7.97	2.71	321.00	0.80	0.01
PH	TURBIDITY (CONCN)	mg/L	0	224	100.0	28	28	28	100.0	7.10	1.05	8.18	6.38	1.00
TURB	RESIDUE PARTICULATE	mg/L	0	153	100.0	22	22	22	100.0	7.41	2.90	200.00	0.46	0.25
RSP	CHEMICAL OXYGEN DEMAND	mg/L	0	219	99.6	28	28	28	100.0	10.12	2.00	242.00	3.50	1.00
COD	PHOSPHORUS UNFLT. TOTAL	mg/L	0	211	99.1	28	28	28	100.0	21.22	2.07	110.00	12.00	5.00
PRIT	AMMONIUM UNFLT. PILT. REAC.	mg/L	0	206	97.6	28	28	28	100.0	52.80	1.83	226.00	18.00	0.01
NH4-N	AMMONIUM UNFLT. PILT. REAC.	mg/L	0	223	91.5	28	28	28	100.0	3.90	6.98	27.70	0.05	0.01
NH4-N	NITRATE UNFLT. PILT. REAC.	mg/L	0	194	88.2	27	28	28	96.4	0.22	5.95	3.30	0.03	0.01
NH4-N	NITRATE UNFLT. PILT. REAC.	mg/L	0	186	83.0	27	28	28	96.4	0.23	7.75	28.40	0.25	0.05
RSP/OL	RESIDUE PAR. LOSS ON IGN.	mg/L	0	58	77.5	14	15	15	93.3	7.47	2.71	162.00	5.20	0.00
PH/OL	PHENOLICS (4AAP)	mg/L	0	33	14.7	15	15	15	53.6	0.07	1.86	0.68	0.10	0.10
METALS AND CYANIDE														
SRUT	STRONTIUM UNFLT. TOTAL	ug/L	0	267	100.0	28	28	28	100.0	340.90	2.14	4500.00	80.00	10.00
ZNUT	ZINC UNFLT. TOTAL	ug/L	0	262	98.1	28	28	28	100.0	53.30	2.48	1400.00	10.00	20.00
HGUT	MERCURY UNFLT. TOTAL	ug/L	0	233	94.4	28	28	28	100.0	0.03	2.62	0.36	0.01	0.01
NITUT	NICKEL UNFLT. TOTAL	ug/L	0	196	74.2	27	28	28	96.4	101.70	3.72	5600.00	40.00	20.00
CUUT	COPPER UNFLT. TOTAL	ug/L	0	171	64.0	24	28	28	85.7	22.10	3.60	900.00	10.00	10.00
CRUT	CHROMIUM UNFLT. TOTAL	ug/L	0	47	60.0	21	28	28	75.0	13.00	2.50	190.00	10.00	10.00
CRUT	CHROMIUM UNFLT. TOTAL	ug/L	0	137	51.3	23	28	28	89.3	6.60	1.56	20.00	10.00	10.00
MOUT	MOLYBDENUM UNFLT. TOTAL	ug/L	0	75	28.1	19	28	28	67.9	6.60	1.56	20.00	10.00	10.00
COUT	COBALT UNFLT. TOTAL	ug/L	0	65	24.4	23	28	28	82.1	6.40	1.54	20.00	10.00	10.00
CDUT	CADMIUM UNFLT. TOTAL	ug/L	0	65	26.7	24.3	20	28	71.4	2.10	2.07	14.00	3.00	3.00
PHUT	PHENOL UNFLT. TOTAL	ug/L	0	42	18.9	13	28	28	46.4	1.30	3.68	43.00	6.00	1.00
LEAD	LEAD UNFLT. TOTAL	ug/L	0	222	17.7	9	28	28	60.7	16.50	1.48	91.00	20.00	30.00
AGUT	SILVER UNFLT. TOTAL	ug/L	0	15	3.4	1	28	28	3.6	1.84	1.84	120.00	10.00	10.00
ARSENIC	ARSENIC UNFLT. TOTAL	ug/L	0	267	1.2	1	28	28	3.6	1.70	1.70	16.00	10.00	10.00
SEUT	SELENIUM UNFLT. TOTAL	ug/L	0	252	1.2	1	28	28	3.6	17.10	1.89	80.00	80.00	30.00
BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS														
PMCHRE	M. CHESOL	ug/L	1	7	3.1	4	28	28	14.3	1.70	1.70	30.50	4.30	3.00
PANPHEN	PHENOL	ug/L	2	4	227	1.8	3	28	10.7	1.65	1.59	17.00	7.00	3.00
PANPHEN	NITROBENZENE	ug/L	1	3	228	1.8	4	28	14.3	1.09	1.54	5.70	4.50	2.00
PANBPP	BUTYL BENZYL PHTHALATE	ug/L	1	3	227	1.3	2	28	7.1	1.09	1.59	25.00	3.80	2.00
XPCHCI	PENTACHLOROPHENOL	ug/L	1	3	228	1.3	2	28	7.1	2.71	1.56	58.70	6.80	5.00
PM2ADP	2,4-DICHLOROPHTHALATE	ug/L	1	2	227	0.9	2	28	7.1	2.68	1.50	15.20	5.30	5.00
PM2ADP	2,4-DICHLOROPHTHALATE	ug/L	1	2	227	0.4	1	28	3.6	2.68	1.52	28.80	28.80	5.00
PM2ADT	2,4-DINITROTOLUENE	ug/L	1	228	0.4	1	28	28	3.6	1.61	1.53	25.90	23.90	3.00
PM2ADT	2,6-DINITROTOLUENE	ug/L	1	228	0.4	1	28	28	3.6	2.68	1.52	28.80	28.80	3.00
PM2NP	2-NITROPHENOL	ug/L	1	227	0.4	1	28	28	3.6	2.68	1.52	28.80	28.80	3.00
PM4ADP	2-METHYL 4,6-DINITROPHENOL	ug/L	2	1	227	0.4	1	28	3.6	2.70	1.60	141.70	143.70	5.00
PM4NP	4-NITROPHENOL	ug/L	2	1	227	0.4	1	28	3.6	2.67	1.50	20.80	20.80	5.00
PM4NP	BIS(2-CHLOROETHOXY)METHANE	ug/L	1	228	0.4	1	28	28	3.6	1.08	1.56	32.80	32.80	2.00
PM4NP	BIS(2-CHLOROETHOXY)METHANE	ug/L	2	1	228	0.4	1	28	3.6	1.61	1.54	30.60	30.60	3.00
PM4NP	DIMETHYL PHTHALATE	ug/L	1	228	0.4	1	28	28	3.6	1.07	1.48	2.60	2.60	2.00
PANSP	NITROSO DIETHYLAMINE	ug/L	1	227	0.4	1	28	28	3.6	1.06	1.58	43.60	43.60	2.00
PANSP	NITROSO DIETHYLAMINE	ug/L	2	1	227	0.4	1	28	3.6	1.06	1.58	43.60	43.60	2.00
PMCKRE	O-CHESOL	ug/L	1	228	0.4	1	28	28	3.6	1.08	1.56	29.40	29.40	2.00
PM4AA	BENZAZANTHRACENE	ug/L	1	228	0.4	1	28	28	3.6	1.08	1.56	29.40	29.40	2.00

TABLE 5-10 b - GLOBAL SUMMARY OF CONTAMINANTS IN SECONDARY EFFLUENTS

CONTAMINANT	CONTAMINANT NAME	UNITS	QC CODE	GLOBAL SAMPLS. DET.	GLOBAL SAMPLS. TESTED	% SAMP. TESTED	GLOBAL DET.	# PLANTS DET.	GLOBAL # PLANTS	% PLANT PREV.	GLOBAL G.P.O. MEAN	GLOBAL SPREAD FACTOR	GLOBAL MAX. CONC.	GLOBAL MIN. CONC. > DL	DET. LIMIT (DL)
BASE, NEUTRAL AND ACID EXTRACTABLE COMPOUNDS															
BIODIOL	BENZAKR1310MAN15ENE	u.g/L	1	1	228	0.4	0.4	1	28	3.6	1.07	1.53	14.60	14.60	2.00
	NAPHTH14ENE	u.g/L	1	1	228	0.4	0.4	1	28	3.6	1.06	1.53	17.80	17.80	2.00
	PHEN15ENE	u.g/L	1	1	228	0.4	0.4	1	28	3.6	1.07	1.53	17.80	17.80	2.00
	PHEN16ENE	u.g/L	1	1	228	0.4	0.4	1	28	3.6	1.61	1.54	29.40	29.40	3.00
	2,3,4,5-TETRAHYDROPHEN16ENE	u.g/L	0	1	227	X.30010	0.4	0.4	1	28	3.6	2.66	1.47	5.10	5.10
BIODIOL	2,4,6-TRICHLOROPHEN16ENE	u.g/L	1	1	227	0.4	0.4	1	28	3.6	2.66	1.48	9.20	9.20	5.00
	2,4,6-TRICHLOROPHEN16ENE	u.g/L	1	1	227	0.4	0.4	1	28	3.6	2.66	1.48	9.20	9.20	5.00
BIODIOL AND ACID EXTRACTABLE COMPOUNDS															
BIODIOL	OCTAC10,10-DIBENZOTRIAZINOLIN	u.g/L	1	4	44	9.1	9.1	4	28	14.3	0.31	3.43	11.00	0.10	0.30
	OCTAC10,10-DIBENZOTRIAZINOLIN	u.g/L	1	2	44	4.6	4.6	2	28	7.1	0.38	3.74	4.30	1.40	0.10
	OCTAC10,10-DIBENZOTRIAZINOLIN	u.g/L	1	2	44	4.6	4.6	2	28	7.1	0.28	2.90	0.50	0.50	0.20
	TETRAHYDROOCTAC10,10-DIBENZOTRIAZINOLIN	u.g/L	2	1	44	2.3	2.3	1	28	3.6	0.15	3.90	0.33	0.33	0.10
PESTICIDES, HERBICIDES, INSECTICIDES															
P324D	2,4-DICHLOROPHENOLACETIC ACID	u.g/L	3	177	227	78.0	78.0	28	28	100.0	0.08	4.71	34.00	0.02	0.02
	GAMMA-BHC(HEXACHLOROCYCLOHEXANE)	u.g/L	2	157	227	69.2	69.2	27	28	96.4	0.02	2.48	0.79	0.01	0.01
P324D	1,2,4-TRICHLOROPHENOLACETIC ACID	u.g/L	3	36	227	15.9	15.9	9	28	32.1	0.01	2.44	0.36	0.02	0.01
	1,2,4-TRICHLOROPHENOLACETIC ACID	u.g/L	3	36	227	15.9	15.9	9	28	32.1	0.01	2.44	0.36	0.02	0.01
P324D	1,2,4-TRICHLOROPHENOLACETIC ACID	u.g/L	3	23	227	10.1	10.1	14	28	50.0	0.03	1.83	2.80	0.05	0.05
	1,2,4-TRICHLOROPHENOLACETIC ACID	u.g/L	3	23	227	10.1	10.1	14	28	50.0	0.03	1.83	2.80	0.05	0.05
P324D	BETA-BHC(HEXACHLOROCYCLOHEXANE)	u.g/L	1	9	227	4.0	4.0	6	28	21.4	0.01	1.38	0.07	0.01	0.01
	BETA-BHC(HEXACHLOROCYCLOHEXANE)	u.g/L	2	9	227	4.0	4.0	6	28	21.4	0.01	1.38	0.07	0.01	0.01
P324D	PERMETHYL	u.g/L	1	7	227	3.1	3.1	5	28	17.9	0.01	1.28	0.04	0.01	0.01
	PERMETHYL	u.g/L	1	7	227	3.1	3.1	5	28	17.9	0.01	1.28	0.04	0.01	0.01
P324D	ALPHA-BHC(HEXACHLOROCYCLOHEXANE)	u.g/L	1	5	227	2.2	2.2	4	28	14.3	0.01	1.31	0.06	0.01	0.01
	ALPHA-BHC(HEXACHLOROCYCLOHEXANE)	u.g/L	1	5	227	2.2	2.2	4	28	14.3	0.01	1.31	0.06	0.01	0.01
P324D	GAMMA-BHC(HEXACHLOROCYCLOHEXANE)	u.g/L	3	5	227	2.2	2.2	4	28	14.3	0.01	1.31	0.06	0.01	0.01
	GAMMA-BHC(HEXACHLOROCYCLOHEXANE)	u.g/L	3	5	227	2.2	2.2	4	28	14.3	0.01	1.31	0.06	0.01	0.01
P324D	HEXACHLOROPHENOL	u.g/L	3	3	227	1.3	1.3	2	28	7.1	0.02	1.22	0.05	0.01	0.01
	HEXACHLOROPHENOL	u.g/L	3	3	227	1.3	1.3	2	28	7.1	0.02	1.22	0.05	0.01	0.01
P324D	TOXAPHENE	u.g/L	0	3	227	1.3	1.3	2	28	7.1	0.02	1.22	0.05	0.01	0.01
	TOXAPHENE	u.g/L	0	3	227	1.3	1.3	2	28	7.1	0.02	1.22	0.05	0.01	0.01
P324D	DELDIN	u.g/L	2	2	227	0.9	0.9	1	28	3.6	0.01	1.19	0.04	0.01	0.01
	DELDIN	u.g/L	2	2	227	0.9	0.9	1	28	3.6	0.01	1.19	0.04	0.01	0.01
P324D	ENDOSULFAN II	u.g/L	3	2	227	0.9	0.9	1	28	3.6	0.01	1.16	0.02	0.02	0.01
	ENDOSULFAN II	u.g/L	3	2	227	0.9	0.9	1	28	3.6	0.01	1.16	0.02	0.02	0.01
P324D	PERMETHYL	u.g/L	3	2	227	0.9	0.9	1	28	3.6	0.01	1.16	0.02	0.02	0.01
	PERMETHYL	u.g/L	3	2	227	0.9	0.9	1	28	3.6	0.01	1.16	0.02	0.02	0.01
P324D	ENDOSULFAN I	u.g/L	3	1	227	0.4	0.4	1	28	3.6	0.01	1.12	0.02	0.02	0.01
	ENDOSULFAN I	u.g/L	3	1	227	0.4	0.4	1	28	3.6	0.01	1.12	0.02	0.02	0.01
P324D	ENDURIN	u.g/L	3	1	227	0.4	0.4	1	28	3.6	0.01	1.14	0.03	0.03	0.01
	ENDURIN	u.g/L	3	1	227	0.4	0.4	1	28	3.6	0.01	1.14	0.03	0.03	0.01
P324D	MIREX	u.g/L	3	1	227	0.4	0.4	1	28	3.6	0.01	1.08	0.01	0.01	0.01
	MIREX	u.g/L	3	1	227	0.4	0.4	1	28	3.6	0.01	1.08	0.01	0.01	0.01
P324D	HEXACHLOROCYCLOPENTADIENE	u.g/L	3	1	227	0.4	0.4	1	28	3.6	0.05	1.20	0.26	0.26	0.10
	HEXACHLOROCYCLOPENTADIENE	u.g/L	3	1	227	0.4	0.4	1	28	3.6	0.05	1.20	0.26	0.26	0.10
VOLATILES															
X111C	X11111 TRICHLOROFETHANE	u.g/L	1	18	224	8.0	8.0	8	28	28.6	1.18	1.86	27.00	2.10	2.00
	X11111 TRICHLOROFETHANE	u.g/L	1	18	224	8.0	8.0	8	28	28.6	1.18	1.86	27.00	2.10	2.00
X111C	X11111 TRICHLOROFETHANE	u.g/L	1	18	224	8.0	8.0	8	28	28.6	1.18	1.86	27.00	2.10	2.00
	X11111 TRICHLOROFETHANE	u.g/L	1	18	224	8.0	8.0	8	28	28.6	1.18	1.86	27.00	2.10	2.00
X111C	M- AND P-XYLENES	u.g/L	1	5	224	2.2	2.2	3	28	10.7	1.05	1.38	17.00	2.70	2.00
	M- AND P-XYLENES	u.g/L	1	5	224	2.2	2.2	3	28	10.7	1.05	1.38	17.00	2.70	2.00
X111C	O-XYLENE	u.g/L	1	4	224	1.8	1.8	3	28	10.7	1.03	1.30	16.00	2.60	2.00
	O-XYLENE	u.g/L	1	4	224	1.8	1.8	3	28	10.7	1.03	1.30	16.00	2.60	2.00
X111C	1,2-DICHLOROFETHANE	u.g/L	1	4	224	1.8	1.8	3	28	10.7	1.03	1.30	16.00	2.60	2.00
	1,2-DICHLOROFETHANE	u.g/L	1	4	224	1.8	1.8	3	28	10.7	1.03	1.30	16.00	2.60	2.00
X111C	CHLOROBROMOMETHANE	u.g/L	1	4	224	1.8	1.8	3	28	10.7	1.02	1.13	12.00	2.30	2.00
	CHLOROBROMOMETHANE	u.g/L	1	4	224	1.8	1.8	3	28	10.7	1.02	1.13	12.00	2.30	2.00

TABLE 5-10 b - GLOBAL SUMMARY OF CONTAMINANTS IN SECONDARY EFFLUENTS

CONTAMINANT	CONTAMINANT NAME	UNITS	Q&QC CODE	GLOBAL # SAMP. DET.	GLOBAL # SAMP. TESTED	GLOBAL % FREQ. DET.	GLOBAL # PLANTS DET.	GLOBAL # PLANTS	GLOBAL % PLANT PREV.	GLOBAL GEO. MEAN	GLOBAL SPREAD FACTOR	GLOBAL MAX. CONC.	GLOBAL MIN. CONC. > DL	DET. LIMIT (DL)
DEBCL	BROMODICHLOROBENZENE	ug/L	1	3	224	1.3	2	28	7.1	4.93	1.21	10.00	10.00	10.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	3	224	1.3	2	28	7.1	1.02	1.21	12.00	2.70	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	3	224	1.3	3	28	10.7	1.01	1.10	2.30	2.20	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	2	224	0.9	2	28	7.1	1.01	1.11	4.10	2.20	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	1	224	0.5	1	28	3.6	1.51	1.16	13.00	13.00	3.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	1	224	0.5	1	28	3.6	1.00	1.06	2.40	2.40	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	1	224	0.5	1	28	3.6	1.50	1.05	3.00	3.00	3.00
DEBCL	BROMODICHLOROBENZENE	ug/L	2	1	224	0.5	1	28	3.6	1.07	1.12	5.00	5.00	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	1	224	0.5	1	28	3.6	1.01	1.12	2.20	2.20	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	1	224	0.5	1	28	3.6	1.00	1.05	2.10	2.10	2.00

VOLATILES

DEBCL	BROMODICHLOROBENZENE	ug/L	1	3	224	1.3	2	28	7.1	4.93	1.21	10.00	10.00	10.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	3	224	1.3	2	28	7.1	1.02	1.21	12.00	2.70	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	3	224	1.3	3	28	10.7	1.01	1.10	2.30	2.20	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	2	224	0.9	2	28	7.1	1.01	1.11	4.10	2.20	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	1	224	0.5	1	28	3.6	1.51	1.16	13.00	13.00	3.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	1	224	0.5	1	28	3.6	1.00	1.06	2.40	2.40	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	1	224	0.5	1	28	3.6	1.50	1.05	3.00	3.00	3.00
DEBCL	BROMODICHLOROBENZENE	ug/L	2	1	224	0.5	1	28	3.6	1.07	1.12	5.00	5.00	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	1	224	0.5	1	28	3.6	1.01	1.12	2.20	2.20	2.00
DEBCL	BROMODICHLOROBENZENE	ug/L	1	1	224	0.5	1	28	3.6	1.00	1.05	2.10	2.10	2.00

5.3.7 Summary of Contaminants in Tertiary Effluents

Only the Guelph WPCP of all the study plants provided tertiary treatment. The 128 contaminants that were not detected in the Guelph tertiary effluent including 12 that were not confirmed, are presented in Table 5-11(a). Also indicated are the 34 contaminants that were not detected in any sample type at any WPCP.

Thirty-one organic contaminants, 12 metals and cyanide were detected in at least one sample. Table 5-11(b) presents the listing of contaminants found in the Guelph tertiary effluent. Metals were the most frequently detected contaminant, with 6 metals (Al, Cr, Cu, Hg, Sr and Zn) detected in more than 75 percent of samples. Only 2 pesticide and herbicide compounds (2,4-Dichlorophenoxyacetic acid and gamma-BHC) were present in more than 50 percent of the samples. The most frequently detected volatile organic compound was detected in only 50 percent of the samples. The most frequently detected base neutral and acid extractable compound was detected in only 30 percent of samples, and there were no dioxin/furan compounds detected in the Guelph tertiary effluent.

5.3.8 Summary of Contaminants in Raw Sludges

Table 5-12(a) presents a list of the 85 contaminants that were not detected in any of the raw sludge from any WPCP, including 6 compounds that were not confirmed. Also indicated are the 34 compounds that were not detected in any sample type at any WPCP.

Table 5-12(b) presents the summary of detected contaminants for all raw sludges. A total of 59 organic compounds, 15 metals and cyanide were detected in any sample. The most prevalent organic compounds were the pesticides and herbicides, with 11 compounds detected in at least 40 percent of the plants. Only 2 of the base neutral and acid extractable compounds, 4 of the volatile compounds and 1 dioxin compound were detected at more than 20 percent of the plants. All of the metals except beryllium were detected at greater than 64 percent of the plants. Cyanide was detected at 10 percent of the plants.

The most frequently detected contaminants were metals, with 12 metals detected in at least 80 percent of the samples, including 5 (Al, Hg, Sr, Zn, Cu) that were detected in all of the samples. Only 1 of the base neutral and acid extractable compounds and 1 volatile compound were detected in more than 30 percent of the samples, and only one dioxin (Octachlorodibenzodioxin) was detected in greater than 12

TABLE 5-11 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN TERTIARY EFFLUENTS

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
BEUT CCNFUR COUT MOUT	<p><u>METALS AND CYANIDE</u></p> <p>BERYLLIUM,UNFLT.TOTAL CYANIDE-FREE,UNFLT.REAC. COBALT,UNFLT.TOTAL MOLYBDENUM,UNFLT.TOTAL</p> <p><u>BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS</u></p>		
P2AMET P2ATRA P4DAZ P4EPAR P4MALA P4MPAR PM24DP PM24DT PM24MP PM42NP PM42DT PM42NP PM46DP PM48PE PM4CPE PM4NP PMANAA PMB2EM PMB2E PMB2NE PMBBP PMBNAA PMDMP PMNTTB PMNSD PMNPN PMOCRE PMPCMC PMPHEN PN2CNA PNACNE PNACNY PNANTH PNBAA PNBAP PNBBFA PNBPH PNBKF PNCHRY PNDAHA PNFLAN PNFLLO PNGHIP PNINP PNNAPH PNPHEN PNPTR PODICH POTOC X300IO X3245 X3246 X3PCPH	<p>AMETRYNE ATRAZINE* DIAZINON* PARATHION ETHYL* MALATHION* METHYLPARATHION* 2,4-DICHLOROPHENOL 2,4-DINITROTOLUENE 2,4-DIMETHYLPHENOL 2,4-DINITROPHENOL* 2,6-DINITROTOLUENE 2-NITROPHENOL 2-METHYL 4,6-DINITROPHENOL 4-BROMOPHENYLPHENYLETHYR 4-CHLOROPHENYLPHENYLETHYR 4-NITROPHENOL ALPHA-NAPHTHYLAMINE* BIS(2-CHLORETHOXY)METHANE BIS(2-CHLOROISOPROPYL)ETHER BIS(2-CHLOROMETHYL)ETHER BUTYL BENZYL PHTHALATE BETA-NAPHTHYLAMINE* DIMETHYL PHTHALATE NITROBENZENE N-NITROSO-DI-PHENYLAMINE N-NITROSO-DI-N-PROPYLAMINE O-CRESOL P-CHLORO-M-CRESOL PHENOL CHLORONAPHTHALENE* ACENAPHTHENE* ACENAPHTHYLENE ANTHRACENE BENZOA)ANTHRACENE BENZOA)PYRENE BENZOB)FLUORANTHENE BIPHENYL BENZOK)FLUORANTHENE CHRYSENE DIBENZOA,H)ANTHRACENE* FLUORANTHENE FLUORENE BENZOG,H,I)PERYLENE* IDENO(1,2,3-CD)PYRENE* NAPHTHALENE PHENANTHRENE PYRENE DICHLORAN* TRI-O-CRESYL PHOSPHATE* 2-CHLOROPHENOL 2,4,5-TRICHLOROPHENOL* 2,4,6-TRICHLOROPHENOL PENTACHLOROPHENOL</p> <p><u>DIOXINS AND FURANS</u></p> <p>TETRACHLORODIBENZODIOXIN* TETRACHLORODIBENZOFURAN* PENTACHLORODIBENZODIOXIN PENTACHLORODIBENZOFURAN HEXACHLORODIBENZODIOXIN HEXACHLORODIBENZOFURAN* HEPTACHLORODIBENZOFURAN OCTACHLORODIBENZOFURAN</p> <p><u>PESTICIDES, HERBICIDES, PCBs</u></p> <p>PCNB ALPHA-BHC(HEXACHLORCYCLOHEXANE) GAMMA-BHC(HEXACHLORCYCLOHEXANE) METHOXYCHLOR MIREX</p>	PMPCRE	P-CRESOL*
P94CDD P94CDF P95CDD P95CDF P96CDD P96CDF P97CDF P98CDF			
P0PCNB P1BICA P1BHCG P1BMDT P1MIRX			
		POCAPN PIEND2 PIENDR PIENDS PIHEPT	CAPTAN* ENDOSULFAN II ENDRIN ENDOSULFAN SULPHATE HEPTACHLOR

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

TABLE 5-11 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN TERTIARY EFFLUENTS

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
P10CHL P1PMIR P1STRO P1TOX X24CB	PESTICIDES, HERBICIDES, PCBS OXYCHLORDANE MIREX PHOTO STROBANE* TOXAPH HEXACHLOROBENZENE	P32AD X1HCB D X1HCPC X2134	2,4-DICHLOROPHENOXACETIC ACID HEXACHLOROBUTADIENE* HEXACHLOROCCYCLOPENTADIENE 1,2,4-TRICHLOROBENZENE
B10CTE B1VBR B2BDCL B2EBNZ B2MPXY B20XYL B2STYR E1DIE L1HEX PMDCEE X1111T X11122 X1112T X111CE X112CE X112CP X113DP X113DR X1ACTO X1BDCM X1BETH X1BROM X1CDBM X1CDCE X1CHLE X1CHLM X1CHLO X1CTET X1DCFM X1DQLE X1T12D X1TCFM X1TETR X1TRIC X1VCL X212CB X213CB X214CB X23CTO X2CBEN X2CPPE	VOLATILES 1-OCTENE VINYL BROMIDE* BROMODICHLOROBENZENE ETHYLBENZENE M-, AND P-XYLENES O-XYLENE STYRENE DIETHYL ETHER HEXANOL 2-CHLOROETHYL VINYLETHER* 1,1,1-TRICHLOROETHANE 1,1,2,2-TETRACHLOROETHANE* 1,1,2-TRICHLOROETHANE* 1,1-DICHLOROETHANE 1,2-DICHLOROETHANE 1,2-DICHLOROPROPANE CIS-1,3-DICHLOROPROPENE TRANS-1,3-DICHLOROPROPENE ALPHA-CHLOROTOLUENE BROMODICHLOROMETHANE BROMOETHANE* BROMOFORM* CHLORODIBROMOMETHANE CIS-1,2-DICHLOROETHYLENE CHLOROETHANE* CHLOROMETHANE* CHLOROFORM CARBON TETRACHLORIDE DICHLORODIFLUOROMETHANE 1,1-DICHLOROETHENE TRI-1,2-DICHLOROETHYLENE* TRICHLORODIFLUOROMETHANE* TETRACHLOROETHYLENE TRICHLOROETHYLENE VINYL CHLORIDE* 1,2-DICHLOROBENZENE 1,3-DICHLOROBENZENE 1,4-DICHLOROBENZENE 3-CHLOROTOLUENE CHLOROBENZENE 3-CHLOROPROPENE*	X1ACRO X1ACRY	ACROLEIN ACRYLONITRILE*

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

TABLE 5-11b - GLOBAL SUMMARY OF CONTAMINANTS IN TERTIARY EFFLUENTS

CONTAM- INANT	CONTAMINANT NAME	UNITS	QA/QC CODE	GLOBAL # SAMP. TESTED	GLOBAL % PLANT DET.	GLOBAL # PLANTS DET.	GLOBAL % PLANT PREV.	GLOBAL GEO. MEAN	GLOBAL SPREAD FACTOR	GLOBAL MAX. CONC.	GLOBAL MIN. CONC.	GLOBAL MAX. CONC. > DL	DET. LIMIT (DL)
CONVENTIONAL													
BOD5	BOD 5 DAY TOTAL DEMAND	mg/L	0	9	100.0	1	100.0	24.64	1.48	56.40	16.00		0.00
COD	CHEMICAL OXYGEN DEMAND	mg/L	0	9	100.0	1	100.0	99.25	1.80	282.00	46.00		0.00
DOC	DISSOLVED ORGANIC CARBON	mg/L	0	9	100.0	1	100.0	8.74	1.19	10.90	7.00		0.00
NNHTR	AMMONIUM TOTAL FILT REAC.	mg/L	0	9	100.0	1	100.0	18.24	1.08	20.80	16.60		0.00
NNHTR	NITROGEN TOTAL FILT REAC.	mg/L	0	9	100.0	1	100.0	23.22	1.17	34.00	19.80		0.00
PH	(LOG10)(CONC)	mg/L	0	9	100.0	1	100.0	7.10	1.04	7.49	6.82		0.00
TP	PHOSPHATE TOTAL	mg/L	0	10	100.0	1	100.0	1.56	1.92	5.93	0.78		0.50
RSPT	RESIDUE PARTICULATE	mg/L	0	8	100.0	1	100.0	31.11	1.74	11.80	17.10		0.00
TURB	TURBIDITY	mg/L	0	5	88.9	1	100.0	10.61	1.95	16.00	6.00		0.00
NNO2R	NITRITE FILT REACT.	mg/L	0	8	100.0	1	100.0	0.05	3.45	0.15	0.03		0.00
NNO3R	NITRATE TOTAL FILT REAC.	mg/L	0	5	55.6	1	100.0	0.14	4.23	0.50	0.40		0.00
METALS AND CYANIDE													
ALUT	ALUMINUM UNFLT TOTAL	ug/L	0	11	100.0	1	100.0	1252.00	1.88	2990.00	560.00		500.00
CRUT	CHROMIUM UNFLT TOTAL	ug/L	0	11	100.0	1	100.0	68.80	1.84	270.00	30.00		500.00
CUUT	COPPER UNFLT TOTAL	ug/L	0	2	100.0	1	100.0	54.80	3.03	120.00	25.00		500.00
HGUT	MERCURY UNFLT TOTAL	ug/L	0	9	100.0	1	100.0	0.17	1.73	0.39	0.07		0.01
SRUT	STRONTIUM UNFLT TOTAL	ug/L	0	11	100.0	1	100.0	1170.70	1.11	1400.00	990.00		500.00
ZNUT	ZINC UNFLT TOTAL	ug/L	0	11	100.0	1	100.0	959.60	1.30	1600.00	650.00		500.00
CNUTR	CYANIDE UNFLT REAC.	ug/L	0	7	9	100.0	1	100.0	14.60	4.19	70.00	7.00	3.00
CDUT	CADMIUM UNFLT REAC.	ug/L	0	11	100.0	1	100.0	6.30	1.74	20.00	11.00		500.00
COUT	COBALT UNFLT TOTAL	ug/L	0	3	100.0	1	100.0	15.00	1.00	100.00	10.00		100.00
PRUT	LEAD UNFLT TOTAL	ug/L	0	2	11	100.0	1	100.0	55.90	1.50	130.00	110.00	500.00
MOUT	SILVER UNFLT TOTAL	ug/L	0	1	9.1	1	100.0	6.60	2.02	10.00	10.00		50.00
MOUT	MOLYBDENUM UNFLT TOTAL	ug/L	0	1	9.1	1	100.0	11.30	1.52	40.00	40.00		250.00
NIUT	NICKEL UNFLT TOTAL	ug/L	0	1	9.1	1	100.0	25.80	1.12	36.00	36.00		3000.00
BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS													
PM2AMP	2,4-DIMETHYLPHENOL	ug/L	2	3	30.0	1	100.0	3.40	1.57	7.50	5.60		500.00
PM2PHN	PHENOL	ug/L	2	3	30.0	1	100.0	1.99	1.59	4.40	3.40		500.00
PM2ADP	2,4-DICHLOROPHENOL	ug/L	1	1	100.0	1	100.0	2.87	1.53	9.90	9.90		500.00
PM2DOP	2,4-DICHLOROPHENOL	ug/L	1	1	100.0	1	100.0	1.70	1.49	5.30	5.30		500.00
PM2NDP	2,6-DINITROPHENOL	ug/L	1	1	100.0	1	100.0	1.77	1.68	7.70	7.70		500.00
PM2DOP	2-NITROPHENOL	ug/L	1	1	100.0	1	100.0	3.18	2.34	27.00	27.00		500.00
PMR2EAM	BIS(2-CHLOROETHOXY)METHANE	ug/L	1	1	100.0	1	100.0	1.31	2.34	14.80	14.80		200.00
PMR2EAM	BIS(2-CHLOROETHOXY)METHANE	ug/L	2	1	100.0	1	100.0	1.86	1.97	12.70	12.70		500.00
PMR2EAM	BIS(2-CHLOROETHOXY)METHANE	ug/L	1	1	100.0	1	100.0	1.13	1.46	3.30	3.30		200.00
PMR2EAM	BIS(2-CHLOROETHOXY)METHANE	ug/L	1	1	100.0	1	100.0	1.34	2.23	19.20	19.20		200.00
PMR2EAM	BIS(2-CHLOROETHOXY)METHANE	ug/L	1	1	100.0	1	100.0	1.29	2.23	12.60	12.60		200.00
PMR2EAM	BIS(2-CHLOROETHOXY)METHANE	ug/L	1	1	100.0	1	100.0	1.85	1.95	14.60	14.60		200.00
PMR2EAM	BIS(2-CHLOROETHOXY)METHANE	ug/L	1	1	100.0	1	100.0	1.85	1.95	12.00	12.00		200.00
PMR2EAM	BIS(2-CHLOROETHOXY)METHANE	ug/L	1	1	100.0	1	100.0	3.10	1.97	21.50	21.50		500.00

TABLE 5-8-b - GLOBAL SUMMARY OF CONTAMINANTS IN TERTIARY EFFLUENTS

CONTAM- INANT	CONTAMINANT NAME	UNITS	QC CODE	GLOBAL # SPTS. DET.	GLOBAL # SPTS. TESTED	GLOBAL % DET.	GLOBAL # SPTS. DET.	GLOBAL PLANTS	GLOBAL % PREV.	GLOBAL MEAN	GLOBAL SPREAD FACTOR	GLOBAL CONC.	GLOBAL CONC. >DL	DST. LIMIT (DL)
PESTICIDES, HERBICIDES, F'BS														
P324D	2,4-DICHLOROPHENOXYACETIC ACID	ug/L	3	9	10	90.0	1	1	100.0	0.12	2.56	0.67	0.05	0.40
P14HCO	GAMMA-BHC (HEXACHLOROCYCLOHEXANE)	ug/L	2	8	10	80.0	1	1	100.0	0.04	3.37	0.75	0.02	0.20
P14HCB	BETA-BHC (HEXACHLOROCYCLOHEXANE)	ug/L	1	3	10	30.0	1	1	100.0	0.02	3.03	0.17	0.05	0.20
P14MDT	METHOXYCHLOR	ug/L	1	2	10	20.0	1	1	100.0	0.07	1.90	0.26	0.20	1.00
P314V	SILVEX	ug/L	3	2	10	20.0	1	1	100.0	0.06	1.34	0.10	0.10	1.00
P14HCO	2,4-DICHLOROPHENOXYACETIC ACID	ug/L	3	1	10	10.0	1	1	100.0	0.01	1.66	0.05	0.05	0.20
P14HCB	GAMMA-BHC (HEXACHLOROCYCLOHEXANE)	ug/L	2	1	10	10.0	1	1	100.0	0.02	1.55	0.06	0.06	0.20
P14MDT	METHOXYCHLOR	ug/L	1	1	10	10.0	1	1	100.0	0.01	1.66	0.05	0.05	0.20
P314V	SILVEX	ug/L	3	1	10	10.0	1	1	100.0	0.01	1.66	0.05	0.05	0.20
VOLATILES														
X12TR	TRICHLOROETHYLENE	ug/L	1	5	10	50.0	1	1	100.0	3.50	4.30	50.00	5.80	40.00
X1111	1,1,1-TRICHLOROETHANE	ug/L	1	4	10	40.0	1	1	100.0	2.30	2.61	25.00	2.60	40.00
X1001	1,1-DICHLOROETHYLENE	ug/L	1	2	10	20.0	1	1	100.0	1.56	2.61	14.00	6.10	40.00
X1001	1,1-DICHLOROETHYLENE	ug/L	1	2	10	20.0	1	1	100.0	1.56	2.61	14.00	6.10	40.00
X111E	DETHYLENE	ug/L	1	1	10	10.0	1	1	100.0	1.28	2.19	12.00	12.00	40.00
X111E	1,1-DICHLOROETHYLENE	ug/L	1	1	10	10.0	1	1	100.0	1.23	1.91	7.80	7.80	40.00
X112E	1,2-DICHLOROETHANE	ug/L	1	1	10	10.0	1	1	100.0	1.10	1.37	2.70	2.70	40.00
X100E	CIS-1,2-DICHLOROETHYLENE	ug/L	1	1	10	10.0	1	1	100.0	1.17	1.66	5.00	5.00	40.00
X111E	1,1-DICHLOROETHYLENE	ug/L	1	1	10	10.0	1	1	100.0	1.26	2.05	9.70	9.70	40.00

TABLE 5-12 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN RAW SLUDGES

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
	<u>BASE NEUTRAL AND ACID EXTRACTABLE COMPOUNDS</u>		
P2AMET	AMETRYNE	PMPCRE	P-CRESOL*
P2ATRA	ATRAZINE*		
P4DIAZ	DIAZINON*		
P4EPAR	PARATHION ETHYL*		
P4MALA	MALATHION*		
P4MPAR	METHYL PARATHION*		
P424DP	2,4-DICHLOROPHENOL		
P424DT	2,4-DINITROTOLUENE		
P424MP	2,4-DIMETHYLPHENOL		
P424NP	2,4-DINITROPHENOL*		
P424DT	2,6-DINITROTOLUENE		
P424NP	2-NITROPHENOL		
P446DP	2-METHYL 6-DINITROPHENOL		
P448PE	4-BROMOPHENYLPHENYLETHER		
P44CPE	4-CHLOROPHENYLPHENYLETHER		
P44NP	4-NITROPHENOL		
P4ANAA	ALPHA-NAPHTHYLAMINE*		
P4B2EM	BIS(2-CHLOROETHOXY)METHANE		
P4B2NE	BIS(2-CHLOROMETHYL)ETHER		
P4BNAA	BETA-NAPHTHYLAMINE*		
P4DMP	DIMETHYL PHTHALATE		
P4N2NP	N-NITROSO-DI-N-PROPYLAMINE		
PMOCRE	O-CRESOL		
PMPCMC	P-CHLORO-M-CRESOL		
PM2CNA	CHLORONAPHTHALENE*		
PM4ACNE	ACENAPHTHENE*		
P4BAA	BENZOAANTHRACENE		
P4BAP	BENZOA)PYRENE		
P4BIPH	BIPHENYL		
P4BKF	BENZOKI)FLUORANTHENE		
P4DAHA	DIBENZOA)ANTHRACENE*		
P4FLUO	FLUORENE		
P4GHIP	BENZOGH)J)PERYLENE*		
P4N2P	IDENOX(1,2,3-CD)PYRENE*		
P4DICH	DICHLORAN*		
P4OTOC	TRIO-CRESYL PHOSPHATE*		
X3001O	2-CHLOROPHENOL		
X3245	2,4,5-TRICHLOROPHENOL*		
X3246	2,4,6-TRICHLOROPHENOL		
	PENTACHLOROPHENOL		
	<u>DIOXINS AND FURAN</u>		
X3PCPH	TETRACHLORODIBENZODIOXIN*		
P4CDD	PENTACHLORODIBENZODIOXIN		
P45CDD	HEXACHLORODIBENZODIOXIN		
P46CDD			
	<u>PESTICIDES, HERBICIDES, PCBs</u>		
	<u>PCNB</u>		
P4PCNB	ELDRIN ALDEHYDE	P4CAPN X1HCBD X2HCE	CAPTAN* HEXACHLOROBUTADIENE* HEXACHLOROETHANE
P4IENDA	MIREX PHOTO		
P4PMIR	STROBANE*		
P4ISTRO	TOXAPHENE		
P4ITOX			
	<u>VOLATILES</u>		
	<u>1-OCTENE</u>		
B1OCTE	VINYL BROMIDE*	X1ACRO X1ACRY	ACROLEIN ACRYLONITRILE*
B1VBR	BROMODICHLOROBENZENE		
B2BDCL	DIETHYL ETHER		
E1DIEE	HEXANOL		
L1HEX	2-CHLOROETHYL VINYLETHER*		
P4OC2E	1,1,1-TRICHLOROETHANE		
X1111T	1,1,2,2-TETRACHLOROETHANE*		
X11112	1,1,2-TRICHLOROETHANE*		
X11112T	1,1-DICHLOROETHANE		
X111CE	1,2-DICHLOROETHANE		
X112CE	1,2-DICHLOROPROPANE		
X112CP	CIS-1,3-DICHLOROPROPENE		
X113DP	TRANS-1,3-DICHLOROPROPENE		
X113DR	ALPHA-CHLOROTOLUENE		
X1ACTO	BROMOETHANE*		
X1BETH	BROMOFORM*		
X1BROM	CIS-1,2-DICHLOROETHYLENE		
X1CDCE	CHLOROETHANE*		
X1CHLE	CHLOROMETHANE*		
X1CHLM	CARBON TETRACHLORIDE		
X1CTET			

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

TABLE 5-12a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN RAW SLUDGES

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
X1DCFM X1DCLE X1TI2D X1TCFM X1TRIC X1VCL X2I2CB X23CTO X2CBEN X2CPPE	<u>VOLATILES</u>		
	DICHLORODIFLUOROMETHANE		
	1,1-DICHLOROETHENE		
	TRI-1,2-DICHLOROETHYLENE*		
	TRICHLOROFLUOROMETHANE*		
	TRICHLOROETHYLENE		
	VINYL CHLORIDE*		
	1,2-DICHLOROBENZENE		
	3-CHLOROTOLUENE		
	CHLOROBENZENE		
	3-CHLOROPROPENE*		

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

TABLE 5-12 b - GLOBAL SUMMARY OF CONTAMINANTS IN RAW SLUDGES

CONTAM. INANT	CONTAMINANT NAME	UNITS (Q/C DRY CODE & SAMP. WEIGHT)	GLOBAL # SAMP. TESTED	GLOBAL % FREQ. DET.	GLOBAL # PLANTS DET.	GLOBAL # PLANTS	GLOBAL % PLANT PREV.	GLOBAL GEO. MEAN	GLOBAL SPREAD FACTOR	GLOBAL MAX. CONC.	GLOBAL MIN. CONC. > DL
DIOXINS AND FURANS											
P97CDD	OCTACHLORODIBENZODIOXIN	ug/g	25	50	20	34	58.8	8.50	3.60	309.10	1.30
P97CDD	HEPTACHLORODIBENZODIOXIN	ug/g	1	12.0	6	34	11.8	5.40	3.42	31.20	1.80
P97CDD	OCTACHLORODIBENZOFURAN	ug/g	1	6	3	34	8.8	2.40	2.94	25.40	3.20
P97CDD	HEPTACHLORODIBENZOFURAN	ug/g	2	6.0	3	34	8.8	3.90	3.57	2.50	2.50
P97CDD	PENTACHLORODIBENZOFURAN	ug/g	1	30	1	34	2.9	5.50	4.45	3.80	3.80
P97CDD	HEXACHLORODIBENZOFURAN	ug/g	1	2.0	1	34	2.9	7.20	4.98	25.30	25.30
P97CDD	HEPTACHLORODIBENZOFURAN	ug/g	1	2.0	1	34	2.9	7.20	4.98	25.30	25.30
PESTICIDES, HERBICIDES, PCB'S											
P97CET	PCB, TOTAL	ug/g	2	78.4	27	34	79.4	88.70	3.18	2895.60	19.50
P97CDD	2,4-DICHLOROPHTHOXYACETIC ACID	ug/g	3	74.5	28	34	82.3	93.20	6.30	3742.70	15.30
P97CDD	GAMMA-BHC(HEXACHLOROCYCLOHEXANE)	ug/g	2	54.9	23	34	67.7	8.90	2.69	101.00	4.70
P97CDD	SIN-VEK	ug/g	3	45.1	19	34	55.9	47.30	3.40	169.60	37.00
P97CDD	PP-DDD	ug/g	1	43.1	18	34	42.9	27.70	2.31	177.00	17.00
P97CDD	2,4,5-TRICHLOROPHTHOXYACETIC ACID	ug/g	2	31.1	16	34	34.0	48.90	3.10	744.70	31.00
P97CDD	PP-DDD	ug/g	2	31.1	16	34	34.0	48.90	3.10	744.70	31.00
P97CDD	GAMMA-CHLORDANE	ug/g	2	31.1	16	34	34.0	48.90	3.10	744.70	31.00
P97CDD	METHOXYCHLOR	ug/g	1	37.3	17	34	50.0	7.20	2.44	36.10	2.90
P97CDD	BETA-BHC(HEXACHLOROCYCLOHEXANE)	ug/g	1	37.3	15	34	44.1	45.80	5.15	3179.70	17.80
P97CDD	ALPHA-CHLORDANE	ug/g	1	35.3	13	34	38.2	7.40	3.29	111.30	2.30
P97CDD	1,2,4-TRICHLOROBENZENE	ug/g	3	35.3	16	34	47.1	5.90	2.38	34.00	2.30
P97CDD	HEXACHLOROBENZENE	ug/g	2	35.3	13	34	41.2	9.30	4.06	322.60	10.60
P97CDD	4,4'-DICHLORODIBENZYL	ug/g	1	35.3	13	34	38.2	5.60	2.65	145.40	1.00
P97CDD	HEXACHLOROCYCLOHEXANE	ug/g	1	23.5	12	34	33.3	5.30	2.74	155.60	2.30
P97CDD	HEXACHLOROCYCLOHEXANE	ug/g	2	23.5	11	34	32.4	5.00	2.62	185.50	3.50
P97CDD	ENDOSULFAN I	ug/g	3	17.7	7	34	20.6	4.60	2.56	129.90	4.80
P97CDD	ENDOSULFAN II	ug/g	3	15.7	8	34	21.5	4.20	2.21	37.40	3.20
P97CDD	OXETHYLCHLORANE	ug/g	2	15.7	7	34	20.6	4.50	2.27	21.70	4.60
P97CDD	HEPTACHLOR	ug/g	3	13.7	7	34	20.6	4.50	2.83	101.00	1.00
P97CDD	ALDRIN	ug/g	1	11.8	6	34	17.7	4.60	2.50	49.80	4.30
P97CDD	ENDOSULFAN I	ug/g	3	11.8	6	34	17.7	4.60	2.50	49.80	4.30
P97CDD	ENDOSULFAN II	ug/g	3	11.8	6	34	17.7	4.60	2.50	49.80	4.30
P97CDD	PP-DDD	ug/g	3	11.8	6	34	17.7	4.60	2.50	49.80	4.30
P97CDD	DELTA-BHC(HEXACHLOROCYCLOHEXANE)	ug/g	3	11.8	6	34	17.7	4.60	2.50	49.80	4.30
P97CDD	MIREX	ug/g	3	5.9	3	34	8.8	4.00	2.22	103.30	15.90
P97CDD	HEXACHLOROCYCLOPENTADIENE	ug/g	1	3.9	2	34	5.9	4.00	2.37	101.00	23.90
P97CDD	HEXACHLOROCYCLOPENTADIENE	ug/g	3	3.9	2	34	5.9	4.00	2.37	101.00	23.90
P97CDD	HEXACHLOROCYCLOPENTADIENE	ug/g	3	3.9	2	34	5.9	4.00	2.37	101.00	23.90
VOLATILES											
B97CDD	M- AND P-XYLENES	ug/g	1	31.4	14	34	41.2	1345.90	3.86	47146.40	1776.00
B97CDD	CHLOROFORM	ug/g	1	23.5	12	34	35.3	1225.10	4.24	95218.10	1688.30
B97CDD	ETHYL BENZENE	ug/g	1	19.6	9	34	26.3	890.70	2.60	24413.90	738.40
B97CDD	ETHYL CHLORIDE	ug/g	1	15.7	8	34	23.5	841.60	2.31	14172.00	1066.90
B97CDD	STYRENE	ug/g	1	2.0	1	34	2.9	659.40	1.96	6010.90	6010.90
B97CDD	BROMODICHLOROMETHANE	ug/g	1	2.0	1	34	2.9	659.40	1.96	6010.90	6010.90
B97CDD	CHLORODIBROMOMETHANE	ug/g	1	2.0	1	34	2.9	659.40	1.96	6010.90	6010.90
B97CDD	1,3-DICHLOROBENZENE	ug/g	1	2.0	1	34	2.9	659.40	1.96	6010.90	6010.90
B97CDD	1,4-DICHLOROBENZENE	ug/g	1	2.0	1	34	2.9	659.40	1.96	6010.90	6010.90

percent of the samples. The most frequently detected pesticide and herbicide compound was detected in 78 percent of the samples and 16 of the pesticide/herbicide compounds were detected in more than 20 percent of samples.

5.3.9 Summary of Contaminants in Treated Sludges

Table 5-13(a) presents a list of the 81 contaminants that were not detected in any treated sludge samples from any WPCP, including 5 contaminants that were not confirmed. Also indicated are the 34 contaminants not detected in any sample type at any WPCP.

Fifteen metals and 64 organic compounds were detected in at least one treated sludge sample. Table 5-13(b) summarizes the contaminants detected in treated sludges. The most prevalent organic compounds were the pesticides and herbicides, with 10 compounds detected at more than 40 percent of the plants. The most prevalent base neutral and acid extractable and volatile compounds were detected at fewer than 35 percent of plants. One dioxin compound (Octachlorodibenzodioxin) was detected at 65 percent of the plants. There were 13 metals detected at more than 86 percent of the plants and 6 (Ag, Al, Cr, Cu, Sr and Zn) detected at all of the plants.

The most frequently detected contaminants were metals, with 13 metals detected in at least 82 percent of samples, and 6 detected in all the samples. None of the base neutral and acid extractable, or volatile compounds were detected in more than 30 percent of samples. One dioxin compound was detected in 53 percent of samples, and the remaining dioxins were detected in less than 20 percent. The most frequently detected pesticide and herbicide compound was detected in 68 percent of samples, and 16 of the pesticide/herbicide compounds were detected in more than 20 percent of the samples.

5.3.10 Summary of Contaminants Detected in Any Sample Type

Table 5-14 presents a summary of contaminants detected in any sample type. The Table provides for each of the five contaminant groups, the number of compounds detected, the maximum percentage prevalence (ie. the maximum percentage of all WPCPs in which the contaminant was identified) for any contaminant in the group and the maximum percentage frequency (ie. the maximum percentage of all samples of a given type in which the contaminant was identified) for any contaminant in the group.

As noted throughout Section 5.3 metals were the most prevalently (most WPCPs) and most frequently detected contaminants in all sample types.

TABLE 5-13 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN TREATED SLUDGES

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
CCNFUR	<u>METALS AND CYANIDE</u>	PMPCRE	P-CRESOL*
	CYANIDE-FREE, UNFLT. REAC.		
	<u>BASE, NEUTRAL AND ACID EXTRACTABLE COMPOUNDS</u>		
P2AMET	AMETRYNE	PMPCRE	P-CRESOL*
P2ATRA	ATRAZINE*		
P4IAZ	DIAZINON*		
P4EPAR	PARATHION ETHYL*		
P4MALA	MALATHION*		
P4MPAR	METHYL PARATHION*		
PM24DP	2,4-DICHLOROPHENOL		
PM24DT	2,4-DINITROTOLUENE		
PM24MP	2,4-DIMETHYLPHENOL		
PM24NP	2,4-DINITROPHENOL*		
PM26DT	2,6-DINITROTOLUENE		
PM2NP	2-NITROPHENOL		
PM46DP	2-METHYL 4,6-DINITROPHENOL		
PM48PE	4-BROMOPHENYLPHENYLETHER		
PM4CPE	4-CHLOROPHENYLPHENYLETHER		
PM4NP	4-NITROPHENOL		
PMANAA	ALPHA-NAPHTHYLAMINE*		
PMB2EM	BIS(2-CHLOROETHOXY)METHANE		
PMB2JE	BIS(2-CHLOROISOPROPYL)ETHER		
PMB2NE	BIS(2-CHLOROMETHYL)ETHER		
PMBNAA	BETA-NAPHTHYLAMINE*		
PMDMP	DIMETHYL PHTHALATE		
PMOCRE	O-CRESOL		
PMPCMC	P-CHLORO-M-CRESOL		
PN2CNA	CHLORONAPHTHALENE*		
PNACNE	ACENAPHTHENE*		
PNBBFA	BENZOBIFLUORANTHENE		
PNDABA	DIBENZ(A,H)ANTHRACENE*		
PNGHIP	BENZOGUINOLIPERYLENE*		
PNDNP	IDENO(1,2,3-CD)PYRENE*		
PODICH	DICHLORAN*		
POTOC	TRI-O-CRESYL PHOSPHATE*		
X301O	2-CHLOROPHENOL		
X324S	2,4,5-TRICHLOROPHENOL*		
X3246	2,4,6-TRICHLOROPHENOL		
X3PCPH	PENTACHLOROPHENOL		
P94CDO P94CDF P96CDF P97CDF P98CDF	<u>DIOXINS AND FURANS</u>		
	TETRACHLORODIBENZODIOXIN*		
	TETRACHLORODIBENZOFURAN		
	HEXACHLORODIBENZOFURAN		
	HEPTACHLORODIBENZOFURAN		
	OCTACHLORODIBENZOFURAN		
PI2NDA PI2STRO PI2TOX	<u>PESTICIDES, HERBICIDES, PCRS</u>	POCAPN X1HCBD	CAPTAN* HEXACHLOROBUTADIENE*
	ELDRIN ALDENHYDE		
	STROBANE*		
	TOXAPHENE		
B10CTE B11VBR B2BDCI B2STYR E1DIEE PM2CEE X11122 X1112T X111CE X1112CE X1112CP X1113DP X1113DR X1ACTO X1BDCM X1BETH X1BROM X1CDCCE X1CHLE X1CHLM X1CTET X1DCFM X1DCLLE	<u>VOLATILES</u>	X1ACRO X1ACRY	ACROLEIN ACRYLONITRILE*
	1-OCTENE		
	VINYL BROMIDE*		
	BROMODICHLOROBENZENE		
	STYRENE		
	DIETHYL ETHER		
	2-CHLOROETHYL VINYLETHER*		
	1,1,2,2-TETRACHLOROETHANE*		
	1,1,2-TRICHLOROETHANE*		
	1,1-DICHLOROETHANE		
	1,2-DICHLOROETHANE		
	1,2-DICHLOROPROPANE		
	CIS-1,3-DICHLOROPROPENE		
	TRANS-1,3-DICHLOROPROPENE		
	ALPHA-CHLOROTOLUENE		
	BROMODICHLOROMETHANE		
	BROMOETHANE*		
	BROMOFORM*		
	CIS-1,2-DICHLOROETHYLENE		
	CHLOROETHANE*		
	CHLOROMETHANE*		
	CARBON TETRACHLORIDE		
	DICHLORODIFLUOROMETHANE		
	1,1-DICHLOROETHENE		

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

TABLE 5-13 a - GLOBAL SUMMARY OF CONTAMINANTS NOT DETECTED IN TREATED SLUDGES

CONFIRMED		NOT CONFIRMED	
CONTAMINANT CODE	CONTAMINANT NAME	CONTAMINANT CODE	CONTAMINANT NAME
X1T12D X1TCFM X1VCL X212CB X213CB X23CTO X2CBEN X2CPPE	<u>VOLATILES</u> TRI-1,2-DICHLOROETHYLENE* TRICHLOROFLUOROMETHANE* VINYL CHLORIDE* 1,2-DICHLOROBENZENE 1,3-DICHLOROBENZENE 3-CHLOROTOLLENE CHLOROBENZENE 3-CHLOROPROPENE*		

* - NOT DETECTED IN ANY STREAM TYPE AT ANY PLANT

Table 5-14
SUMMARY OF CONTAMINANTS DETECTED IN ANY SAMPLE TYPE

Sample Type	No. Plants	No. Samples Det. (1)	Metals and Cyanide			Base Neutral and Acid Extractable			Dioxin/Furan (2)			Pesticides and Herbicides			Volatiles		
			No.	Max %	Max %	No.	Max %	Max %	No.	Max %	Max %	No.	Max %	Max %	No.	Max %	Max %
			Det. (3)	Plant Prev.	Freq. Det.	Det. (3)	Plant Prev.	Freq. Det.	Det. (3)	Plant Prev.	Freq. Det.	Det. (3)	Plant Prev.	Freq. Det.	Det. (3)	Plant Prev.	Freq. Det.
Raw Sewage	37	275	15	100	99.7	31	86.5	60.7	3	10.8	7.4	29	100	77.5	22	37.8	15.7
Primary Effluent	7	39	14	100	100	6	71.4	46.2	2	26.8	25.0	16	85.7	72.5	12	85.7	55.3
Lagoon Effluent (4)	2	12	10	100	100	0	-	-	0	-	-	7	100	100	0	-	-
Secondary Effluent	28	224	15	100	100	24	14.3	3.1	4	14.3	9.1	24	100	78.0	19	64.3	16.5
Tertiary Effluent (4)	1	10	13	100	100	15	100	30.0	0	-	-	8	100	90.0	9	100	50.0
Raw Sludge (2)	34	51	16	100	100	15	85.3	82.3	7	58.5	50.0	27	79.4	78.4	10	41.2	31.4
Treated Sludge (2)	34	50	15	100	100	19	35.3	30.0	5	64.7	53.1	30	73.5	68.0	10	32.4	30.0

Notes:

(1) The number of samples may vary depending on compound group. The number given is representative of most compound groups.

Refer to Table 5-4(a) - 5-10(a) for exact numbers.

(2) Samples are 5-day composites.

(3) The number of contaminants detected in the sample type.

(4) Summary data for lagoon and tertiary effluents is included for the sake of completeness. Due to the small number of facilities sampled, this data should be interpreted with caution.

Only 5 base neutral and acid extractable compounds (M-cresol, Phenol, Phenanthrene, Butylbenzyl phthalate and Naphthalene) were detected at more than 20% of the plants studied for any sample type. With the exception of the 5 compounds, base neutral and acid extractable compounds were detected at a maximum of 14 percent of plants and in a maximum of 8 percent of samples, for all sample types. Interestingly, the maximum prevalence and frequency of detection of base/neutral acid extractable compounds in all secondary effluent samples was substantially smaller than in the raw sewage, primary effluent or sludge sample types.

The maximum frequency of detection and prevalence of dioxins and furans was relatively low in raw sewage and in primary and secondary effluent streams. In contrast, the number of dioxin/furan compounds detected, and the maximum frequency of detection and prevalence were markedly greater in sludge streams.

Approximately the same number of compounds in the pesticide/herbicide group were detected in raw sewage, secondary effluent and raw treated sludge streams. About 50 percent fewer compounds were detected in primary effluents. This may be attributable to a lower number of primary plants monitored. The maximum frequency of detection and prevalence is quite large and reasonably uniform for all sample types.

The largest number of volatile compounds were detected in the raw sewage and secondary effluent streams. Maximum frequency of detection and prevalence were quite variable among the sample types ranging from 32 percent to 85 percent prevalence and 15 percent to 55 percent frequency of detection.

This interim report on the WPCP Pilot Monitoring Study was prepared to present the study program methodology and the analytical data in conjunction with the QA/QC results. Also included are the individual plant information summaries (Appendix A).

The final report will present a more detailed review and analysis of the study results. More specifically, the report will include the following:

- o An assessment of the impact of industrial, residential and sanitary sewer inputs on the nature and loadings of HCs observed in the raw wastewater and sludges.
- o An estimate of HC loadings discharged in the sludges and liquid effluents of the 37 WPCPs.
- o An assessment of the ability of WPCPs to remove HCs and identification of the factors affecting HC removal efficiency.
- o A prioritized list of contaminants observed at the 37 WPCPs.
- o The major concerns affecting the implementation of the monitoring regulation and recommendations to address problem areas.

1. Ontario Ministry of the Environment (1988). "The Effluent Monitoring Priority Pollutants List (1987)", Ontario MOE, Hazardous Contaminants Coordination Branch, Toronto, Ontario. (ISBN 0-7729-0784-7).
2. U.S. EPA (1984). "Development Document for Effluent Limitations Guidelines and Standards for the Plastics Moulding and Forming Point Source Category". U.S. EPA, Office of Water Regulations and Standards, Industrial Technology Division, Washington, D.C. (EPA-440-/1-84-069).
3. Zenon Environmental Inc. "Joint MOE/Environment Canada/MEA Municipal Sewage Treatment Plants Pilot Monitoring Project Volume 1 - Final Report" January 1988. Prepared for Ontario Ministry of the Environment.
4. Survey of Industrial Discharges to 37 WPCPs (1987), Ministry of the Environment Internal Report.
5. Ontario Ministry of the Environment (1988). "Estimation of Analytical Detection Limits", MISA Report. Ontario MOE, Toronto, Canada. (ISBN 0-7729, 0784-7).
6. Ontario Ministry of the Environment, LCS-QA/QC Section, Laboratory Services Branch (1988). "QA/QC Report on the Spiked Effluent and Sewage Samples from the 40 Sewage Treatment Plants Toxic Survey Project".



